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GC-MS Analysis of Phyto-Chemical Constituents in Aqueous, Methanol and Ethyl Acetate Extracts of *Vitis Vinifera* Peel and Its Characterization Studies by Using AGNPs

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Abstract

Vitis Vinifera peel (Red) has major components such as Organic acids, Malate, higher moisture content and it is the source of medicine with high antioxidant activity for hundreds of years, *Vitis vinifera* belongs to family Vitaceae. But in our present work is designed to identify the possible phyto-chemicals compounds present in the Aqueous extracts, methanolic extract, and Ethyl acetate extracts of of *Vitis vinifera* peel by using GC-MS spectrum with their retention times indicating the peaks. This Vitis vinifera peel were shown nearly Thirty to Forty compounds from the Aqueous, Methanol and Ethyl acetate extracts the major chemical compounds were Xylose, Cyclo-hexanidiol, Octane, Butanoic acids and Octa-deca-diyndioic acids etc and from the Aqueous extracts we synthesized the AgNPs for preliminary characterization studies such as UV, DLS and SEM analysis.

Keywords

Vitis vinifera peel, AgNPs, Aqueous extract, Methanolic extract, Ethyl acetate extract, GC-MS spectrum.

1. INTRODUCTION:

Vitis vinifera is a member of the Vitaceae family, native to the Mediterranean region, central Europe and southwest Asia and cultivated today in all temperature regions of the world [1]. *Vitis vinifera* is used in conditions like hemorrhages, anemia, leprosy, skin diseases, syphilis, asthma, jaundice, bronchitis, anti-inflammatory, anti-carcinogenic, platelet aggregation inhibiting, and metal chelating

properties [2-4]. Red Grapes or *Vitis vinifera* is a Berry fruit and belongs to the group of versatile fruits which are used in a wide range as popular foods from raisins to jelly to wine. Over 72 million tons of grapes are grown every year and 7.2 trillion gallons of wine is produced. Grapes are rich source of many vital nutrients and antioxidants. [5] *V.vinifera* seed contains lipid, protein, carbohydrates and 5-8% polyphenols. The pulp majorly contains water (65%-



85% of grape weight), sugars (15%-25%), it directs the sweetness. It is attributed to different mix of acids such as malic acid, tartaric acid, succinic acid, ascorbic acid, citric acid, phenols, and flavonoids. [6] These are considered to have biological properties, not only limited to antioxidant, anti-inflammatory, anti-cancer, antimicrobial, antiviral, cardioprotective, neuroprotective, hepatoprotective activities but also as a nutraceutical [7].

GC-MS is a method that couples two different analytical techniques, gas-liquid chromatography, and mass spectrometry to identify different phytochemical compounds present in a test sample. GC can separate volatile as well as semi-volatile compounds with higher resolution, but it cannot identify them. MS can be used to obtain structural information of the compound, but it cannot separate readily. It is used to analyze complex biochemical and organic mixtures and it is also highly compatible. [8] In addition, the size of the peaks determined by the spectrum is a directly proportional to the amount of material present in the test sample.

Several studies have indicated that extracts obtained from grape seed inhibit enzyme systems that are responsible for the production of free radicals, and that they have anti-mutagenic and anti-carcinogenic. It has a protective effect on oxidant-induced production and deposition of extracellular matrix components [9].

Hence the objective of the present study was aimed to identify the possible Phyto-chemical compounds using GC-MS Aqueous, Methanol and Ethyl acetate extracts of *Vitis vinifera* peel with the aid of GC-MS Technique and the AgNPs were synthesized by using the aqueous extracts of peel for UV, DLS and SEM analysis.

2. MATERIALS AND METHODS:

2.1. Preparation of *Vitis vinifera* peel aqueous extract.

The cleaned and air dried *Vitis vinifera* peels was taken and it was grounded by mortar and pestle until it turns in to thin fine powder then the powder was stored in refrigerator at 4°C for further analysis. 10g of *Vitis vinifera* powder was taken and dissolved in 100ml of distilled water and heated at 80°C for 1h then the extract was filtered by using Whatman No. 1 filter paper and collected in plastic bottle and stored at 4±C for further characterization and experimentation [10-11].

2.2. Preparation of *Vitis vinifera* peel extract mediated silver (Ag) nanoparticles.

Silver nitrate (>99% pure) was purchased from Sigma-Aldrich, India. To prepare the AgNPs, a 90-mL aqueous solution of 1.0x10⁻³M silver nitrate was

mixed with a 10-mL of 5% aqueous solution of vitis vinifera peel extract. The vitis vinifera Ag solution was yellow in color and the solution was stirred repeatedly for an hour, and it was observed that the color of the solution has been changed to brown of which visually confirms the formation Nanoparticles. These vitis vinifera silver Nanoparticles were characterized by using the techniques such as UV-Vis spectrophotometry, Dynamic light scattering (Particle size), zeta potential and Scanning electron microscopy (SEM) analysis [12].

2.3. Vitis vinifera peel sample extractions

Twenty grams of the powdered peels were extracted with 100mL of 40% methanol overnight in a stopped bottle and with occasional stirring at room temperature (28°C). The sample was first sieved using muslin cloth and then filtered using Whatman No.1 filter paper. This process was repeated three times to remove the contaminants in sample. The filtrate was concentrated under rotary vacuum evaporator for one hour at 50°C and then lyophilized to get a compounds extract. The dry extract was preserved under 4°C until further use and the same procedure used for ethyl acetate compound extraction [13].

2.4. Gas chromatography-mass spectrometry (GC-MS) and identification of phytocompounds

GC-MS analysis was carried out on a GC-MS (Model: QP2010 PLUS Shimadzu, Japan) comprising a AOC-20i auto-sampler and chromatograph interfaced to a mass spectrometer (GC-MS). The instrument was equipped with a VF 5 ms fused silica capillary column of 30 m length, 0.25 mm diameter and 0.25µm film thickness. The temperatures employed were column oven temperature 80°C, Injection Temp 250°C at a pressure of 108.0 kPa, scan range 40-800 u and an injection volume of 1µL of the plant extract (split ratio 10:1). The total running time of GC-MS was 30 min. The relative percentage of the extract was expressed as percentage with peak area normalization. The fragmentation pattern spectra of the unknown components were compared with those of known components stored in the NISTV.3.2 library [14]. The compound bioactivity prediction was based on Dr. Dukes Phytochemical and Ethnobotanical Databases. The name, molecular weight, and structure of the components of the test materials were ascertained [15].

3. RESULTA AND DISCUSSION:

3.1. UV-visible spectrum of *Vitis vinifera* Peel aqueous extract mediated synthesized AgNPs

In this study, extract when interacting with the silver nitrate salt solution form a dark brown solution due



to the reduction of the silver ion to AgNPs followed by a colour change indicating the biotransformation of ionic silver to reduced silver and the subsequent formation of AgNPs in an aqueous medium. The colour change was monitored visually and the peak at 316nm in the UV-visible spectra indicated the presence of AgNPs which may be due to the excitation of surface plasmon resonance vibrations in AgNPs (Figure 1) [16].

3.2. Dynamic light scattering analysis of *Vitis vinifera* Peel aqueous extract mediated synthesized AgNPs

The hydrodynamic diameter (size) of the AgNPs was found to be 87.6nm (Figure 2a) and was measured as a function of scattering angle of the laser from the surface of the particle. Further, zeta potential of AgNPs was also measured and was recorded as 48.7mV (Figure 2b). The zeta potential clearly indicates stability of the prepared AgNPs [17].

3.3. Scanning electron microscopic analysis of *Vitis vinifera* Peel aqueous extract mediated synthesized AgNPs

Surface morphology of silver nanoparticles was studied from the SEM micrograph. It is evident that AgNPs were spherical, irregular in shape and some of them were poly dispersed. The measured average size of AgNPs was 30-50µm (Figure 3) [12, 18].

3.4. Identification of Phyto-compounds present in Aqueous, Methanolic and Ethyl acetate extractions from *Vitis vinifera* peel

3.4.1. GC-MS analysis of Aqueous extract of Vitis vinifera peel

GC-MS chromatogram of the aqueous extract of *Vitis vinifera* peel showed four peaks (Figure. 4) in chromatogram indicating the presence of four phyto-chemical constituents. On comparison of the mass spectra of the constituents with the NISTO8, WILEY8 and FAME libraries the four phyto-constituents were characterized and identified in (Figure 5)

The retention time present at **2.86** each showing the compound of amphetamine-3-methyl.

The retention time at **19.74** each showing the compounds of Xylose, 1-P-Toluidino-1-Deoxy-Beta-D-Idopyranose, 2-1, 2-Dihydroxyethyl-9-Beta-D-

RibofuranosylHypoxanthine,Methyl-Alpha-D-

Ribofuranoside,He xanoicacid,Hexanoicacid,6-Bromo,9-Bromononanoicacid,D-Allose,Allo-

Inositol, Ribonicacid. Gamma-Lactone, 1,2,5,6-Di-O-Isopropylidene-3-o-

Methanesulfonyl,glucofurnose,Beta-D-

Glucopyranose,1,6-Anhydro,5-Thio-D-

Glucopyranose,1,6-Anhydro-Beta-D-

Talopyranose, D-Galactose, 6-Deoxy, Lactose, Beta-

Glucopyranose,1,6-Anhydro,5-Cyano-

Desoxinojirimycin, D-Chiro-Inositol, 3-o-2-Amini-4-Carboxyminomethylamino-2, 3, 4, 6-TE.

The retention time at **26.05** showing the compounds of 1, 3-Cyclohexanediol, 1, 3-Cyclohexanediol,Cis-Ethylamine,1-Methyl-2-5-Methyl-1H-Pyrazol-3-yl-, 1, 3-Cyclohexanediol,Trans-1H-Imidazole-4-

Ethanamine,n, 5-Dimethyl-, 4H-1, 3, 4-Triazol-3-Amine, N-Dimethyl amino methylene-, Ethyl2-2-Chloroacetamido-3,3,3-Trifluor olactate,1H-Pyrazole,1-Methyl-4-Methylaminomethyl-,1H-

Pyrazole,1-Methyl-4-Methylaminomethyl-,1-Beta -D-Ribofuranosyl-s-Triazol(2,3-A-S-Triazin-5,7-

Dione, Ribavirin, Glucopyranuronamide, 1-4-Amino-2oxo-1(2H) -Pyrmidinyl-1-4-Dideoxy, 1H-Imidazole, 2-Ethyl-4, 5-Dihydro-, Histamie, N-Triflouroacetyl-2-Amino-, Cyclohexan e-3-5-diol, Cis-, 2-

Hydroxymethylcyclopentanol,1H,3H-

Furo(3,4Furan,Tetrahydro-,1H-Imidazole,2-Ethyl-4,5-Dihydro-4-Methyl.

The retention time at **26.58** showing the compounds 22, 23-Dibromostigmasterolacetate, 1-Hydroxy-1, 7-Dimethyl-4-Isopropyl-2, 7-Cyclodecadiene, 10-12-Pentacosadiynoicacid, Methyl, 10, 12-Pentacosadiyonate, 5, 8, 11, 14-eicosatetraenoicacid, Methyl ester, All-Z-, Pregn-4-Ene-3, 20-Dione, 17, 21-dihydroxy-Bis-o-Methyloxime.

3.4.2. GC-MS analysis of Ethyl acetate extract of *Vitis vinifera* peel

GC-MS chromatogram of the ethyl acetate extract of *Vitis vinifera* peel showed thirteen peaks (Figure 6) in chromatogram indicating the presence of thirteen phyto-chemical constituents. On comparison of the mass spectra of the constituents with the NISTO8, WILEY8 and FAME libraries the thirteen phyto-constituents were characterized and identified in (Figure 7).

The retention time present at **2.85** each showing the compounds of S (-)- Cathionone, N-Acetyl, Acetamide, 2, 2, 2-Triflouro-, Acetamide, 222, Trifluro-, 2, 3-Pyridinecarboxylic acid.

The retention time present at **19.27** each showing
the compounds of 1,3-Cyclohexanedioldiacetatecis-,
1,1,4-CyclohexanediolDiacetateTrans-,1,3-

Cyclohexanediol diacetate Trans-, L-Alanine N-Acetyl-3-Chloro-Methyl ester, 3-Cyclohexen-1-ol-Acetate, 4(H)-Pyridine N Acetyl-

The retention time present at **19.75** each showing the compounds of 2-Butyl-1-2-Oxaborolane, Ether Hexyl Pentyl, Dichloroacetic acid 4-Methyl Pentyl ester, Sulfurous acid Iso hexyl 2-Pentyl ester,6,8-Doixatetradecane, 5iIsoxazolecarboxylix acid 4,5-Dihydro-5-Methyl ester(R), Di(Tetrahydrofurfuryl) Adipate, Pyrrolidine 1,1-Methylenebis,2-Ethyl-1-Butanol Triflouroacetate, sulphurous acid Hexyl Pentyl ester, Succinic acid Hexadecyl



Tetrahydrofurfuryl 1-Butanol 4ester, Hexyloxy, Succinic acid Octadecyl Tetrahydrofurfuryl Isobutyrate, Succinic ester,2-Ethylbutyl acid Heptadecyl Tetrahydrofurfuryl ester, 1-Octacosanol 2,4,6,8 Tetramethyl-All-R,Sulfurous acid Isohexyl Pentyl ester, Sulfurous acid Nonyl 2-Propyl ester, Butanoic acid 2-Ethyl Butyl Ester, Dichloroacetic acid 6-Ethyl-3-Octyl ester.

The retention time present at **20.51** each showing the compounds of 2,5-Diamino-2-Methylpentanoic acid,4-cycloocten-q-Amine Ν Methyl,2-T Butylperoxy-2-Ethylbutan-1-ol **Butyrate** ester, Octane 1,1-Oxybis, Methoxyacetic acid octyl ester,Acetic acid Trichloro-Octyl ester,10acid octyl ester, butanoic Undecenoic acid octylester, 2-Bromoproponoicacidoctylester, 5-

Octadecenal, 4-Amino-6-Methyl-Piperidin-2-

one,Chloromethyl octylether,2,4(1H,3H)-Pyridinedione1-Beta-D-Ribofuranosyl,Sulfurous acid Decyl 2-Propyl ester, Hexyl Octyl ether,2-Propenoic acid Pentadecyl ester,1,2,3-cyclohexanetriol,2-Propenoic acid Tridecyl ester.

The retention time present at**21.23** each showingthe compounds of Octadecane2-Methyl,Hexadecane, Nonadecane2-Methyl,Eicosane2-Methyl,Tricosane2-Methyl,Tricosane2-Methyl,Tricosane2-Methyl,Docosane2,21-Dimethyl,Heptadecane2,6,10,14-

Tetramethyl, Hexadecane, 1-lodo-2-

Methylundecane,Penta decane,Nonadecane2-Methyl,Undecane2,10-Dimethyl,Heptadecane2-

Methyl, Hexadecane, Octadecane, 1-

Iodo, Tetradecane, Heptadecane, 4-

Methyl,Pentadecane 2-Methyl,Decane 2,6,8-Trimethyl.

The retention time present at **22.29** each showing the compounds of Tricyclo[4.4.0.0(3,8)Dec-9-En-4-Ol,2,4Decadien1-

OI(E,Z),9,10Dimethylenetricyclo[4.2.1.1(2,5)Decane, 1,2,4-Methenocyclobut[CD]Inden-3-Dione3A,4,7,7A-Tetrahydro-5-Methyl,4,7-Methanoisobenzofuran-1,3-Dione,3A,4,7,7A-Tetrahydro-5-Methyl,1-2-

Methylenecyclohexyl-3-Phenylpropan-1-

OL,1,3,5Dodecatriene,6-[(1E)-1,3-Butadienyl-1,4-

Cycloheptadiene,4,7-Methanoisobenzofuran-1,3-

Dione3A,4,7,7A-Tetrahydromethyl,3-[(1Z)-1,3-

Butadienyl]-1-4-Vinylcyclopentene,6-[(1Z-1,3-Butadienyl]-1,4-Cycloheptadiene,Cyclobutane,1-

(1,3-butadienyl-2-Vinyl,1,3,7,11-

Cyclotetradecatetraene,2,7-

Methanopthalene, 1, 2, 4A, 7, 8, 8A-

Hexahydro,Cyclobutane1,2-Bis(1,3Butadienyl),1,2-Bis(3-Cyclohexenyl)ethylene,1,4,4A,5,8,8A-Hexahydronaphthalene,1,2,4a,4B,7,8,8A,8B- Octahydrobiphenylene, Tricyclo [4.3.0.0(3,7)] Non-8-En-4-Ol.

The retention time present at **23.08** each showing the compounds of Methyl 5-2-Phenyl propionyl Hexanoate, 2-Methyl-1-Phenyl-2-Propen-1-Ol, Tricyclo [4.2.1.1(2, 5)DecA-3, 7-Dien-9-One 10-Hydroxy-10-Methyl-Stere.

The retention time present at **26.54** each showing the compound of 1H-4-Azacycloprop [CD]Indene octahydro-4-Methyl-.

The retention time present at **30.58** each showing the compounds of Tricyclo[4.4.0.0(2,7)Dec-3-Ene-3-Methanol,1-Methyl-8-1-Methyethyl,2,6-Dimethyl-4-Nitro-3-Phenyl-Cyclohexanone,2,5-

Octadecadiynoicacid, methyl, Ester, 10, 12-

Docasadiyndiocacid, Falcarinol, 10-Heneicosene 11-Phenyl, 4, 7-

 ${\it Octade cadiynoicacid} Methylester, Methyl 5, 7-$

Hexadecadiyonate,Estra-5-(10)-En-3-One-17-OLacetate,(4,4-Dimethyl-2,4,5,6-Tetrahydro-1H-Inden-2-YL)Aceticacid,Methyl5,7-

Hexadecadiynoate, Estra-5(10)-En-3-One-17-Olacetate, (4,4-Dimethyl-2,4,5,6-Tetrahydro-1H-Inden-2-YL) Aceticacid, Methyl-8,10-

Octadecadiynoate, Methyl-7,9-Octadecadiynoate,1-3,3-Dimethyl-1-YL-2,2-Dimethylcyclopropene-3-Carboxylic acid [19].

3.4.3. GC-MS analysis of Methanol extract of *Vitis vinifera* peel

GC-MS chromatogram of the methanol extract of *Vitis vinifera* peel showed sixteen peaks (Figure 8) in chromatogram indicating the presence of sixteen phyto-chemical constituents. On comparison of the mass spectra of the constituents with the NISTO8, WILEY8 and FAME libraries the sixteen phyto-constituents were characterized and identified in (Figure 9)

The retention time present at **9.58** each showing the compounds of 4-Methyl-1-3-Oxazine-2,6[3H]-Dione,Dec-9-En-6-Oxo-1-Ylamide,1,2,5-Oxadiazole-3-Carbohydroxymicacid,4-

Amino,4(1H)Pyrimidinone,6-Amino-2-Methyl-5nitroso-,Propanenitrile3-Amino-2-Methyl-5-Nitroso-,Prpanenitrile,3-Amino-2,3-

Dihydroxymino, Glutamine, N-Methyl-Oxime, 3(2)H-Isoxazolone, 4, 5-Dimethyl, 2, 4(1H, 3H)-

Pyrimidinededione,1-[3,5-Dihydroxy-6-

Hydroxymethyl],1,2,3-Triazole-4-Methanol,1-4-Aminofurazan-3-YL),3-Methyl-3-5-

(Cyanoethyl)Tetrahydro-4-

Thiopyranone,Cycloheptano[D]Imidazolidine,1,3-

Dihydroxy-2-Methyl-,1H-Imidazole,2,4-Dimethyl-

,Ronidazole,Propanal,2-Methyl-Oxime,L-

Guanindinosuccinimide, 5-Amino-3, 4-Dimethyl-Isoxazole, Spiro [Tetrahydrofuran-3, 5'-Hydantoin, 1, 3-



Oxazin-2-one, Tetrahydro-3-Methyl-5-6-TrimethyleneCis-,1H-1,2,3-Triazole-5-Methanol,1-4-Amino-1,2,5-Oxadiazole-3-YL)-The retention time present at **12.27** each consisting compounds of 4-Chloro-1-Butanol,4-Chloro-1-ButanolOxirane, Propyl-, Propylaldoxime, 2-Methyl-Anti, 3-Ethyl-3-Methyldiaziidine, 4-Hydroxybutylacrylate, 4-Hydroxybutylacrylate, 1, 2, 5-Triazole-2-oxide,1-sec-Butyl-3-Nitro-4-Amino-,1-Dimethylamino-2-Nitroethylene,L-Gala-L-Ido-Octose, 5-Amino-3, 4-Dimethyl-Isoxazole, Pentanoicacid, 3-Hydroxy-4-Methyl-Methylester, Cycloheptanol, 3, 8, 9-Trioxabicyclo[4.2.1]nonane,aceticacid,Pentylester,C yclohexanol2-Methyl-Trans-, Isothiourea, 2-2-Octylsulfonyl ethyl-4-amino-1-Hexanol. The retention time present at **14.47** each consisting compounds of Furazanamine,4-Azido-,3the Ethylamino-5-Hexene-2-OL. The retention time present at 17.49 each consisting the compounds of Cyclopropaneteteradecanoicacid,2-octyl-methyl 1,3,2,5-Diformal-L-Rhaminitol ester, acetate, Dodecanoicacid, 2, 3bis(Acetyloxy)Propylester,Lyxitol,1-0-nonyl,2-Butanyl, 3-1, 3-Dimethylbutoxy-3-Pentanol-3-Methyl, 3-Pentanol-3-Methyl-, Alpha-D-Glucopyranoside, Methyl3, 6-Anhydro, Pyrimidine-2,4,6(1H,3H,5H-Trione,1-Octadecyl-,3-Octanol,3,7-Dimethyl-,3-Octanol,3,6-Dimethyl-,3-octanol,3,7-Dimethyl-,2-Methyl-3-Decanol, Eicosanoicacid, 1, 2, 3, 4-Pentadecanetetrol,[2R-(2R,3s,4s)],1-Octanamine,N-Methyl-N-Nitroso-,4-Heptanol,2,6-dimethyl-4-Propyl,1,2,3,4-Hexadecanetetrol,2R-2R,3S,4S)],D-Mannohexadecane-1,2-3,4,5-Pentol,Undecanol-4. The retention time present at **18.42** each consisting the compounds of 2-Pentenoicacid, 4-oxo-Methylester(z),Pentanoicacid,3,5-dioxo-Methylester, Hexanoicacid, 3-Oxo-Methylester, 2-Pentanoicacid, 4-oxo-Methylester, (E), 2(5H)-Furanone, 5, 5-Dimethyl-, 1, 3-Dioxolane, 2, 2-Dimethyl-4-Hydroxymethyl-5-2-Hydroxypropyl,Xylopyranoside,Methyl4-Thio-Triacetate, Alpha-D-, 2(5H)-Furanone, 5, 5-Dimethyl,1,3-Dixolane,2,2-Dimethyl-4-Hydroxymethyl-5-2-Hydroxypropyl,Xylopyranosidemethyl-4-Thio-Triacetate, Alpha-D, 2(5H) Furanone, 5, 5-Dimethyl-,Heptanedioic acid 4-Methyl-Dimethyl ester, Acetic acid,2-Methyl-6-oxo-Heptyl ester,Hexanoicacid-5oxo-methyl ester, Heptanedioc acid 3-Methyl-Dimethylester, Methyl-4-oxo-2-Pentenoate, 5-Isoxazolidinecarboxylicacid, 5-Methyl-2-5-O-Methyl-2-3-O-1-Methy, 2-Propanol, 1, 3-Dimethylbutoxy, 3-

Ethyl-4-Methyl-3 Heptanol,2-Furanhexanoicacid, Tetrahydro-Beta, Delta, Dihydroxy-5-Metho, 5-Isoxazolidine, carboylicacid, 5, Methyl-2-5-O-2-3-O-1-Methylethyl,1,4-DI-O-Acetyl-2,5-DI-O-Methyl-3,6-Dideoxy-D-Glucitol. The retention time present at **18.27** each consisting the compounds of 5-Hexenoicacid Methylester,3-Dodecen-1-OL,2-Heptadecenal,4-OctadecenoicacidMethylester,Undecanenitrile,4chloro-3-N-Hexyltetrahydropyran,Hexanenitrile,5-Methyl-, Decanitrile, Methyl-3-Cyclopropylpropanoate, Nonanenitrile, E-2-Octadecen-1-OL, Undecanenitrile, Z-3-Octadecen-1-OL-Acetate, 5-Hexanoic, acid, methylester, 1, 19-Eicosadiene, z-2-Octadecen-1-OL, Acetate, Decanenitrile, Cyclohexanol-2-Methyl-AcetateCis,Oxalicacid Cyclohexyl methyl propyl ester,2-Dodecen-1-OL. The retention time present at **20.04** each consisting the compound of 2-Propenoicacid, 3-2-2-Dimethyl-1-3-Dioxolan-4-YL-Methyl ester(s)-The retention time present at **22.56** each consisting compounds Butane,1,1'the of Ethylidennebis(Oxy)Bis-2-Methyl,1,6-Heptadien-4-OL,9-Octadecen-12-ynoic acid Methyl ester,5-Dimethylamino-2-Methyl-4-Oxazolecarbonitrile, Disulphide Isopentyl methyl,8,11,14-Eicosatrienoic acid Methyl ester(Z,Z,Z)-, Butanoyl chloride, Methyl Pentyl disulfide,5-Methoxy-cyclooctene,Bicyclo[2.2.2]Oct-5-En-2 Y-Ldimethylamine,1,4-dioxane 2.6 -Dimethyl,2,6-Dimethylmorpholine-4-Carbothioic acid, 2-1-2-Pyrimidyl]Ethynide. The retention time present at 24.63 each consisting the compounds of Cyclohexane,2,4-Disopropyl-1,1-Dimethyl, Cyclohexane, 1, 5-Disopropyl-2, 3-Dimethyl-, Cyclohexane,1,1'-1 Methyl propylidene Bis, Oxalic acid Heptadecylic 1-Menthyl ester, Oxalic acid 1-Menthyl pentadactyl ester. The retention time present at **26.41** each consisting the compounds of 1-Hexanol,2-Hydroxymethyl, AminopropionamideN-Methyl-N-4-1-Pyrrolidinyl-2-Butynyl-N-T-But2-Octenal(E),3-Methyl-4-Phenylthio-2-Prop-2-enyl-2,5-Dihydrothiophene 1,1-Dioxi, Fromamide N-Methyl-N-4-1-Pyrrolidinyl-2-Butynyl. [20-21], both worked on GC-MS of grape seeds and their results demonstrated 33 and 16 constituents respectively against the outcome of this study where 7 peaks were observed. Also, [22] in aqueous skin extract reported 16 compounds. Out of the compounds posited by other researchers only palmitic and stearic acid were in affirmation with the result. The disparity in the number and nature of compounds found in this research could be due to



differences in plant parts, solvents of extraction and geographical location.

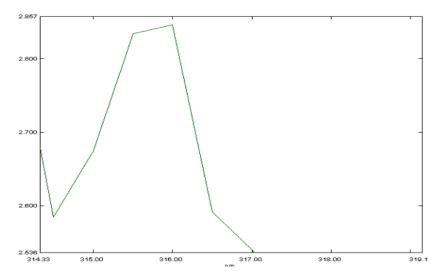


Figure.1 Showing UV-Visible spectrum analysis of *Vitis vinifera* peel extracted mediated synthesized Silver nanoparticles.

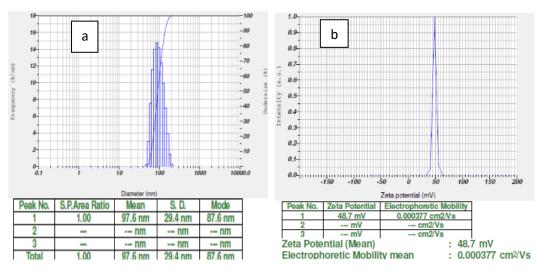
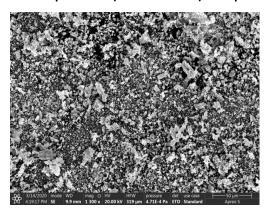


Figure.2 Showing Dynamic light scattering analysis of *Vitis vinifera* peel extracted mediated synthesized Silver nanoparticles a) Particle size b) Zeta potential.



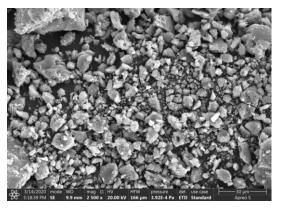
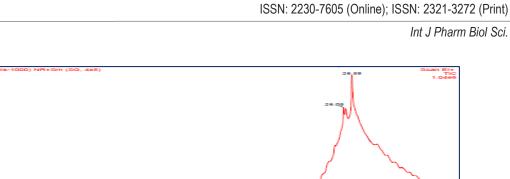


Figure.3 Showing Scanning electron microscopy analysis of *Vitis vinifera* peel extracted mediated synthesized Silver nanoparticles.



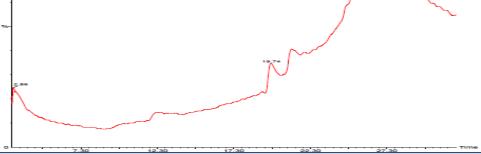
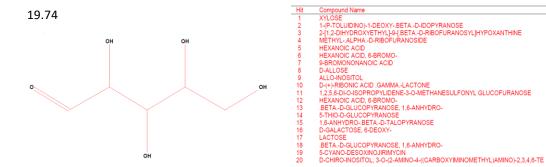
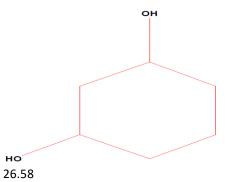


Figure.4 showing GC-MS Chromatogram of Aqueous extract of the Vitis vinifera peel





26.05



Hit		mpound Name	M.W.	Formula
1		3-CYCLOHEXANEDIOL	116	C6H12O2
2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17	Hit	Compound Name	M.W.	Formula
	1	22,23-DIBROMOSTIGMASTEROL ACETATE	612	C31H50O2Br2
	2	1-HYDROXY-1,7-DIMETHYL-4-ISOPROPYL-2,7-CYCLODECADIENE	222	C15H26O
	3	10-12-PENTACOSADIYNOIC ACID	374	C25H42O2
	4	METHYL 10,12-PENTACOSADIYNOATE	388	C26H44O2
	5	5,8,11,14-EICOSATETRAENOIC ACID, METHYL ESTER, (ALL-Z)-	318	C21H34O2
	6	PREGN-4-ENE-3,20-DIONE, 17,21-DIHYDROXY-, BIS(0-METHYLOXIME)	404	C23H36O4N2
		I,3H-FURO[3,4-C]FURAN, TETRAHYDRO- I-IMIDAZOLE, 2-ETHYL-4.5-DIHYDRO-4-METHYL-	114 112	C6H10O2 C6H12N2

C7H12O4N2 C14H25O9N

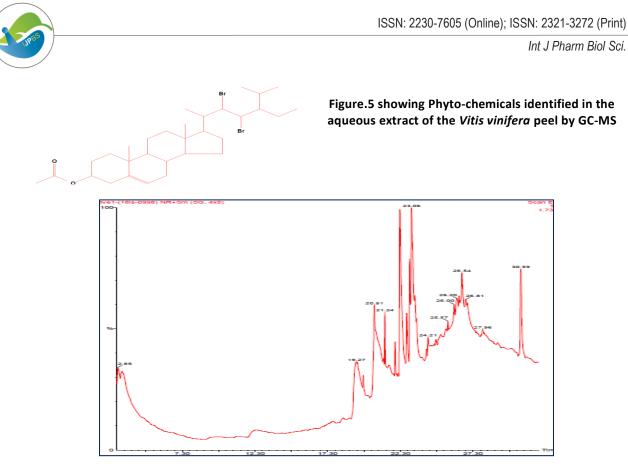
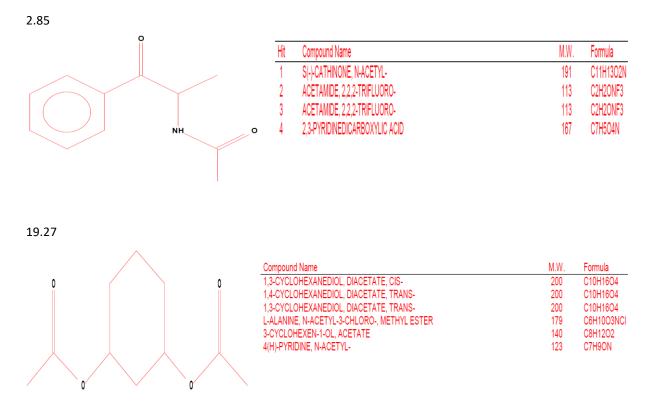


Figure.6 Showing GC-MS Chromatogram of Ethyl acetate extract of the Vitis vinifera peel



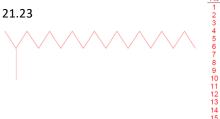


19.75

Int J Pharm Biol Sci.

HitCompound NameM.W.Formula12-BUTYL-1.2-OXABOROLANE126C7H150B2ETHER, HEXYL PENTYL172C11H2403DICHLOROACETIC ACID, ISOHEXYL 2-PENTYL ESTER212C8H1402CI24SULFUROUS ACID, ISOHEXYL 2-PENTYL ESTER202C12H260256,8-DOIXATETRADECANE202C12H260265-ISOXAZOLECARBOXYLIC ACID, 4.5-DIHYDRO-5-METHYL-, METHYL ESTER, (R)-143C6H903N7DI(TETRAHYDROFURFURYL)ADIPATE154C9H18N292-ETHYL-1-BUTANOL, TRIFLUOROACETATE198C8H1302F310SULFUROUS ACID, HEXADECYL TETRAHYDROFURFURYL ESTER226C11H2403S11SUCCINIC ACID, HEXADECYL TETRAHYDROFURFURYL ESTER426C25H4605121-BUTANOL, 4(HEXYLOXY)-174C10H202213SUCCINIC ACID, HEXADECYL TETRAHYDROFURFURYL ESTER440C26H4805142-ETHYLBUTYL ISOBUTYRATE172C10H200215SUCCINIC ACID, HEYALDEYL TETRAHYDROFURFURYL ESTER440C26H4805142-ETHYLBUTYL ISOBUTYRATE172C10H200215SUCCINIC ACID, IETRAHYDROFURFURYL ESTER440C26H4805161-OCTACOSANOL, 2,4,6,8-TETRAMETHYL-, (ALL-R)-466C32H66017SULFUROUS ACID, ISOHEXYL PENTYL ESTER236C11H2403S18SULFUROUS ACID, ISOHEXYL PENTYL ESTER236C11H2403S19BUTANOIC ACID, 2-ETHYL-, BUTYL ESTER260C12H2203S19BUTANOIC ACID, 6-ETHYL-3-OCTYL ESTER268

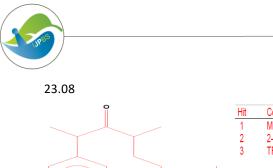
Compound Name 2:5-DIAMINO-2-METHYLPENTANOIC ACID 4-CYCLOOCTEN-1-AMINE, N-METHYL-2:T-BUTYLPERCXYL2-ETHYLBUTAN-1-OL, BUTYRATE ESTER OCTANE, 1,1'OXYBIS-METHOXYACETIC ACID, OCTYL ESTER 10-UNDECENOIC ACID, OCTYL ESTER BUTANOIC ACID, OCTYL ESTER 2-BROMPOPIONIC ACID, OCTYL ESTER 2-BROMPOPIONIC ACID, OCTYL ESTER 2-BROMPOPIONIC ACID, OCTYL ESTER 2-BROMPOPIONIC ACID, OCTYL ESTER 4-AMINO-6-METHYL-PIPERIDIN-2-ONE CHLOROMETHYL OCTYL ETHER 2.4(114,3H)-PYRIDINEDIONE, 1-.BETA--D-RIBOFURANOSYL-SULFUROUS ACID, DECYL 2-PROPPIL ESTER HEXYL OCTYL ETHER 2-PROPENOIC ACID, PENTADECYL ESTER 1.2.3-CYCLOHEXANETRIOL 2-PROPENOIC ACID, TRIDECYL ESTER 20.511 Formula C6H14O2N2 C9H17N C14H28O4 C16H34O C16H34O C16H34O C11H22O3 C10H17O2CI3 C19H36O2 Hit M.W 146 139 260 242 242 274 290 264 266 128 178 243 264 214 282 132 254 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 C19H36O2 2H24O2 1H21O2B ОН C18H34O C6H12ON2 C9H19OC C10H13O 4H30O 8H34O2 C6H12O3 C16H30O2



Hit	Compound Name	M.W.	Formula
1	OCTADECANE, 2-METHYL-	268	C19H40
2	HEXADECANE	226	C16H34
3	NONADECANE, 2-METHYL-	282	C20H42
4	EICOSANE, 2-METHYL-	296	C21H44
5	TRICOSANE, 2-METHYL-	338	C24H50
6	TRICOSANE, 2-METHYL-	338	C24H50
7	DOCOSANE, 2,21-DIMETHYL-	338	C24H50
8	HEPTADECANE, 2,6,10,14-TETRAMETHYL-	296	C21H44
9	HEXADECANE	226	C16H34
10	1-IODO-2-METHYLUNDECANE	296	C12H25
11	PENTADECANE	212	C15H32
12	NONADECANE, 2-METHYL-	282	C20H42
13	UNDECANE, 2,10-DIMETHYL-	184	C13H28
14	HEPTADECANE, 2-METHYL-	254	C18H38
15	HEXADECANE	226	C16H34
16	OCTADECANE, 1-IODO-	380	C18H37
17	TETRADECANÉ	198	C14H30
18	HEPTADECANE, 4-METHYL-	254	C18H38
19	PENTADECANE, 2-METHYL-	226	C16H34
20	DECANE, 2.6,8-TRIMETHYL-	184	C13H28

22.29

	Hit	Compound Name	M.W.	Formula
	1	TRICYCLO[4.4.0.0(3,8)]DEC-9-EN-4-OL	150	C10H14O
	2	2,4-DECADIEN-1-OL, (E,Z)-	154	C10H18O
	3	9,10-DIMETHYLENETRICYCLO[4.2.1.1(2,5)]DECANE	160	C12H16
	4	1,2,4-METHENOCYCLOBUT[CD]INDEN-3(1H)-ONE, OCTAHYDRO-	160	C11H12O
	5	BENZENE, [(CYCLOHEX-1-EN-3-YL)METHYL]-	172	C13H16
	6	4,7-METHANOISOBENZOFURAN-1,3-DIONE, 3A,4,7,7A-TETRAHYDRO-5-METHYL-	178	C10H10O3
	7	1-(2-METHYLENECYCLOHEXYL)-3-PHENYLPROPAN-1-OL	230	C16H22O
	8	1,3,5-DODECATRIENE	164	C12H20
\land	9	6-[(1E)-1,3-BUTADIENYL]-1,4-CYCLOHEPTADIENE	146	C11H14
	10	4,7-METHANOISOBENZOFURAN-1,3-DIONE, 3A,4,7,7A-TETRAHYDROMETHYL-	178	C10H10O3
	11	3-[(1Z)-1,3-BUTADIENYL]-4-VINYLCYCLOPENTENE	146	C11H14
Y Y	12	6-[(1Z)-1,3-BUTADIENYL]-1,4-CYCLOHEPTADIENE	146	C11H14
	13	CÝCLÓBUTANE, 1-(1,3-BUTADIENYL)-2-VINYL-	134	C10H14
	14	1.3.7.11-CYCLOTETRADECATETRAÉNE	188	C14H20
	15	CYCLOBUTANE, 1,2-BIS(1,3-BUTADIENYL)-	160	C12H16
	16	1,2-BIS(3-CYCLOHEXENYL)ETHYLENE	188	C14H20
ОН	17	2,7-METHANONAPHTHALÉNE, 1,2,4A,7,8,8A-HEXAHYDRO-	146	C11H14
	18	1,4,4A,5,8,8A-HEXAHYDRO-NAPHTHALENE	134	C10H14
	19	1,2,4A,4B,7,8,8A,8B-OCTAHYDROBIPHENYLENE	160	C12H16
	20	TRICYCL0[4.3.0.0(3,7)]NON-8-EN-4-OL	136	C9H12O



Compound Name M.W Formula METHYL 5-(2-PHENYLPROPIONYL)HEXANOATE C16H22O3 262 2-METHYL-1-PHENYL-2-PROPEN-1-OL 148 C10H12O TRICYCLO[4.2.1.1(2,5)]DECA-3,7-DIEN-9-ONE, 10-HYDROXY-10-METHYL-, STERE 176 C11H12O2 ő 26.54 Hit Compound Name M.W. Formula 1H-4-AZACYCLOPROP[CD]INDENE, OCTAHYDRO-4-METHYL-137 C9H15N

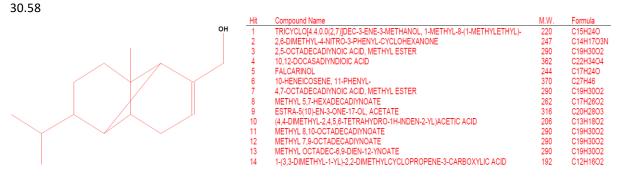
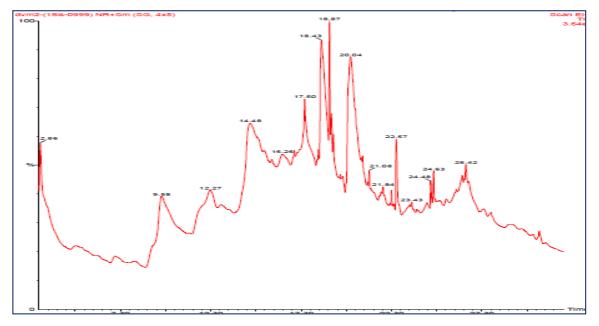
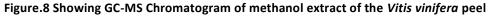
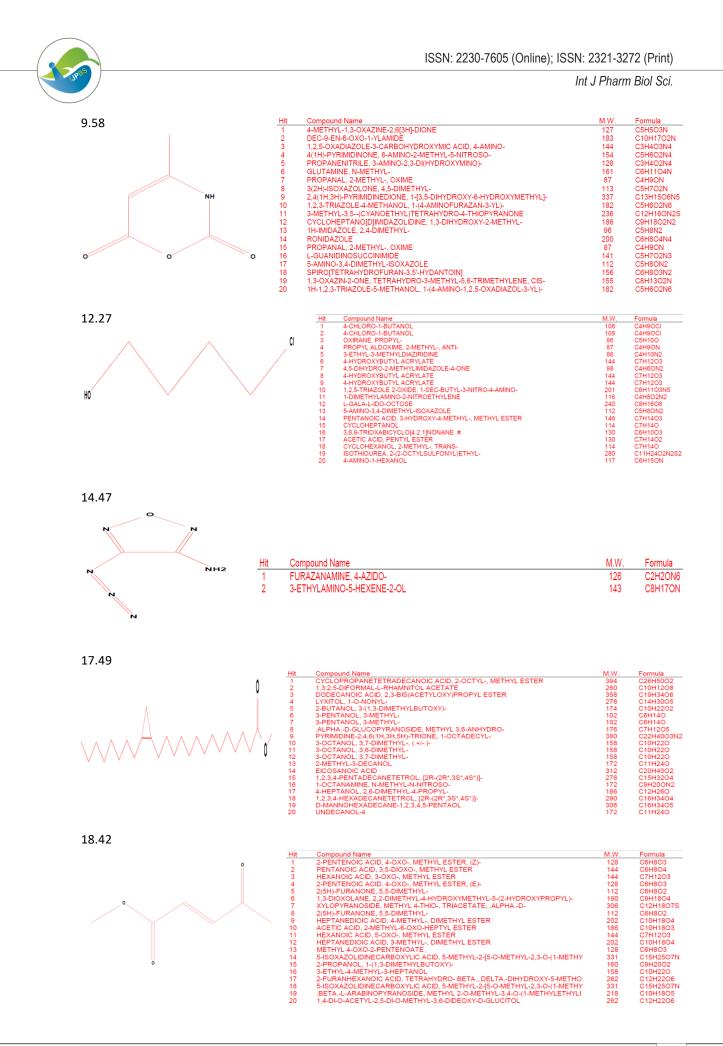


Figure.7 Showing Phyto-chemicals identified in the Ethyl acetate extract of the Vitis vinifera peel by GC-MS







D

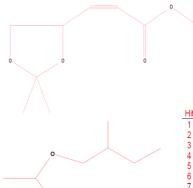


18.87

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Hit	Compound Name	M.W.	Formula
1	5-HEXENOIC ACID, METHYL ESTER	128	C7H12O2
2	3-DODECEN-1-OL	184	C12H24O
3	2-HEPTADECENAL	252	C17H32O
4	4-OCTADECENOIC ACID, METHYL ESTER	296	C19H36O2
5	UNDECANENITRILE	167	C11H21N
6	4-CHLORO-3-N-HEXYLTETRAHYDROPYRAN	204	C11H21OCI
7	HEXANENITRILE, 5-METHYL-	111	C7H13N
8	DECANENITRILE	153	C10H19N
9	METHYL 3-CYCLOPROPYLPROPANOATE	128	C7H12O2
10	NONANENITRILE	139	C9H17N
11	E-2-OCTADECADECEN-1-OL	268	C18H36O
12	UNDECANENITRILE	167	C11H21N
13	Z-3-OCTADECEN-1-OL ACETATE	310	C20H38O2
14	5-HEXENOIC ACID, METHYL ESTER	128	C7H12O2
15	1,19-EICOSADIENÉ	278	C20H38
16	Z-2-OCTADECEN-1-OL ACETATE	310	C20H38O2
17	DECANENITRILE	153	C10H19N
18	CYCLOHEXANOL, 2-METHYL-, ACETATE, CIS-	156	C9H16O2
19	OXALIC ACID, CYCLOHEXYLMETHYL PROPYL ESTER	228	C12H20O4
20	2-DODECEN-1-OL	184	C12H24O

20.04



Hit	Compound Name	M.W.	Formula
1	2-PROPENOIC ACID, 3-(2,2-DIMETHYL-1,3-DIOXOLAN-4-YL)-, METHYL ESTER, (S)-	186	C9H14O4

Hit	Compound Name	M.W.	Formula
1	BUTANE, 1,1'-[ETHYLIDENEBIS(OXY)]BIS[2-METHYL-	202	C12H26O2
2	1,6-HEPTADIEN-4-OL	112	C7H12O
3	9-OCTADECEN-12-YNOIC ACID, METHYL ESTER	292	C19H32O2
4	5-DIMETHYLAMINO-2-METHYL-4-OXAZOLECARBONITRILE	151	C7H9ON3
5	DISULFIDE, ISOPENTYL METHYL	150	C6H14S2
6	8,11,14-EICOSATRIENOIC ACID, METHYL ESTER, (Z,Z,Z)-	320	C21H36O2
7	BUTANOYL CHLORIDE	106	C4H7OCI
8	METHYL PENTYL DISULFIDE	150	C6H14S2
9	5-METHOXY-CYCLOOCTENE	140	C9H16O
10	BICYCLO[2.2.2]OCT-5-EN-2-YLDIMETHYLAMINE	151	C10H17N
11	1,4-DIOXÁNE, 2,6-DIMETHYL-	116	C6H12O2
12	2,6-DIMETHYLMORPHOLINE-4-CARBOTHIOIC ACID, 2-[1-[2-PYRIMIDYL]ETHYLIDE	293	C13H19ON5S

22.56

24.63



Compound Name	M.W.	Formula
CYCLOHEXANE, 2,4-DIISOPROPYL-1,1-DIMETHYL-	196	C14H28
CYCLOHEXANE, 1,5-DIISOPROPYL-2,3-DIMETHYL-	196	C14H28
CYCLOHEXANE, 1,1'-(1-METHYLPROPYLIDENE)BIS-	222	C16H30
OXALIC ACID, HEPTADECYL 1-MENTHYL ESTER	466	C29H54O4
OXALIC ACID, 1-MENTHYL PENTADECYL ESTER	438	C27H50O4
	CYCLOHEXANE, 2,4-DIISOPROPYL-1,1-DIMETHYL- CYCLOHEXANE, 1,5-DIISOPROPYL-2,3-DIMETHYL- CYCLOHEXANE, 1,1'-(1-METHYLPROPYLIDENE)BIS- OXALIC ACID, HEPTADECYL 1-MENTHYL ESTER	CYCLOHEXANE, 2,4-DIISOPROPYL-1,1-DIMETHYL-196CYCLOHEXANE, 1,5-DIISOPROPYL-2,3-DIMETHYL-196CYCLOHEXANE, 1,1'-(1-METHYLPROPYLIDENE)BIS-222OXALIC ACID, HEPTADECYL 1-MENTHYL ESTER466

26.41

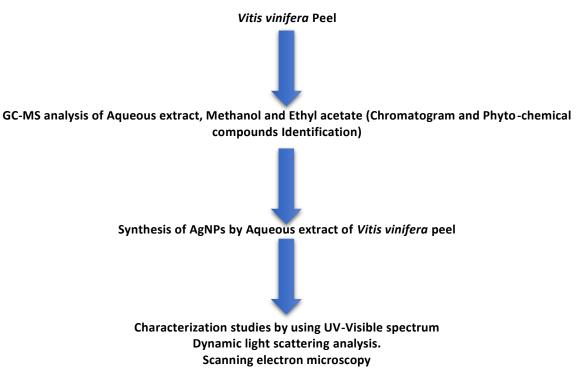


Hit	Compound Name	M.W.	Formula
1	1-HEXANOL, 2-(HYDROXYMETHYL)-	132	C7H16O2
2	AMINOPROPIONAMIDE, N-METHYL-N-[4-(1-PYRROLIDINYL)-2-BUTYNYL]-N'-T-BUT	323	C17H29O3N3
3	2-OCTENAL, (E)-	126	C8H14O
4	3-METHYL-4-(PHENYLTHIO)-2-PROP-2-ENYL-2,5-DIHYDROTHIOPHENE 1,1-DIOXI	280	C14H16O2S2
5	FORMAMIDE, N-METHYL-N-4-[1-(PYRROLIDINYL)-2-BUTYNYL]-	180	C10H16ON2

Figure.9 Showing Phyto-chemicals identified in the methanol extract of the Vitis vinifera peel by GC-MS

D





Flow chart of the Study

4. CONCLUSION:

In the present study more than twenty-five constituents have been identified from Aqueous, Ethyl acetate and methanolic extracts of *Vitis vinifera* peel by GC-MS analysis. The presence of various bioactive compounds justifies their use for various ailments by traditional practitioners Xylose and Cyclohexane were present. However, further studies are undertaken to identify the mechanisms of these compounds and their potential applications for the diseases in the field of Pharmaceuticals because the peel is having the medical applications (Phytochemical and bio-active compounds) for health benefits.

ACKNOWLEDGEMENT:

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REFERENCES:

- Gruenwald J., Brendler B.A. and Jaenicke C., PDR for herbalmedicines, 3 Rd ed., Thomson PDR: Montvale, NJ. (2004).
- [2] Anjaria J, Parabia M., Bhatt G., and Khamar R., Nature heals a glossary of selected indigenous medicinal plants of India. Sristi Innovations, Ahmedabad, India, (2002).
- [3] Sriram K., Benkovic SA., Hebert MA., Miller DB., O'Callaghan JP., Induction of gp130-related cytokines and activation of JAK2/STAT3 pathway in astrocytes precedes upregulation of glial fibrillary acidic protein

in the 1-methyl-4-phenyl-, 2, 3, 6-tetrahydropyridine model of neurodegeneration: Key signaling pathway for astrogliosis in vivo. J Biol Chem, 279:19936–9947, (2004)

- [4] Balu M., Sangeetha P., Haripriya D., Panneerselvam C., Rejuvenation of antioxidant system in central nervous system of aged rats by grape seed extract. Neurosci. Lett, 383: 295-300, (2005).
- [5] Aslanian A., Dizaji AA., Fahoomand P., Shahrary HA., Maheri N., Rouhnavaz S., Characterization of the nutritive value and protein fractions the Cornell net carbohydrateotien system in white and red grape (Vitis vinivera sp.) pomace. Research journal of biological sciences, 6(7): 298 – 303, (2011).
- [6] Cetin ES., Altinoz D., Tarcan E., Baydar NG., Chemical composition of grape canes. Industrial Crops and Products, 34(1): 994 – 998, (2011).
- [7] Vasil Georgiev., Anthony Ananga., Violeta Tsolova., Recent Advances and Uses of Grape Flavonoids as Nutraceuticals. Nutrients, 6: 391–415, (2014).
- [8] Syed Zameer Hussain., Khushnuma Maqbool., GCMS: Principle, Technique and its application in Food Science; Division of Post-Harvest Technology. Int J Curr Sci, 13: 116 – 126, (2014).
- [9] Dulundu E., Ozel Y., Topaloglu U., Grape seed extract reduces oxidative stress and fibrosis in experimental biliary obstruction. J. Gastroenterol Hepatol, 22:885-892, (2007).
- [10] Divya R., Supraja N., David E., Synthesis and Characterization of AgNPs from *Vitis Vinifera* Peel Extract and Its Antimicrobial Efficacy. Research and Development in material sciences, 9(3): ISSN: 2576-8840, (2019)

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- [11] Supraja N., Kishore B., Rajasekhar KK., Padmavathamma M., Synthesis of Carica papaya (Leaf, peel and seed) extracts mediated Ag nanoparticles for industrial and medical applications. Chem.sci.eng.res, 2(5): 29-39, (2020)
- [12] Divya R., Supraja N., David E., Synthesis and Characterization of ZnONPs from Vitis Vinifera Peel Extract and Its Antimicrobial Efficacy. Advancements in Bioequivalence and bioavailability, 2(2): ISSN: 2640-9275, (2019)
- [13] Naresh S., Sunil KS., Akki Suma, Ashika BD., Chitrali Laha Roy., Dr. Balasubramanian Sathyamurthy., GC-MS and FTIR analysis on the methanolic extract of red Vitis vinifera pulp. wjpls, 4(8): 153-159, (2018)
- [14] Anonymous, Duke's phytochemical and ethnobotanical database. http://www.ars-grin.gov/ cgi-bin/duke/ethnobot.pl. (2014)
- [15] Adewole E., Adewumi DF, Jonathan J., Fadaka AO., Phytochemical constituents and proximate analysis of orange peel (Citrus fruit). J. Adv. Bot. Zool., 1: 1-2, (2014).
- [16] Supraja N., Prasad TNVKV., Gandhi AD., Anbumani D., Kavitha P., Babujanarthanam R., Synthesis, Characterization and Evaluation of Antimicrobial Efficacy and Brine Shrimp Lethality Assay of Alstonia Scholaris Stem Bark Extract Mediated ZnONPs. Biochem. Biophys. Rep, 14: 69-77, (2018).
- [17] Supraja N., Prasad TNVKV., Soundariya M., Babujanarthanam R., Synthesis, characterization, and

dose dependent antimicrobial and anticancerous activity of phycogenic silver nanoparticles against human hepatic carcinoma (HepG2) cell line. AIMS Bioengineering, 3 (4): 425-440, (2016)

- [18] Supraja N., Avinash B., Prasad TNVKV. Nelumbo Nucifera Extracts Mediated Synthesis of Silver Nanoparticles for the Potential Applications in Medicine and Environmental Remediation. Adv. Nano Res, 5: 373 (2017).
- [19] Uraku Anayo Joseph., Uraku Oluchi Helen., Nweke Friday Nwalo., Orji Obasi Uche., Igwenyi Ikechukwu Okorie., Edwin Nzubechukwu., Ezeani Nkiru Nwamaka., Phytochemical and GC-MS Evaluation of Bioactive Principle of Vitis vinifera Peels. Asian Journal of Applied Sciences, ISSN 1996-3343, DOI: 10.3923/ajaps.2018.192.198, (2018)
- [20] Kadhim MJ., Al-Rubaye AB., Hameed IH., Determination of bioactive compounds of methanolic extract of Vitis vinifera using GC-MS. Int. J. Toxicol. Pharmacol. Res, 9: 113-126, (2017).
- [21] Kumar KA., Vijayalakshmi K., GC-MS analysis of phytochemical constituents in ethanolic extract of Punica granatum Peel and Vitis vinifera seeds. Int. J. Pharma Bio Sci, 2: B461-B468, (2011).
- [22] Bupesh G., Vijayakumar T., Manivannan S., Beerammal M., Manikadan E., Shanthi P., Vijaya AA., Identification of secondary metabolites, antimicrobial and antioxidant activity of grapefruit (Vitis vinifera) skin extract. Diabetes Obesity Int, 1(1): (2016).