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DETERMINING THE *IN-VITRO* CHOLESTEROL-REDUCING EFFICIENCY OF LACTOBACILLUS AND ENTEROCOCCUS STRAINS ISOLATEDFROM HUMAN BREAST MILK, FECES OF BREAST-FED INFANTS AND ANIMAL MILK (GOAT, COW AND BUFFALO)

Shaik Parvinnisa¹, Farhath Khanum^{*1} and Chandrasekhar.N²

¹Defence Food Research Laboratory- Defence Research and Development Organization (DFRL-DRDO), Mysore-570011

²Research and Development Center, Department of Chemistry, Shridevi Institute of Engineering and Technology, Sira Road, Tumakuru - 572106, Karnataka, India.

*Corresponding Author Email: <u>farhathkhanum@gmail.com</u>

ABSTRACT

The present study was designed to evaluate the cholesterol-removal efficacy in-vitro of four Lactobacillus and four Enterococcus strains. All 8 isolates exhibited Bile Salt Hydrolase (BSH) activity (1.64 to 3.09 mm of precipitation zone) and cholesterol assimilation with and without bile salt (7.7% to 95.58%) but significantly higher-cholesterol assimilation was observed with bile salts (9.157% to 95.58%), Among these NB 16 and NB12 isolates manifested significantly higher cholesterol assimilation superior to other 6isolates with deoxycholic acid (95.58% and 94.85%) and cholic acid (92.17% and 88.9%) than ox bile and TDCA, and also exhibited a high cholesterol reduction ability in natural [egg yolk (NB7-34.46% to NB16-73.82%) and skimmed milk (NB7-28.22% to NB16-72.88%)] cholesterol media than synthetic cholesterol media without bile salt and cholesterol reduction potential by the 8 isolates were optimized by different cultural conditions, among these NB16 displayed an elevated cholesterol removal ability with 83.51% at1%inoculum size, 83.9% at 24 h inoculums age., 83% at pH-7, 83.17% at 37°C incubation temperature, 83.27% at 24 h incubation time and 83.51% at 70 µg/ ml cholesterol concentration these probiotic strain could be exploited as a potential biotherapeutic agent to reduce cholesterol levels and the risk of cardiovascular diseases.

KEY WORDS

Bile salt hydrolase, Cholesterol- lowering effect, Enterococcus faecium, Lactobacillus para casei.

I. INTRODUCTION

Cholesterol is a vital structural component of the animal cell membrane [1] and its imbalance in the blood is called hypercholesterolemia which is a leading risk aspect for cardiovascular diseases and also the main cause of death [2]. It may be averted by practicing drug therapies but usually, they possess undesirable side effects certain as gastrointestinal discomfort [3]. Hence, necessity is there for more natural approaches among which dietary intervention using probiotics have potent health-promoting benefits namely bio therapeutic agents [4, 5]. Previous studies have stated that total and low-density lipoprotein (LDL-C) cholesterol decreased by probiotics in which it was strongly related with elevated CVD risks. Due to it has led to an improved interest in probiotics as it was much less expensive then should be considered a "natural health remedy" [6]. In the 1970s experimental reports have stated that wild

Lactobacillus strain have a cholesterol removal effect in human beings [7], after that many *in vitro* and *in vivo*



experiments have been performed to examine the cholesterol removal effect about lactic acid bacteria specifically in the strains of *Bifido bacterium* and *Lactobacillus* [8, 9,10]. The previous review on the cholesterol removal effect of probiotics concluded that the day by day consumption of products including lactic acid bacteria might also remain a dietary solution for long-term lowering effects of cardiovascular disease. [11, 12, 13].

Number of mechanisms for cholesterol reduction through probiotics have been recommended, such as like deconjugation about bile salts through bile-salt hydrolase (BSH) [14], Assimilation about cholesterol into cell membranes of the bacteria [15, 16], production about short-chain fat acids (SCFAs) for the duration of the growth about probiotics [17] then cholesterol transformation among coprostanol [18, 19] proposed as the BSH activity mechanism maximizes the amount of excretion. Such mechanism could be used in controlling serum cholesterol ranges by way of colonic microbes [20], due to this in past few decades the wide attention of the consumers, physicians and the researchers have attracted and used probiotic bacteria to decrease serum cholesterol levels [21]. It was once postulated that cholesterol assimilation was related with the availability of bile salts, it increased together with increasing concentration about bile salts within the medium [22] and numerous in vitro studies reported significantly higher cholesterol assimilation was observed in the existence of bile salt [23,24,25]. In accordance with the experimental findings [15, 26, 27, 28] cholesterol removal by probiotic appeared to not only strain specific but also growth dependent [15].

In the existent study of *Lactobacillus* and *Enterococcus* characterized as cholesterol-reducing probiotics by *in vitro* evidence.

II.MATERIALS AND METHODS

Bacterial strains: Strains were isolated from raw animal milk [Goat milk (*Enterococcus faecium NB44*), Cow milk (*Enterococcus faecalis NB7, Enterococcus faecium NB10*) and Buffalo milk (*Enterococcus faecium NB94*)], human milk (*Lactobacillus para casei NB12, Lactobacillus para casei NB14, Lactobacillus casei NB16*) and breastfeed Infant's fecal matter (*Lactobacillus para casei NB113*) by spread plate method [29,30] and maintained in MRS broth with 40 % sterile glycerol at – 20°C [31] and evaluated for probiotics potential by using

standard methods [32] and which exhibited acid tolerance (58-92%), bile tolerance (60-92%), synthetic gastric juice (60-90%), bacterial adhesion to hydrocarbons (93-96%), antimicrobial activity against food borne pathogens, antibiotic-resistant to 16 tested antibiotics out of 17, β -galactosidase activity upto1309±8mg/15 min, auto-aggregation (50-90%), fermented 17 sugars out 20, co- aggregation up to (62%), adhesion potential with 386 ± 15 bacterial cells (28.8%±1.5) per 100 cells of Caco-2 cell-line and identified as Lactobacillus para casei (NB12, 14 and 113), *Lactobacillus casei* (NB16), *Enterococcus faecium* (NB10, NB44 and NB94), and *Enterococcus faecalis* (NB7) by 16SrDNA sequencing method.

Chemicals: Cholesterol, Egg yolk, Skimmed milk, Bile salts, Calcium Chloride, De Man Rogosa and Sharpe broth/agar, were obtained from Hi-Media, Mumbai, India. Polyoxyethanyl cholesteryl sebacate, O-phthalaldehyde from Sigma, Saint Louis, USA. All the chemicals used were about an analytical grade.

Screening for bile salt hydrolase activity of *Lactobacillus* and *Enterococcus*.

Sterile filter discs placed on MRS agar plates with fresh culture, containing sodium salt of Tauro deoxycholic acid (0.5%) (w/v) and calcium chloride (0.037%) then the plates were incubated at 37°C for 72h an aerobically. Precipitation zone surrounding the filter disc around the colonies indicated the BSH activity of isolates, agar plates without supplementation were used as control [20, 33, 34].

In vitro cholesterol removal.

MRS broth with 4% commercial hen egg yolk, 12% Skimmed milk cholesterol, Water-soluble cholesterol (polyoxyethanyl cholesteryl sebacate) and Synthetic cholesterol (70 µg/ml) as a sources through filtersterilization with 0.45 mm filter and inoculated with an 18 h culture (1% v/v) then incubated for 24 h at 37 $^{\circ}$ C. Then the cells were eliminated from culture broth by centrifugation for 15 min at 4°C and 6000 rpm [6, 15, 35]. Cholesterol remaining in the spent broth was determined by using the O-phthalaldehyde technique. One thousand micro liter of the supernatant was added to 1000 μ l of potassium hydroxide (33% w/v) and 2000 μ l of ethanol (96%) was vortexed for 1 min, followed by heating for 15 min at 37 °C then 2000µl of the distilled water and 3000µl of the hexane were added after cooling, the tubes were then vortexed for 1 min then 1000µl of hexane layer was transferred to a glass tube



then evaporated on the water bath at 65°C. The residue was quickly dissolved into 2000 μ l of o-phthalaldehyde. After perfect mixing, 500 μ l concentrated sulphuric acid was added and the mixture was vortexed for 1 min. and allowed to stand for 10 min. Absorbance was read at

550 nm. [20, 34, 36, 37]. MRS broth including cholesterol without inoculation used as a control. All experiments have been done in triplicates. The proportion of cholesterol elimination was estimated the usage of the following formula:

= <u>Conc. of cholesterol in control – Conc. of cholesterol in test sample</u> X 100 Conc. of cholesterol in control.

In-vitro cholesterol removal with different bile-salts from media

MRS broth was mixed with 0.30% of different bile salts including ox gall, CA, DCA, and TDCA and water-soluble cholesterol (70 μ g/ml) through filter-sterilization (0.45 mm filter) and inoculated with (1% v/v) of an 18 h culture and incubated for 24 h at 37 °C [37,38,11], the cells were eliminated by centrifugation then the rest of the cholesterol concentration in the cell-free broth was estimated by using O-phthalaldehyde method [36].

In-vitro cholesterol removal by the dead, resting and growing cells with or without ox bile:

MRS broth (15 ml) was inoculated with cultures (18 h) and incubated at 37°C for 24 h. The cells were recollected by using centrifugation for 15 min at 4°C for 6000 rpm, then washed and re suspended in distilled water (15 ml). Then the cell suspension was separated into three portions. First one was heat treated for 15 min at 121°C, centrifuged and recollected cells suspension was inoculated in cholesterol media (70 μ g/ml) with and without ox bile (0.30%). The second one was centrifuged, and a recollected suspension was inoculated with 0.05 M phosphate buffer (pH 6.8) having cholesterol with and without ox bile. The third one was suspended in cholesterol media with and without ox bile and incubated at 37°C for 24 h. The cells were eliminated by centrifugation for 15 min at 4°C and 6000 rpm [15, 37, 39]. Then the spent stock was examined for cholesterol by the O-phthalaldehyde technique [36].

Optimization of cholesterol reduction by the isolates: Effects of culture conditions on cholesterol reduction by isolates were studied in MRS broth with different concentrations of cholesterol (50, 70, 100, 150 and 200 μ g/ml), incubation temperature (20, 30, 37 and 45°C), pH (3, 5, 6.5, 7 and 9), incubation time (12, 24, 48, 62 and 72 h), inoculum size (0.5, 1,1.5, 2, 3 and 4%) and inoculum age (8, 12, 18, 24,36 and 48 h)[40].

Statistical analysis

All assays were conducted in triplicates and the data were analyzed in Mean \pm SEM (n=3) and standard deviations, by using Microsoft Excel (Version 7.0).

II. RESULTS AND DISCUSSION

Qualitative determination of bile salts hydrolase activity of LAB isolates:

Bile salt hydrolase activity was regarded as detoxifying activity wherein the conjugated bile salt is converted to free bile by producing bile salt hydrolase enzyme which catalyzes deconjugation activity [25, 41]. In the present study, the BSH activity was observed in 8 isolates. Wherein, the conjugated bile acid (Na-TDCA) taurine was deconjugated and produced ample amount of deoxycholic acid by BSH enzyme and diffused into the surrounding in the form of precipitation zone around the active colonies at different levels (1.64 to 3.09 mm). Among these, Lactobacillus casei NB16 displayed highest BSH activity (3.09mm) and Enterococcus faecalis NB7 displayed lowest BSH activity (1.67mm) (Fig. 1) and (Table.1). Our results were correlated with previous studies [10, 11, 34, 42, 43]. Since detoxified bile acids have a greater possibility of getting excreted through the intestinal tract because they are much less soluble and are much less possibility to be absorbed through the intestinal lumen than conjugated bile salts, deconjugated bile. This will increase the requirement of cholesterol because of de novo synthesis of bile acids to substitute their loss through feces [44].

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Figure.1. BSH activity of Lactic acid bacteria on plate assay method

BSH activity of isolates manifested in the form of precipitation zone around the (disc contain) colonies at different levels (1.64 to 3.09 mm).

| Precipitation zone diameter (mm) ^a | | |
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Table.1. Screening of the bile salt hydrolase by plate assay method

a. Diameter of precipitation zone, b. Not detected, Values is expressed as mean ± SD in triplicates.

In vitro cholesterol removal from different Cholesterol media without bile salt:

In the present study, ability of 8 LAB isolates to reduce biodegrade cholesterol from 4 different sources was investigated. All LAB isolates show the cholesterol assimilation ability and exhibited different levels of cholesterol reduction ranging from 7.7% to 73.82%. Egg yolk cholesterol-lowering ability ranged from 31.93% (NB7) to 73.82% (NB16), skimmed milk (NB7-28.22% to NB16-72.88%), water-soluble cholesterol (NB7-9.157% to NB16-47.07%) and synthetic cholesterol (NB7-7.7% to NB16-42.42%). All LAB Isolates exhibited a high cholesterol reduction ability in natural (egg yolk and skimmed milk) cholesterol media than water soluble and synthetic cholesterol media without bile salt. Among these NB16 and NB12 strains showed excellent cholesterol-lowering characteristics superior to other 6 isolates (Fig. 2). Our findings correlate with previous studies [38, 45, 46]. Previous experimental findings reported that about 11 lactobacilli strains isolated from raw cow milk exhibited cholesterol degrading activity ranging from 42.88 to 97.20%, *Enterococcus hirae* up to 75.3% and the extent of cholesterol reduction has been varying between the strains i.e *Lactococcus lactis KF 147* with 66.8% and *L. lactis subsp. Lactis N7* with 97.0% of cholesterol reduction [6, 15, 26, 28, 47, 48, 49]. Several reports have proven that *in vitro* model LAB can remove cholesterol including bile salt [10, 11, 15, 38, 42, 43]. This contradictory result suggests that cholesterol elimination seems to be strain specific [15, 26, 27, 28].



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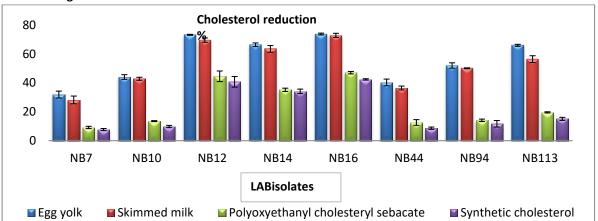


Figure.2. *In vitro* cholesterol removal from different Cholesterol media without bile salt

Egg yolk, Skimmed milk, Water-soluble (polyoxyethanyl cholesteryl sebacate) and Synthetic cholesterol removal ratio by Lactobacillus casei NB12, Values are expressed as mean ± SD in triplicates.

In-vitro cholesterol assimilation from media with different bile salts:

In vitro cholesterol assimilation potential of LAB isolates including bile salts in media has been shown by numerous studies [11, 37].

In the present study *in vitro* cholesterol removing ability of 8 LAB isolates with 4different bile salts (ox bile [conjugated-97% + deconjugated- 3%) and TDCA (conjugated) CA and DCA (deconjugated)] was assessed. All the 8 LAB isolates showed significantly higher cholesterol removal ability in the presence of bile salts (41.74% to 95.58%) than without bile salt. Cholesterol removal ability of 8 LAB isolates with poly oxyethanyl cholesteryl sebacate without bile salt ranged from 9.157% (NB7) to 47.07% (NB16), with TDCA the capacity increased to 41.74% (NB7) and the highest was 80.72% (NB16), with ox bile NB7 showed 52.67% cholesterol removal capacity and NB16 showed 83.27%, while in presence of cholic acid NB7 showed 56.38 % capacity and NB16 showed 92.17% and with deoxycholic acid (NB7-62.52% to NB16-95.58%). Among these NB16 and isolates manifested significantly **NB12** higher cholesterol assimilation superior to other 6 isolates with deoxycholic acid (95.58% and 94.85%) and cholic acid (92.17% and 88.9%) compared to the ox bile and TDCA (Fig.3). These results were in agreement with the previous reports [38, 50]. Which stated that higher cholesterol assimilation was observed in the presence of deconjugated bile acid than conjugated bile acids might be as a result of deconjugated bile acid high solubility, the detergent recreation of cholesterol then a high inhibitory effect of conjugated bile towards isolates may be one of the viable reasons [11, 24, 38].

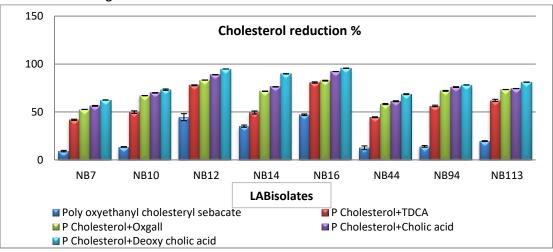


Figure.3. In-vitro cholesterol assimilation from media with different bile salts

In-vitro cholesterol assimilation/ cholesterol removal percentage (%) from media with and without Oxbile by 8 isolates, Values are expressed as mean ± SD in triplicates.



Cholesterol removal by the heat killed, resting and growing cells:

In vitro cholesterol reduction potential by the heat killed, resting and growing LAB strains from media containing ox bile has been shown by several studies [37, 38, 51].

In the present study, cholesterol removal ability of all 8 LAB isolates ranged from 0.1% to 83.16%. All 8 isolates growing on media containing ox bile showed significantly higher cholesterol reduction (NB7-50.77% toNB16- 83.16%) as compared to cells growing in the absence of ox bile (NB7-7.7% toNB16- 42.42%) and also as compared to the resting cells [(NB7-2% to NB16- 18.29%) and without ox bile(NB7-0.11% toNB16- 9.6%)] and heat-killed cells [(NB7-0.5% toNB16- 3.09%) and without ox bile (NB7-0.13% toNB16- 2.58%)]. This could be as a result of the transiting microbes deconjugation

activity about bile salts [52]. Among these NB 16 and isolates manifested significantly **NB12** higher cholesterol assimilation compared to other 6 isolates (Figure.4). However, there was no considerable difference in the level of cholesterol removal between resting cells and heat-killed cells without ox bile. This indicated that heat treatment did not reduce the cholesterol removal potential however slightly higher cholesterol reduction potential was observed in resting cells including ox bile and many reviews have confirmed that availability of bile salts has a positive effect due to their high cell permeability [24, 45, 46]. However, heatkilled and resting with and without bile salt showed a small rate of cholesterol removal, indicating that cholesterol may also be removed via binding on to the cell surface [38, 53]. These results were correlated with the previous studies [11, 15, 37, 38].

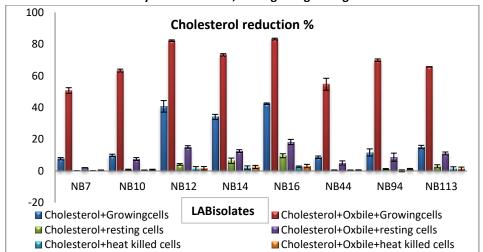


Figure.4. Cholesterol removal by the heat killed, resting and growing cells with and without ox bile

Cholesterol removal percentage (%) by the heat killed, resting and growing cells with and without ox bile by LAB isolates, Values are expressed as mean ± SD in triplicates.

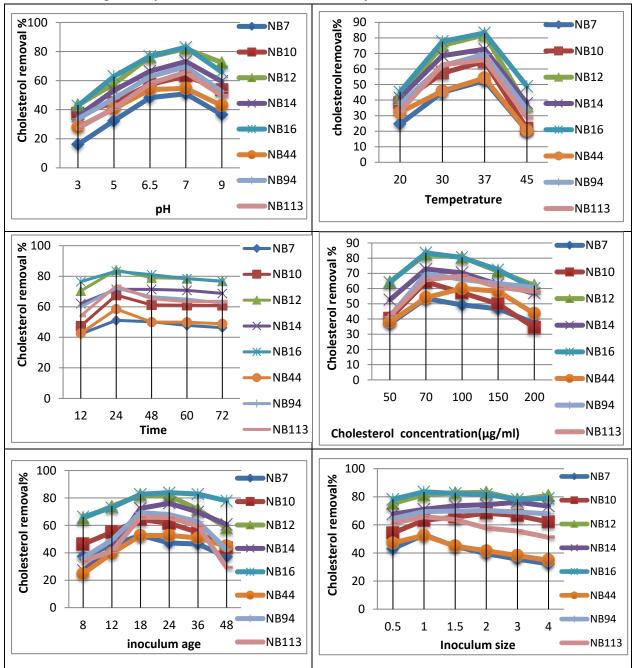
Optimization of cholesterol reduction by the selected bacterial isolates:

The effect of pH (3-9) on cholesterol reduction ability by 8 LAB isolates was assessed. In all isolates it increased gradually with decreasing acidity and reached to maximum values (NB7- 50.94% to NB16-83%) and decreased gradually (NB7-36.61% to NB16-65.95%) with increasing alkalinity of culture medium, maximum cholesterol reduction by the 8 isolates was observed at pH 7(neutral) (Figure 5.a). These results are in agreement with the previous reports [40]. The previous experimental finding reported that for *L. casei LA-1*, *Streptomyces fradiae* show high cholesterol reduction ability at pH-7.2 and for *Rhodococcus erythropolis ATCC* 25544 at pH-6.75 [54, 55]. The production of ideal enzymes is significantly altered by pH, via metabolic process therefore, the pH of the culture medium is of utmost importance for their structure and function [56]. The effect of temperatures (20 to 45°C) on cholesterol reduction ability by the 8 isolates was observed. It increased by increasing incubation temperature and reached to maximum values (NB7-52.47% to NB16-83.17%) at 37°C and then decreased when the temperature was increased (NB7-20.22% to NB16-48.92%) to 45°C (Figure.5. b.). The results are in accordance with the experimental findings reported earlier [40]. Previous experimental finding reported high cholesterol reduction ability for *E. hirae* shows at



37°C, *R. erythropolis ATCC 25544* at 29°C and *Bacillus subtilis SFF34* at 30°C [28,57,58,]. The optimal pH for decomposition of cholesterol is known to depend on the

growth of the microorganisms which influences by environmental and nutritional factors [40].





a). pH (3-9), b). Temperatures (20 to 45°C), c). Time (12-72h), d). Cholesterol concentrations (50- 200 μg/ml), e). Inoculums age (8-48h), f). inoculum size (0.5-4%), values are expressed as mean ± SD in triplicates

The effect of incubation time (12-72h) on cholesterol removal ability by the 8 isolates was observed. The maximum value was observed at 24 h of incubation ranging from (NB7-51.1% to NB12-84.06% and NB16-83.27). After that, cholesterol removal ability of 8

isolates decreases gradually and finally reached stability at 48 h with slight decreasing value (NB7-50.18% to NB12-79.01% and NB16-81%) (Figure 5c). Our results correlate with the previous reports [40], the previous experimental finding reported high cholesterol



reduction ability of L. *casei LA-1, L. lactis KF147, Enterococcus hirae* at 24 h and Lacto *bacillus acidophilus P10*6 at 20 h. [28, 49, 54, 59].

The effect of cholesterol concentrations from (50 to 200 μ g/ml) on cholesterol removal ability was observed. The cholesterol removal ability of all the 8 isolates increased gradually with increasing cholesterol concentration upto70 and 100 μ g/ml, in that 6 isolates (NB7-53.47%, NB10-64.48%, NB12-82.51%, NB14-72.89%, NB16-83.51% % and NB94-69.69%), showed maximum value at 70 µg/ml and remaining 2 isolates(NB44-60.15% and NB113-68.2%) reached the maximum value of at 100 µg/ml cholesterol, then cholesterol removal ability reduced with improved cholesterol concentrations (Figure 5.d). Our findings are in agreement with previous findings [40] which state that Lactobacillus acidophilus P106 removed 90% at 70 µg/ml, L. lactis KF147 removed 66.8%, Enterococcus strains removed (41.29%-56.61%), lactic acid bacteria and bifidobacteria removed (47%) at 100 µg/ml cholesterol [11, 49, 59, 60].

Excessive cholesterol concentration reduced water activity then the onset of plasmolysis may

suppress bacterial cholesterol assimilation ability [61]. The effect of age of the inoculate (8-48h) on cholesterol removal potential was studied. The maximum reduction potential of 5 isolates out of 8 isolates (NB7-52.8%, NB10-64.31%, NB12-82.18%, NB94-69.35% and NB113-65.93%) was observed at 18 h, remaining 3 isolates (NB14-76.11%, NB16-83.9% and NB44-52.67%) showed maximum reduction at 24 h inoculum age (Figure 5.e). High cholesterol assimilation was executed during the log phase and high biomass production executed when the cultures reached the stationary phase when the medium nutrients were depleted then enriched with inhibitory products leading to reduce in bacterial biomass and rate of cholesterol assimilation [40, 54].

The effect of inoculum size (0.5-4%) on the cholesterol removal ability was studied. Out of 8 isolates,4 isolates (NB7-52.97%, NB16-83.51%, NB44-52.52% yet NB113-65.81%) showed high cholesterol reduction capacity at 1% inoculum size and 3 isolates (NB10-68.34%, NB12-83.19% and NB94-70.26%) at 2% inoculum and NB14-76.11% at 3% inoculum size. NB16 showed high cholesterol reduction with 83.51% at 1% inoculum (Figure 5f). The high cholesterol assimilation was recorded with 1 to 2% inoculums size. Our results correlate with the previous findings [40, 54]. However,

some experimental studies reported maximum cholesterol assimilation for *Lactobacillus fermentum* and *Lacto bacillus acido philus P106* at 1% inoculum size [11, 59].

IV.CONCLUSION

All lactobacillus and enterococcus strains demonstrated high bile salt hydrolase activity and cholesterol removal with and without bile salt in different cholesterol media. these, Lactobacillus casei NB16 Among and Lactobacillus para casei NB12 isolates executed significantly higher cholesterol assimilation with deoxycholic acid (95.58% and 94.85%). These results also indicate that milk is an excellent resource for cholesterol-reducing lactic acid bacteria. However, in vivo investigations are still required to confirm the hypo cholesterolemic effects and only after that one can recommend for the development of new functional food preparations for public health especially where cholesterol reduction in food is the main target.

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VI. REFERENCES

- Boone LR, Lagor WR, de la Llera Moya M, Niesen MI, Rothblat GH, Ness GC. Thyroid hormone enhances the ability of serum to accept cellular cholesterol via the ABCA1 transporter. Atherosclerosis. 2011 Sep 1: 218(1):77-82.
- Ai M, Otokozawa S, Asztalos BF, Ito Y, Nakajima K, White CC, Cupples LA, Wilson PW, Schaefer EJ. Small dense LDL cholesterol and coronary heart disease: results from the Framingham Offspring Study. Clinical chemistry. 2010 Jun 1:56 (6):967-76.
- Davidson MH, Dillon MA, Gordon B, Jones P, Samuels J, Weiss S, Isaacsohn J, Toth P, Burke SK. Colesevelam hydrochloride (cholestagel): a new, potent bile acid sequestrant associated with a low incidence of gastrointestinal side effects. Archives of Internal Medicine. 1999 Sep 13;159(16):1893-900.
- Park YH, Kim JG, Shin YW, Kim SH, Whang KY. Effect of dietary inclusion of Lactobacillus acidophilus ATCC 43121 on cholesterol metabolism in rats. Journal of Microbiology and Biotechnology. 2007 Apr.17(4):655-62.



- Begley M, Hill C, Gahan CG. Bile salt hydrolase activity in probiotics. Applied and environmental microbiology. 2006 Mar 1;72(3):1729-38.
- Liu H, Yang C, Jing Y, Li Z, Zhong W, Li G. Ability of lactic acid bacteria isolated from mink to remove cholesterol: in vitro and in vivo studies. Canadian journal of microbiology. 2013 Jun 20:59(8):563-9.
- Mann GV, Spoerry A. Studies of a surfactant and cholesteremia in the Maasai. The American Journal of Clinical Nutrition. 1974 May 1;27(5):464-9.
- Pan DD, Zeng XQ, Yan YT. Characterisation of Lactobacillus fermentum SM-7 isolated from koumiss, a potential probiotic bacterium with cholesterol-lowering effects. Journal of the Science of Food and Agriculture. 2011 Feb. 91(3):512-8.
- Oner O, Aslim B, Aydaş SB. Mechanisms of cholesterollowering effects of lactobacilli and bifidobacteria strains as potential probiotics with their bsh gene analysis. Journal of molecular microbiology and biotechnology. 2014:24(1):12-8.
- Wang J, Zhang H, Chen X, Chen Y, Bao Q. Selection of potential probiotic lactobacilli for cholesterol-lowering properties and their effect on cholesterol metabolism in rats fed a high-lipid diet. Journal of dairy science. 2012 Apr. 95(4):1645-54.
- Pereira DI, McCartney AL, Gibson GR. An in vitro study of the probiotic potential of a bile-salt-hydrolyzing Lactobacillus fermentum strain, and determination of its cholesterol-lowering properties. Applied and environmental microbiology. 2003 Aug 1;69(8):4743-52.
- Anderson JW, Gilliland SE. Effect of fermented milk (yogurt) containing Lactobacillus acidophilus L1 on serum cholesterol in hypercholesterolemic humans. Journal of the American College of Nutrition. 1999 Feb 1;18(1):43-50.
- Noh DO, Kim SH, Gilliland SE. Incorporation of cholesterol into the cellular membrane of Lactobacillus acidophilus ATCC 431211. Journal of Dairy Science. 1997 Dec 1;80(12):3107-13.
- 14. Ahn YT, Kim GB, Lim KS, Baek YJ, Kim HU. Deconjugation of bile salts by Lactobacillus acidophilus isolates. International Dairy Journal. 2003 Jan 1;13(4):303-11.
- Kimoto H, Ohmomo S, Okamoto T. Cholesterol removal from media by lactococci. Journal of dairy science. 2002 Dec 1;85(12):3182-8.
- 16. Tabuchi M. Hypocholesterolemic effects of viable and heat-sterilized cells of Lactobacillus GG in rats fed a highcholesterol diet. Milchwissenschaft. 2004:59(5):249-53.
- 17. Trautwein EA, Rieckhoff D, Erbersdobler HF. Dietary inulin lowers plasma cholesterol and triacylglycerol and alters biliary bile acid profile in hamsters. The Journal of nutrition. 1998 Nov 1;128(11):1937-43.
- Lye HS, Rahmat-Ali GR, Liong MT. Mechanisms of cholesterol removal by lactobacilli under conditions that

mimic the human gastrointestinal tract. International Dairy Journal. 2010 Mar 1;20(3):169-75.

- Sanders ME, Huis J. Bringing a probiotic-containing functional food to the market: microbiological, product, regulatory and labeling issues. InLactic Acid Bacteria: Genetics, Metabolism and Applications 1999 (pp. 293-315). Springer, Dordrecht.
- 20. Tsai CC, Lin PP, Hsieh YM, Zhang ZY, Wu HC, Huang CC. Cholesterol-lowering potentials of lactic acid bacteria based on bile-salt hydrolase activity and effect of potent strains on cholesterol metabolism in vitro and in vivo. The Scientific World Journal. 2014.
- Baroutkoub A, Mehdi RZ, Beglarian R, Hassan J, Zahra S, Mohammad MS. Effects of probiotic yoghurt consumption on the serum cholesterol levels in hypercholestromic cases in Shiraz, Southern Iran. Scientific Research and Essays. 2010 Aug 18;5(16):2206-9.
- 22. Cohn JS, Kamili A, Wat E, Chung RW, Tandy S. Dietary phospholipids and intestinal cholesterol absorption. Nutrients. 2010 Feb 8;2(2):116-27.
- Ziarno M, Sękul E, Lafraya AA. Cholesterol assimilation by commercial yoghurt starter cultures. Acta Scientiarum Polonorum Technologia Alimentaria. 2007 Mar 30;6(1):83-94.
- 24. Tok E, Aslim B. Cholesterol removal by some lactic acid bacteria that can be used as probiotic. Microbiology and immunology. 2010 May;54(5):257-64.
- Gilliland SE, Nelson CR, Maxwell C. Assimilation of cholesterol by Lactobacillus acidophilus. Applied and environmental microbiology. 1985 Feb 1;49(2):377-81.
- Sieladie DV, Zambou NF, Kaktcham PM, Cresci A, Fonteh F. Probiotic properties of lactobacilli strains isolated from raw cow milk in the western highlands of Cameroon. Innovative Romanian Food Biotechnology. 2011 Sep 1; 9:12. -28.
- Huang Y, Wang X, Wang J, Wu F, Sui Y, Yang L, Wang Z. Lactobacillus plantarum strains as potential probiotic cultures with cholesterol-lowering activity. Journal of Dairy Science. 2013 May 1;96(5):2746-53.
- Yehia HM, Hassanein WA, Ibraheim SM. Purification and characterisation of the extracellular cholesterol oxidase enzyme from Enterococcus hirae. BMC microbiology. 2015 Dec;15(1):178.
- Sharpe ME, Fryer TF, Smith DG. Identification of the lactic acid bacteria. Identification methods for microbiologists. 1979; 2:233-59.
- Harrigan WF, McCance ME. Laboratory methods in food and dairy microbiology. Academic Press Inc.(London) Ltd.; 1976.
- Shafakatullah N, Chandra M. Screening of Raw Buffalo's Milk from Karnataka for Potential Probiotic Strains.Research Journal of Recent Sciences.2014Sep 3:9,73-78.

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- Garrity GM, Bell JA, Lilburn TG. Taxonomic outline of the prokaryotes. Bergey's manual of systematic bacteriology. Springer, New York, Berlin, Heidelberg. 2004 May.
- Toit MD, Dicks LM, Holzapfel WH. Identification of heterofermentative lactobacilli isolated from pig faeces by numerical analysis of total soluble cell protein patterns and RAPD-PCR. Letters in applied microbiology. 2003 Jul;37(1):12-6.
- 34. Shehata MG, El Sohaimy SA, El-Sahn MA, Youssef MM. Screening of isolated potential probiotic lactic acid bacteria for cholesterol lowering property and bile salt hydrolase activity. Annals of Agricultural Sciences. 2016 Jun 1;61(1):65-75.
- Ju H, Liu SL, Ao L, WANG Y. Study on characteristics of growth, screening and identification of cholesterolreducing Lactic acid bacteria. China Dairy Industry. 2007;35(8):7-10.
- Rudel LL, Morris MD. Determination of cholesterol using o-phthalaldehyde. Journal of Lipid Research. 1973 May 1;14(3):364-6.
- Anila K, Kunzes A, Bhalla TC. In vitro cholesterol assimilation and functional enzymatic activities of putative probiotic Lactobacillus sp. isolated from fermented foods/beverages of North West India. J Nutr Food Sci. 2016:6(2).
- Liong MT, Shah NP. Optimization of cholesterol removal by probiotics in the presence of prebiotics by using a response surface method. Applied and environmental microbiology. 2005 Apr 1;71(4):1745-53.
- Searcy RL, Bergquist LM. A new color reaction for the quantitation of serum cholesterol. Clinica chimica acta. 1960 Mar 1;5(2):192-9.
- Aboseidah AA, Rasmey AH, Osman MM, Desouky SG, Kamal N. Cholesterol reduction in vitro by novel probiotic lactic acid bacterial strains of Enterococcus isolated from healthy infants' stool. African Journal of Microbiology Research. 2017 Oct 14;11(38):1434-44.
- Ridlon JM, Kang DJ, Hylemon PB. Bile salt biotransformations by human intestinal bacteria. Journal of lipid research. 2006 Feb 1;47(2):241-59.
- Lavanya B, Sowmiya S, Balaji S, Muthuvelan B. Screening and characterization of lactic acid bacteria from fermented milk. British Journal of Dairy Sciences. 2011 Apr 1;2(1):5-10.
- Miremadi F, Ayyash M, Sherkat F, Stojanovska L. Cholesterol reduction mechanisms and fatty acid composition of cellular membranes of probiotic Lactobacilli and Bifidobacteria. Journal of Functional Foods. 2014 Jul 1;9:295-305.
- Kumar M, Nagpal R, Kumar R, Hemalatha R, Verma V, Kumar A, Chakraborty C, Singh B, Marotta F, Jain S, Yadav H. Cholesterol-lowering probiotics as potential biotherapeutics for metabolic diseases. Experimental diabetes research. 2012 May 3;2012.

- Lye HS, Rusul G, Liong MT. Removal of cholesterol by lactobacilli via incorporation and conversion to coprostanol. Journal of dairy science. 2010 Apr 1;93(4):1383-92.
- Philipp B. Bacterial degradation of bile salts. Applied microbiology and biotechnology. 2011 Feb 1;89(4):903-15.
- Kulkarni NS, Lokhande AP, Pachori RR, Agrawal PN, Dalal JM. Screening of the cholesterol degrading bacteria from cow's milk. Curr Res Microbiol Biotechnol. 2013;1(3):92-4.
- Lim HJ, Kim SY, Lee WK. Isolation of cholesterol-lowering lactic acid bacteria from human intestine for probiotic use. J Vet Sci. 2004 Dec 1;5(4):391-5.
- 49. Hassanein WA, Awny NM, Ibraheim SM. Cholesterol reduction by Lactococcus lactis KF147. African Journal of Microbiology Research. 2013 Aug 23;7(34):4338-49.
- Anandharaj M, Sivasankari B. Isolation of potential probiotic Lactobacillus oris HMI68 from mother's milk with cholesterol-reducing property. Journal of bioscience and bioengineering. 2014 Aug 31;118(2):153-9.
- 51. Girishkumar B and prapulla SG. 2010. Beneficial Properties of Lactic acid bacteria (LAB) Isolated from breast fed infants faecal flora: *in vitro* evidences Asian Jr. of Microbiol. Biotech. Env. Sc. 2010 :12 (4): 1-3.
- Sindhu SC, Khetarpaul N. Effect of feeding probiotic fermented indigenous food mixture on serum cholesterol levels in mice. Nutrition research. 2003 Aug 1;23(8):1071-80.
- Zeng XQ, Pan DD, Zhou PD. Functional Characteristics of Lactobacillus fermentum F1. Current microbiology. 2011 Jan 1;62(1):27-31.
- Kumar A, Kumar M, Ghosh M, Ganguli A. Modeling in vitro cholesterol reduction in relation to growth of probiotic Lactobacillus casei. Microbiology and immunology. 2013 Feb;57(2):100-10.
- 55. Yazdi MT, Zahraei M, Aghaepour K, Kamranpour N. Purification and partial characterization of a cholesterol oxidase from Streptomyces fradiae. Enzyme and microbial technology. 2001 Mar 8;28(4-5):410-4.
- Lekha PK, Lonsane BK. Production and application of tannin acyl hydrolase: state of the art. Advances in applied microbiology. 1997 Jan 1; 44:216-60.
- Sojo MM, Bru RR, García-Carmona FF. Rhodococcus erythropolis ATCC 25544 as a suitable source of cholesterol oxidase: cell-linked and extracellular enzyme synthesis, purification and concentration. BMC biotechnology. 2002 Dec;2(1):3.
- Lashkarian H, Raheb J, Shahzamani K, Hossein S, Shamsara M., 2010. Extracellular Cholesterol Oxidase from Rhodococcus sp. Isolation and Molecular Characterization. Iran Biomed J.;4(1 & 2):49–57.
- 59. Mahrous H. Probiotics bacteria from Egyptian infant's cause cholesterol removal in media and survive in

yoghurt. Food and Nutrition Sciences. 2011 Apr 26;2(02):150-155.

60. Guo L, Li T, Tang Y, Yang L, Huo G. Probiotic properties of E nterococcus strains isolated from traditional naturally

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fermented cream in C hina. Microbial biotechnology. 2016 Nov;9(6):737-45.

 Roukas T. Ethanol production from carob pods by Saccharomyces cerevisiae. Food biotechnology. 1993 Jul 1;7 (2):159-76.

*Corresponding Author:

Farhath Khanum* Email: farhathkhanum@gmail.com

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