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CHARACTERIZATION OF HETEROTROPHIC BACTERIA FROM THE SEDIMENT OF MARINE CAGE FARM OF POLEM, GOA

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ABSTRACT

Present study aimed at characterising the sediment bacterial population from the cage farm of Polem, Goa with reference to phenotypic and biochemical characterization. Total of 9 isolates were selected for the present study based on the colony morphology and were identified by conventional biochemical and molecular techniques as Bacillus firmus, Bacillus cereus, Bacillus subtilis, Bacillus aryabhattai, Acinetobacter sps, Vibrio harveyi, Vibrio algenolyticus, Ferrimonas balearica and Oceanomonas smirnovii. The study suggests that the sediment bacteria from the cage farm can be used as biological indicator of farming practices.

KEY WORDS

Cage farm, Bacteria, Sediment, Polem

INTRODUCTION

Marine ecosystem is highly dynamic, providing a high diversity of ecological niches. Biodegradation of organic matter and energy transfer in marine sediment is mainly influenced the presence of heterotrophic bacteria [1] and the structure of microbial assemblage is sensitive to changes in environmental conditions and tropic state [2]. Further, it is thought that marine sediment harbours around 55–85% of all microbial life [3]. A large number of studies are conducted on the role of heterotrophic bacteria in releasing the organic substrate and inorganic nutrients in to the higher trophic levels [4,5].

Marine finfish culture is rapidly growing food sector in the Asia pacific. Previous studies shown that disturbance in coastal areas due to cage farming practices might determine changes in the community structure and activity of benthic bacteria [4]. Knowledge on the microbial community composition may give important information about the health of the ecosystem, since bacteria responds quickly to the abiotic changes in the environment [5]. Measurement of microbial assemblages in the sediment has been recently proposed as tools for monitoring the impact of intensive aquaculture activities [4]. Although bacterial communities have been frequently studied, recent studies show that lots of bacterial communities need to be still characterized in marine environment [6,7,8]. Further, the information on the total culturable heterotrophic bacterial population from the cage farms in India is insufficient. The aim of this work was to isolate and identify the bacterial population from sediment of cage farm by conventional methods and 16s rDNA sequencing.

MATERIALS AND METHODS

Study area

Present study was carried out in marine fish farm located in Southern part of Goa, Central west coast of India (Polem village 14° 54' 21.12" N ;74° 04' 32.20" E). The cage farm consisting of 25 no's of floating circular GI cages of 6m diameter with Asian seabass, pompano and cobia cultured in it.



Isolation of bacteria and biochemical characterization

Sediment samples were collected from the bottom of the cage farm during September 2014 to May 2016. Sediment samples were collected in sterile polythene bags by using Van veer grab sampler (0.042m²) and transported to the laboratory in an ice cold condition. Microbiological analysis of the culturable heterotrophic bacterial fraction was carried out by pour plate method. For qualitative estimation of bacteria, distinct colonies were selected from each plate and isolated in pure culture on Marine Agar (Himedia, India) and incubated at 30°C \pm 0.2°C for 24 to 48 hrs. For taxonomic identification, purified bacterial samples were subjected to morphological and biochemical characterization such as Gram stain, catalase test (3% H₂O₂), oxidase test, indole [9], oxidative/fermentative acid production from glucose, citrate utilization, nitrate reduction, Methyl red (MR)-Voges Proskauer (VP), degradation of urea, followed by growth on TCBS (thiosulphate citrate bile salts sucrose), growth at different salt (NaCl) concentrations and different temperature, were performed according to the standard methods [10].

Molecular characterization

For molecular identification, pure cultures of the phenotypic isolate in log phase in Luria-Bertani (LB) broth were used for pure genomic DNA extraction using bacterial genomic DNA extraction kit (Sigma, USA). For identification of bacterial isolates (1-7) 16S rRNA gene was amplified in a thermal cycler (Eppendorf, Germany) using the combination of universal primers 27F (5' AGAGTTTGATCCTGGCTCAG 3') and 1492R (5' GGTTACCTTGTTACGACTT 3'), while specific primers were used for confirming the Vibrio harveyi (VAHF1) 5'-ATCATGAATAAAAC TATTACGTTACT-3' and (VAHR1) 5'-AAAGGATGGTTTGACAAT-3' following Fukui and Sawabe [11] and Vibrio algenolyticus VA-F (5' -CGAGTACAGTCACTTGAAAGCC- 3') and VA-R (5' -CACAACAGAACTCGCGTTACC- 3') following Abdel-Aziz et al. [12]. The amplified PCR products were confirmed by running the amplification product in 1% agarose gel in 1X TAE. The amplified products (1500 bp) were purified individually using the nucleic acid purification kit (Axygen Biosciences, USA) and sanger sequenced. Bacterial identity was deduced by searching Genbank using BLAST algorithm. The partial 16S rDNA sequences derived in this study have been deposited in GenBank.

RESULTS AND DISCUSSION

Marine substrates generally provide a nutrient-rich environment where bacteria grow in complex interactive biofilm communities [13]. Total of 9 isolates were selected for present study for biochemical and molecular characterization. These included Bacillus firmus, Bacillus cereus (MG782844), Bacillus subtilis (MG782866), Oceanomonas smirnovii (MG800693), Ferrimonas balearica (KP406159), Acinetobacter sps (KP718120), Bacillus aryabhattai (KP718121), Vibrio Vibrio algenolyticus. harveyi and Phenotypic characterization of bacterial isolates showed creamy yellow, dull white, pale yellow colour with most of the isolates having round to irregular shaped colony (Table 1). All the isolates except Acinetobacter sp were motile. Gram-positive bacteria comprise a relatively large proportion in the marine sediments and are likely to play important microbiological roles in the marine environment mainly the biogeochemical processes. In the present study, the selected isolates comprised of 40% gram positive and 60% were gram negative. Among the gram positive group, the genus *Bacillus* is one of the well-known genera and members of these genera have strong adaptability to diverse conditions and several species belonging to the genus are capable of producing highly resistant spores [14]. During the present study, 4 isolates were found to be gram positive which includes Bacillus cereus, Bacillus subtilis, Bacillus aryabhattai and Bacillus firmus. Miranda et al. [15] reported that, Bacillus cereus and Bacillus subtilis were the most frequently (78%) occurring strains from marine sediments. B. cereus causes two distinct types of food poisoning i.e. diarrhoeal and emetic (vomiting), causing two types of illness [16] and also fatal meningitis [17]. This organism is also responsible for spoilage of different food products especially of plant origin, but also isolated from the meat, egg and dairy products [18]. Isolates of Bacillus subtilis from the marine sediments has many industrial application. B. subtilis were used to increase the rotifer production and control bacterial growth in the cultures [14]. Study also shown that B. subtilis can be used in laboratory based bioremediation experiments along with other species of Bacillus viz B. megaterium and B. polymyxa [19]. Ray et al. [20] isolated Bacillus aryabhattai the plant growth promoting bacteria from the wetlands of east Kolkata and reported that the isolate can survive in extreme condition. Shvaiji et al. [21] initially isolated this

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bacterium from cryotubes used to collect air samples from the Earth's stratosphere at an altitude between 27 and 41 km. Lee et al. [22]; Pailan et al. [23] and Yan et al. [24] isolated this bacterium from rhizosphere soil from South Korea, India, and Tibet. During the present study this bacterium was isolated from the sediment collected from marine cage farm but the source of this bacterium in sediment is unclear. *Bacillus firmus*, lead resistant bacteria used for its ability in biosorption of lead was isolated from the cage farm sediments. Alireza et al. [25] isolated to *B. firmus* from the surface sediment of Imam Khomeini port.

Acinetobacter is a gram-negative, coccobacilli, nonspore-forming, and non-motile bacteria with milky white colored colonies. Members of the genus Acinetobacter are ubiquitous in nature and can be isolated from soil, water samples and sewage with appropriate enrichment techniques [26]. During present study this bacterium was isolated from the marine cage farm sediment and presence of Acinetobacter in the sediment indicates the competitive degradation of organic matter [27]. Among the gram negative bacteria, vibrios are the most commonly occurring group in the marine environment either in the free form in the water and sediment or associated with the diverse marine animals. Vibrio algenolyticus and Vibrio harveyii are most frequently isolated from the coastal sediments and water and are considered as opportunistic pathogens for aquatic animals such as fishes, shellfishes and crustaceans causing serious economic damage in the culture by causing the disease [28]. During the present study, V. algenolyticus and V. harveyii were isolated from marine farm sediment from Polem. Similler reports were made by Liu et al. [29] in the sea cage farm of Xiangshan Bay, China. Ferrimonas balearica a new facultative iron reducing bacteria isolated from the sediment during the present study. Similar reports on isolation of this bacteria were made by Rossello' -Mora et al. [30] from marine sediment at the Mediterranean coast of Mallorca (Spain). The genus Oceanimonas are the group of marine Proteobacteria, which are motile- gram negative rods comprising of three known species O. baumannii, O. doudoroffii and O. smirnovii. Among this O. smirnovii. is a novel bacterium isolated from the marine sediments [31].



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	Bacillus Bacillus Bacillus Acinetobacter Ferrimonas Oceanomonas Vibrio Vibrio								
	Bacillus firmus	cereus	subtilis	aryabhattai	sps	balearica	smirnovii	harveyi	algenolyticus
Morphology	Creamy yellow	Opaque	Dull white	Creamy	Pale yellow	Pale yellow	Light creamy	Pale yellow	White
Shape	Round	Irregular	Irregular	undulated	Smooth	Round	round	Round	Round
Gram Stain	Rods +ve	Rods +ve	Rods +ve	Rods +ve	-ve Bacilli	Rods –ve	Rods –ve	Rods –ve	-ve
Motility	+	+	+	+	-	+	+	+	vc +
Spore formation	+	- -	+	+	_	_		-	-
Indole	-	-		-		_			
MR	-		-	_		-	т	+	+
VP	-	-	-	-	-	_		т	т
Citrate	-	+	+	Ŧ	-	-		-	-
	-	+	+	-	+	-	+	+	+
Catalase	+	+	+	+	+	+	+	+	+
Oxidase	+	-	+	+	-	+	+	+	+
H ₂ S production	-	-	-	-	ND	+	-	-	-
Urease	-	-	-	+	-	+	-	ND	-
Nitrate	+	+	+	+	-	+	+	+	+
Glucose	+	+	+	+	+	-	+	-	+
Sucrose	+	+	+	-	-	-	-	-	+
Gas production	-	-	-	-	-	-	-	-	-
Lactose	-	-	+	+	+	ND	-	-	-
Starch	+	+	+	+	-	-	-	+	+
TCBS	-	-	-	-	-	Green	ND	Green	Yellow
Growth at									
5º C	-	+	-	-	-	-	-	-	-
10° C	-	+	+	+	+	+	+	-	
40° C	+	+	+	+	+	+	+	-	
50° C	-	-	+	-	-	-	-	-	-
NaCl growth (%)	2-6%	2-6%	2-8%	2-6%	0-6%	0-8%	0-8%	2-8%	2-6%
Gen Bank Acce. No	NS	MG782844	MG782866	KP718121	KP718120	KP406159	MG800693	NS	NS

Table 1: Biochemical characteristics of the bacterial isolates

VP, Voges Proscaeur; MR, Methyl red; ND, not done; (+), growth; (-), no growth; NS (not submitted)

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CONCLUSION

Identification of microbial fauna from the sea sediment is one the difficult area of biodiversity research which helps in understanding the role of these organisms in community assembly and ecological processes. The heterotrophic bacterial diversity in marine cage sediments is little known, which can have considered as bio indicator for assessing the health of farm environment.

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