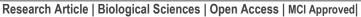


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COMPARITIVE PHARMACOGNOSTICAL, PHYTOCHEMICAL AND ANTIOXIDANT PROPERTIES OF THREE DIFFERENT *EUPATORIUM* SPECIES

Nithya.V1 and Kamalam.M2

¹Research scholar, ²Associate Professor, Department of Botany, PSGR Krishnammal college for women, Coimbatore- 641004.

*Corresponding Author Email: kamaluma12@gmail.com

ABSTRACT

The leaves of three different Eupatorium species such as Eupatorium glandulosum, Eupatorium odoratum and Eupatorium triplinerve belongs to the family Asteraceae, were screened in order to study the pharmacognostical, phytochemical and antioxidant property. The Physicochemical parameters such as Loss on drying, total ash, Acid insoluble ash, Water soluble ash and percentage of solubility was calculated. The leaf powders of different species were extracted with different solvents like petroleum ether, benzene, chloroform, acetone, methanol and water and the results revealed that polar solvents showed higher extractive value than non-polar solvents. The results of qualitative phytochemical analysis confirmed the presence of alkaloids, anthraquinone glycosides, phenols, glycosides, protein, amino acids, flavonoids, fats, fixed oils and saponins in the ethanol and water extracts. The leaves were also screened for the antioxidant property using standard ascorbic acid as a control and found that they possess good antioxidant property. Thus, the preliminary studies of the three-plant species could help us to understand the medicinal properties of the plant and also help us to construct monograph of the plant.

KEY WORDS

Pharmacological, phytochemical, antioxidant, Eupatorium glandulosum, Eupatorium odoratum, Eupatorium triplinerve

I. INTRODUCTION

Plants are an essential source of medicines and play a key role in world health. The contribution of medicinal plants is important to the global economy as approximately 85% of traditional medicine preparations involve the use of plants or plant extracts [1]. Natural products derived from plants are the essential sources of reliable bioactive compounds such as secondary metabolites and antioxidants. They are concentrated at different parts of plant such as leaves, flower, stem, bark, fruit, roots and seeds. Many plants contain several secondary metabolites have important applications in the fields of agriculture, human health and veterinary medicine [2].

Standardization of herbal drugs is an essential factor in order to assess the quality, purity, safety and efficacy of drugs. Development of standards for plant-based drugs being a challenging task, it needs innovative and creative approaches. The quality of plant will be analysed by different parameters such as identification, organoleptic, pharmacognostic, physiochemical and phytochemical properties [3].

E. glandulosum belongs to the family Asteraceae, is a native of Mexico, introduced as an ornamental shrub in several countries. In India, the tribes of Nilgiris use the leaves of the plant to heal wounds and small injuries [4]. In folklore medicine it is used as an astringent, thermogenic and stimulant [5].



The plant *E. odoratum* is a fast-growing perennial shrub, native of Central and South America has spread in tropical and subtropical regions of the world. The tribes of Indonesia used the leaf extract to cure skin diseases, poison bites, wounds, burns, cough, diabetes, diarrhea, fever, inflammation and rheumatism. The boiled roots are used to cure urinary disorders [6, 7, 8].

E triplinerve is a tropical American shrub commonly called as Ayapana is an ornamental erect perennial herb having aromatic leaves. In tribal medicine, it is used to cure fever with convulsions, pneumonia, indigestion, and cough [9].

Thus, to consider the importance these plants, the present study is aimed to investigate the comparative account of physicochemical, phytochemical and antioxidant properties of three different species of *Eupatorium*.

II. MATERIALS AND METHODS

Collection of plant material

The leaves of *E. glandulosum, E. odoratum* and *E. triplinerve* were collected from Nilgiri Hills of Western

Ghats, Coimbatore plain and Kanjikode in Kerela respectively and certified by Botanical survey of India in Coimbatore, Tamil Nadu. The collected leaves were washed thoroughly, dried, powdered and stored in air tight container for further study.

Extraction of plant material

The leaf powder of selected plant materials were extracted with different solvents like Petroleum ether, acetone, benzene, chloroform, ethanol and water.

Physicochemical parameters [10, 11]

The plant powder was subjected to calculate total ash, acid insoluble ash and water-soluble ash, loss on drying, solubility percentage in alcohol and water and extractive values.

Loss on drying

Freshly collected and pre-weighed samples were dried in Hot air oven at 45°C until it reaches a constant weight.

Total ash

3gm of leaf powder was taken in silica crucible and ignited in an electric muffle furnace at 100°C until the sample free from carbon. The percentage of total ash was calculated with reference to the air-dried sample.

Percentage of <u>ash value= Weight of fresh sample – Weight of dried sample X 100</u> Initial weight of the sample

Acid insoluble ash

Total ash obtained was heated with 25ml of diluted hydrochloric acid for 10 minutes, filtered in ash less filter paper (Whatman No.1) and the residue was incinerated in the furnace to get a constant weight. The weight of the insoluble matter was subtracted from the weight of total ash, represents the acid insoluble ash. The percentage of acid insoluble ash was calculated with reference to the air-dried drug.

Water soluble ash

The total ash obtained above was boiled with 25 ml of distilled water for 5 minutes. The insoluble matter was collected on an ash less filter paper, washed with hot water and dried to get constant weight at low temperature. The weight of the insoluble matter was subtracted from the weight of total ash, represents the water-soluble ash. The percentage of water-soluble ash was calculated with reference to the air-dried drug.

Solubility percentage

Alcohol

1gm of powdered material was mixed with 20ml ethyl alcohol and shaken frequently for 6 hours and kept undisturbed overnight. The extract was concentrated

and the solubility percentage was calculated on dry weight basis.

Water

The procedure adopted for solubility percentage of alcohol, is used to calculate the solubility percentage of water.

Extractive values

The powdered materials were extracted with different solvents like petroleum ether, benzene, chloroform, acetone, methanol and water in a soxhlet apparatus. The extracts were concentrated and the extractive values were calculated on dry weight basis.

Fluorescent analysis

Powdered plant materials were treated with different solvents and their illuminations were observed under ordinary and Ultra-violet light conditions [12].

Qualitative phytochemical analysis

Qualitative phytochemical analysis was done by using the procedure of Kokate [13]. Presence of alkaloids, flavonoids, glycosides, tannins, phenols, fixed oils, fats and saponins were analysed qualitatively.



Alkaloids (Mayer's Test)

Plant extract was treated with Mayer's reagent and formation of yellow colour precipitate indicates the presence of alkaloids.

Glycosides (Fehling's test)

Fehling reagent mixed with plant extract and gave red colour precipitate indicates the presence of glycosides.

Phenols and Tannins (Ferric Chloride Test)

Plant extracts were treated with 3-4 drops of 5% ferric chloride solution. Formation of bluish black colour indicates the presence of phenols.

Flavonoids (Alkaline Reagent Test)

Few drops of sodium hydroxide solution were added to the plant extract. Initially Formation of intense yellow colour, which becomes colourless on addition of dilute acid which indicates the presence of flavonoids.

Proteins and Amino acids (Ninhydrin test)

Ninhydrin reagent was added to the plant extract and boiled for few minutes. Formation of blue colour indicates the presence of proteins.

Anthroquinone Glycosides

To the plant extract, 5% potassium hydroxides solution was added. Appearance of red color indicates the presence of anthroquinones.

Fats and fixed oils

The extracts are treated with 0.5N alcoholic potassium hydroxide along with a drop of phenolphthalein and then heated on water bath for few minutes. Formation of soap or partial neutralization of alkali indicates the presence of fixed oils and fats.

Phytosterols (Salkowski's Test)

The plant extracts were treated with chloroform and filtered. The filtrates were treated with few drops of Con. Sulphuric acid, shaken and allowed to stand for few minutes. Appearance of golden yellow colour indicates the presence of phytosterols.

Saponins

1ml of the plant extract diluted with distilled water and made up to 20 ml and shaken in a test tube for 15 minutes. Formation of foamy layer indicates the presence of saponins.

Antioxidant activity

Preparation of standard solution

The standard solution was prepared by dissolving 1mg of Ascorbic acid in 1ml of methanol to obtain various concentrations such as 20, 40, 60, 80 and 100 μ g/ml.

Preparation of test sample

The ethanol and water extracts were dried and about 10mg of dried extract were dissolved in 10ml of methanol to give concentration of mg/ml.

DPPH Free Radical Scavenging Assay [14]

A solution of the radical is prepared by dissolving 2.4 mg DPPH in 100 ml methanol. A test solution of various concentrations such as 20, 40, 60, 80 and 100µl were added to 3.98, 3.96, 3.94, 3.92 and 3 ml of DPPH respectively. The mixture was shaken vigorously and kept at room temperature for 20 min in the dark. All the determinations were performed in triplicate. The DPPH reagent itself served as control. After 20 minutes the absorbance was measured at 515nm in spectrophotometer. The scavenging percentage of the extract was calculated using the following formula:

Hydrogen peroxide scavenging activity [15]

The leaf powder extracts (4ml) were prepared using distilled water at various concentrations. To the plant extract, 0.6 ml of 4mM Hydrogen peroxide solution which is prepared in 0.1M phosphate buffer (pH-7.4)

was added. The mixture was incubated for 10 minutes and absorbance was read at 230 nm in UV visible spectrophotometer. Ascorbic acid (control) was used as standard reference.

Hydrogen peroxide scavenging activity of powder extracts were calculated using the following formula,

% scavenged (H₂O₂) =
$$\frac{\text{(A control- A sample)}}{\text{(A control)}}$$
 X 100

Where,

A control - Absorbance of standard reference (Ascorbic acid)

A sample - Absorbance of the powder extract.



III. RESULTS

Physicochemical analysis

The leaf powder of three different *Eupatorium* species were screened for analytical values like moisture content, total ash, acid insoluble ash, water soluble ash and solubility percentage of ash in alcohol and water. The results observed that moisture content of E. odoratum was 19% and in E. glandulosum 24 %. The value of total ash and acid insoluble ash in E. glandulosum was found to be 2.1% and 0.4% respectively. The percentage of water-soluble ash content was almost equal in all the three samples. Among the three plants, the leaf powder of E. odoratum showed maximum solubility percentage in both water (24%) and ethanol (25%) (Table 1). Comparing the results of two solvents, plant powder of selected three plants extracted with ethanol showed maximum solubility percentage than water.

Extractive Value

The leaf powders of three plants were subjected to successive solvent extraction using different solvents in the soxhlet apparatus. The extractive values were observed to be better in polar solvents (water and ethanol) than non-polar solvents (Table 2). The extractive value was found to be maximum in ethanol (3.99%) and water (3.31%) extract of *E. odoratum* compared to other two plants.

Fluorescence analysis

The fluorescence behaviour of the powdered plant materials were studied by treating them with various solvents and observed under normal and UV light. No significant variations were observed among the three plants except *E. odoratum*. It revealed characteristic variation when it treated with ethanol (Table 3).

Phytochemical analysis

The preliminary screening of leaf extracts of all the three plants showed maximum phytoconstituents in ethanol and water extract (Table 4). The extract of *E. glandulosum* showed the presence of all the phytoconstituents tested except fats and fixed oils. Whereas in *E. odoratum*, phytosterols was absent in ethanol and water extract (Table 5) and in *E. triplinerve*, anthroquinone glycosides, fats, fixed oils and phytosterols were absent (Table 6).

Antioxidant activity

Antioxidant studies were carried out to find out the antioxidant's properties of the selected plant materials

by hydrogen peroxide scavenging method and DPPH method. Methanol and water extracts were used for the study. In DPPH method, the antioxidant activity was higher in all the extracts tested compared to the Hydrogen peroxide scavenging activity. The antioxidant activity of standard (ascorbic acid) increased with increasing concentration. In DPPH method, the ethanolic extract of *E. odoratum* and *E. glandulosum* exhibited higher antioxidant activity of about 81.35% and 75.87% respectively. Whereas *E. triplinerve* showed minimum antioxidant activity compared to other two plants. The scavenging percentage of plant extracts were increase with increasing concentration. (Figure 1 & 2).

IV. DISCUSSION

The leaves of three different *Eupatorium* species were analysed to identify their physicochemical, phytochemical and antioxidant property.

The physicochemical parameters help us to check the quality, standard and adulterants in the plant powder. Moisture is one of the major factors responsible for the deterioration of the drugs and affecting their shelf life [16]. In the present investigation, the moisture content of the leaves of all the three plants was found to be very low. Determination of ash value provides criteria for judging the purity of the drug [17]. A high ash value is indicator of contamination, substitution, adulteration or carelessness in drug preparation or drug formulation for marketing. In the present investigation, ash value was found to be very low in all the selected plants. The acid insoluble ash content was ranging between 0.3-0.4% in all the three plants which was found to be very low when comparing to the earlier report available in the leaves of Sesbania grandiflora and Wedelia trilobata [18, 19]. The remaining ash content showed the existence of inorganic components in the plant sample which shows the purity of the samples.

Extractive value plays an important role in evaluation of crude drugs. Less extractive value indicates addition of exhausted material, adulteration or incorrect processing during drying or storage or formulation [20]. In the present study, the extractive values were observed to be better in polar solvents than non polar solvents. The ethanol and water extracts showed highest percentage of extractive value in Ethanol extract



of *E. odoratum* (3.99%) and *E. glandulosum* (3.21%) indicated the quality of the drugs.

The fluorescence analysis is a sensitive and enables the precise and accurate determination of drug. The fluorescence colour is specific for each compound. A non-fluorescent compound may fluoresce if mixed with impurities which are fluorescent. The colour of the plant powder treated with different organic and inorganic solvents was observed under ordinary and UV light and found that there was no fluorescent compounds and not much colour variation among the species were observed.

The phytochemical screening of the three selected plants confirmed the existence of alkaloids, glycosides, flavonoids, saponins, tannins and phenols were observed in ethanol and water extract of all the three leaves. This result resembles the earlier reports on the leaves of *E. glandulosum* [21, 22], *E. odoratum* [23, 24, 25, 26] and *E.triplinerve* [27].

Antioxidant protects the body against the damaging effects of free radicals produced naturally within the body. These free radicals' production could cause damage to proteins, DNA and the genetic material within the cells [28]. In the present study, ethanol and water extracts of all the three plants showed good

antioxidant activity. Among the three plants ethanol extracts of *E. odoratum* (81.35%) and *E. glandulosum* (75.87%) revealed maximum antioxidant activity tested by DPPH method. This is due to the presence of flavonoids in plants and act as antioxidants. According to earlier report, the antioxidant activity of flavonoids has the ability to reduce free radical formation and scavenge the free radicals [29, 30]. In the present study, all the plant extracts showed the presence of flavonoids which are the reason behind the better antioxidant property. This finding resembles the earlier report available in *E. odoratum* [31, 32].

V. CONCLUSION

From the above findings, it is concluded that *E. glandulosum*, *E. odoratum* and *E.triplinerve* were subjected to various analysis such as physicochemical, phytochemical and antioxidant properties and found that there is no distinct variation among *E. glandulosum* and *E. odoratum* in physicochemical and phytochemical properties. But in *E.triplinerve*, it showed less antioxidant properties and lack of fats, fixed oils and phytosterols.

Table 1 Physicochemical analysis

S. No	Parameter studied	Value expressed in % (W/W)					
		E. glandulosum	E. odoratum	E. triplinerve			
1	Moisture content	24	19	25			
2	Total ash	2.1	1.9	1.5			
3	Acid insoluble ash	0.4	0.3	0.3			
4	Water insoluble ash	0.2	0.2	0.3			
5	Solubility %						
	1) Water	20	24	18			
	2) Alcohol	23	25	21			

Table 2 Extractive values

S.No	Solvent	Yield (%)				
		E. glandulosum	E. odoratum	E. triplinerve		
1	Petroleum ether	0.91	0.89	0.77		
2	Benzene	1.02	1.19	0.93		
3	Chloroform	1.67	1.56	1.19		
4	Acetone	2.13	2.29	1.87		
5	Ethanol	3.21	3.99	2.25		
6	Water	3.12	3.31	2.42		



Table 3 Fluorescence analysis

S.	Solvents	E. glandulosum		E. odoratum		E. triplinerve		
No		Normal light	UV light	Normal light	UV light	Normal light	UV light	
1	Powder as such	Green	Dark green	Green	Dark green	Green	Dark green	
2	Con.H ₂ SO ₄	Dark green	Dark green	Green	Dark green	Green	Dark green	
3	Con.HCl	Green	Blackish	Pale green	Blackish green	Pale green	Dark green	
			green					
4	Con.HNO₃	Dark green	Fluroscent	Light green	Dark green	Pale green	Dark green	
			green					
5	1N NaOH	Green	Dark green	Green	Dark green	Green	Dark green	
6	Ethanol	Light green	Blackish	Green	Reddish	Dark green	Blackish green	
			green		brown			
7	Water	Dark green	Reddish	Green	Pale green	Dark green	Blackish green	
			green					

Table 4 Qualitative phytochemical studies on Eupatorium glandulosum

S.No	Constituents	Petroleum Ether	Benzene	Chloroform	Acetone	Ethanol	Water
1	Alkaloids	+	+	+	+	+	+
2	Glycosides	-	+	-	+	+	+
3	Phenols & tannins	+	+	+	+	+	+
4	Flavonoids	-	-	-	+	+	+
5	Protein & amino acids	+	+	-	+	+	+
6	Anthroquinone Glycosides	-	+	-	+	+	+
7	Fats & fixed oils	-	-	-	-	-	-
8	Phytosterols	-	-	-	+	+	+
9	Saponins	-	-	-	-	+	+

⁺⁼Presence; -=Absence

Table 5 Qualitative phytochemical studies on Eupatorium odoratum

S.No	Constituents	Petroleum	Benzene	Chloroform	Acetone	Ethanol	Water
		Ether					
1	Alkaloids	+	+	+	+	+	+
2	Glycosides	-	-	+	+	+	+
3	Phenols & tannins	+	+	+	+	+	+
4	Flavonoids	+	+	+	+	+	+
5	Protein & amino acids	-	-	-	-	+	+
6	Anthroquinone	-	-	-	-	+	+
	Glycosides						
7	Fats & fixed oils	-	-	+	+	+	-
8	Phytosterols	-	-	-	-	-	-
9	Saponins	-	-	-	-	+	+

⁺⁼Presence; -=Absence

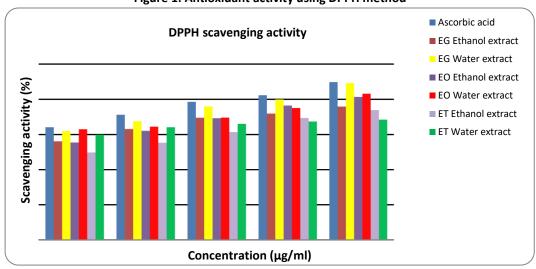


Table 6 Qualitative phytochemical studies on Eupatorium triplinerve

S.No	Constituents	Petroleum	Benzene	Chloroform	Acetone	Ethanol	Water
		Ether					
1	Alkaloids	-	-	+	+	+	+
2	Glycosides	-	-	-	+	+	+
3	Phenols & tannins	-	+	+	+	+	+
4	Flavonoids	-	-	-	+	+	+
5	Protein & amino acids	-	-	+	+	+	+
6	Anthroquinone	-	-	-	-	-	-
	Glycosides						
7	Fats & fixed oils	+	+	-	-	-	-
8	Phytosterols	-	-	-	-	-	-
9	Saponins	-	-	-	+	+	+

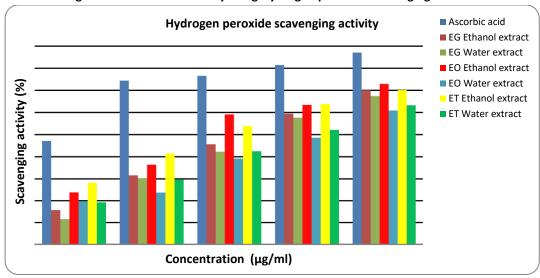
+=Presence; -=Absence

Figure 1: Antioxidant activity using DPPH method



EG-E. glandulosum, EO- E. odoratum, ET-E. triplinerve

Figure 2: Antioxidant activity using Hydrogen peroxide scavenging method



EG-E. glandulosum, EO- E. odoratum, ET-E. triplinerve



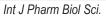
REFERENCES

- 1. Vieira RF, and Skorupa LA. Brazilian medicinal plants gene bank. *Acta Hort*, *330*: 51-58, (1993).
- 2. Kim D, Jeond S, Lee C. Antioxidant capacity of phenolic phytochemicals from various cultivars of plums. *Food Chem.*, *81*: 321-326, (2003).
- Rajani M, Kanaki NS. Phytochemical standardization of herbal drugs and polyherbal formulations. Bioactive Molecules and Medicinal Plants, (Ramawat KG, Mérillon JM (Eds.) Springer: 349-369, (2008).
- Monisha desingh, Jessy Mathew Jesudas, Prasun Balasubramaniam, Karthikeyan, Mayakrishnan, Bharath Ganesan and Ramya Mohan. Phytochemical analysis and anti-microbial activity of Eupatorium glandulosum. International journal of current microbiology and Applied science; 3(7): 882-885, (2014).
- Kritikar KR, Basu BD. Indian Medicinal Plants, Derhadun, India, 3: 1331-1333, (1987).
- Taylor RSL, Hudson JB, Manandhar, NP, Towers, GHN. Antiviral activities of medicinal plants of southern Nepal. *J., Ethanopharmacol*, 53: 97-102, (1996).
- Irobi ON. Antibiotic properties of ethanol extract of Chromolaena odorata (Asteriaceae). Inter J Pharmacognosy, 35: 111-126, (1997).
- 8. Amatya S, Tuladhar, SM. Invitro antioxidant activity of extracts from *Eupatorium odoratum*. *Res J .L., medicinal plant, 5*:79.-86, (2011).
- 9. Shahadat Hossan, Abu Hanif, Mujib Khan, Sazzadul Bari, Rownak Jahan, Mohammed Rahmatullah. Ethnobotanical survey of the Tripura tribe of Bangladesh. *American-Euras ian Journal of Sus tainable Agriculture*, 3(2): 253-261, (2009).
- 10. Trease, GE and Evans WE. Pharmacognosy, 13th edition, Bailliere Tindall, London, (2005).
- 11. Kokate CK. Evaluation of crude drug. (ed). Practical Pharmacognosy, 5th ed.: M.K.Jain, 125-127 (2014).
- 12. Chase CR and Pratt RJ. Fluorescence of powdered vegetable drugs with particular reference to development of a system of identification. *Journal American pharmacology association*, 38:32, (1949).
- Kokate CK. Analytical Pharmacognosy. (ed).
 Pharmacognosy, 33rd ed.: Nirali Prakashan, 97-132, (2005).
- 14. Brand-Williams W, Cuvelier ME, Berset C. Use of free radical method to evaluate antioxidant activity. *Lebens-Wiss Technol*, 28:25-30, (1995).
- 15. Ruch RJ, Cheng SJ, Klaunig JF. Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chinese green tea. *Carcinogenesis*, *10*: 1003–1008, (1989).
- 16. Ruchi Tripathi, Suryakant Verma TS, Easwari and Harish Shah. Standardization of some herbal antidiabetic drugs

- in polyherbal formulation & their comparative study. *IJPSR*, *4*(*8*): 3256-3265, (2013).
- 17. Victor Kuete. Medicinal spices and vegetables from Africa. Academic press- An imprint of Elsevier, 181-183, (2017).
- Momin, RK and Kadam VB. Determination of ash values of some medicinal plants of genus sesbania of marathwada region in Maharashtra. *Journal of Phytology*, 3(12): 52-54, (2011).
- Karthika, C and Manivannan, S. Pharmacognostic, physicochemical analysis and phytochemical screening of the leaves of W. trilobata L. International Journal of ChemTech Research, 11(02): 124-131, (2018).
- 20. Chandel, HS, Pathak AK and Tailang M. Standardization of some herbal antidiabetic drugs in polyherbal formulation. *Pharm. Res., 3(1)*: 49-56, (2011).
- 21. Arvind Negi, Upadhyay, Amit Semwal and Arun Kumar Wahi,. Pharmacognostical studies on the leaves of *Eupatorium adenophorum* Sprengs. *Pharmacognosy, 2* (15): 01-08, (2010).
- 22. Rajeswary, M and Govindarajan, M. Mosquito larvicidal and phytochemical properties of *Ageratina adenophora* (Asteraceae) against three important mosquitoes. *J Vector Borne Dis*, 141–143, (2013).
- Arvind Negi, Upadhyay, Amit Semwal and Arun Kumar Wahi, Pharmacognostical studies on the leaves of Eupatorium adenophorum Sprengs. Pharmacognosy, 2 (15): 01-08, (2010).
- 24. Okwu, DE. Phytochemical and vitamin content of indigenous spices of South Eastern Nigeria. *J., Agric. Environ.*, *6*(1): 30-34, (2004).
- 25. Liu RH. Potential synergy of phytochemicals in cancer prevention: mechanism of action. *Nutr. J., 134*: 3479-3485, (2004).
- 26. Afolabi, C, Akinmoladun, EO, Ibukun IA, Dan-Ologe. Phytochemical constituents and antioxidant properties of extracts from the leaves of *Chromolaena odorata*. *Scientific Research and Essay, 2 (6)*:191-194, (2007).
- 27. Sugumar S, Karthikeyan and Gowdhami T. Preliminary Photochemical Screening on the Leaf Extract of Eupatorium triplinerve Vahl. International Journal of Pharmaceutical & Biological Archives, 5(5): 141 144, (2014).
- 28. Weisburger JH. Tea In: The Cambridge world History of food. (Eds:K.Kipple and K.C.Orneal). Cambridge Univ.Press. Cambridge, 712-720, (2000).
- 29. Pier-Giorgio Pietta. Flavonoids as Antioxidants. *J. Nat. Prod*, 63:1035-1042, (2000).
- 30. Amarowicz, R, Pegg, RB. Legumes as a source of natural antioxidants. *Eur J Lipid Sci Technol* 110: 865-878, (2008).
- 31. Anup Chakraborty, Harikrishna Roy, Shailaja Bastia.

 Evaluation of antioxidant activity of the leaves of

 Eupatorium odoratum L. International Journal of





Pharmacy and Pharmaceutical Sciences, 2 (4): 77-79, (2010).

32. Rajalakshmi P, Sumathi V, Pugalenthi M. Antioxidant activity of Erigeron Karvinskianus DC. and *Ageratina*

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Adenophora (Spreng) King (leaves). *International Journal of Food Science and Nutrition*, 1(5): 64-68, (2016).

*Corresponding Author: *M.Kamlam**

Email: kamaluma12@gmail.com