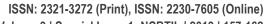
# **International Journal of Pharmacy and Biological Sciences**



IJPBS | Volume 9 | Special Issue 1- NCBTIL | 2019 | 157-162

National Conference on Biochemistry –Transcending and Integrating Life Sciences (NCBTIL) -2019 Held @ Kristu Jayanti College, K Narayanapura, Kothanur PO, Bengaluru-560077, January 17th & 18th

| Conference Proceedings | Research Article | Biological Sciences | Open Access | MCI Approved |

**|UGC Approved Journal|** 



# Biosynthesis of Silver Nanoparticles from Trichoderma Harzianum And Its Impact on Germination Status of Rice and Brinjal

Saravanan. K\*, Anitha. M, Rajabairavi.N, P.Arivazhagan and S. Jayakumar

PG and Research Department of Biotechnology, Selvamm Arts and Science College, Namakkal.

Received: 10 Dec 2018 / Accepted: 30 Dec 2018 / Published online: 10 Jan 2019 Corresponding Author Email: <a href="mailto:snrsaro88@gmail.com">snrsaro88@gmail.com</a>

# **Abstract**

The present study highlighting on the biosynthesis of silver nanoparticles by using the fungus *Trichoderma harzianum*. The cell filtrate of *Trichoderma harzianum* reacted with 1 mM silver nitrate solution, resulting the formation of silver nanoparticles within 3 hours. The silver nanoparticles were characterized by Visual analysis, UV-Vis absorption spectroscopy and Transmission electron microscopy (TEM). Biosynthesized silver nanoparticles exhibited maximum absorbance at 440nm in UV-Vis spectroscopy. TEM showed polydisperse spherical and occasionally ellipsoid nanoparticles in the size range from 20-60 nm and average size 35 nm. Disease-free healthy-looking seeds of Brinjal (Disco) and Rice (Samba) per-soaked in 3 days old silver nanoparticles solution of *Trichoderma harzianum* for 2hr and 5hr soaking period and. It is clear form the result that percentage of seed yield was enhanced irrespective of the synthesized silver nanoparticles solution. *T. harzianum* synthesized silver nanoparticles showed increase in percentage of yield with increased in soaking time of silver nanoparticles solution. *T. harzianum* synthesized silver nanoparticles have biological assay used in agricultural purposes to increases the viability of yield.

### **Keywords**

Silver nanoparticles, *Trichoderma harzianum*, Transmission Electron Microscopy, biological assay.

## INTRODUCTION:

Nanotechnology is the science and technology of small things i.e. 'nano'. This term originates from Greek

word meaning 'dwarf'. This term was firstly used by 'Richard Feynman' in 1959. Therefore, nanotechnology term was used from last 53 years. Microbes play an

157



important role in nanotechnology due to the synthesis of nanoparticles by biological method. Microbes are the microscopic organisms which are single or multicellular found in all universes. They include the bacteria, algae, fungi and protozoa. Microbes play an important role in balancing the ecosystem by various processes like soil fertility bio-degradation, sewage treatment and improving agricultural productivity. Microbes are known as decomposer in the ecosystem especially fungi well it grows on living or on dead organism.

A fungus produces several hydrolytic enzymes like Amylases, proteases, Lactase, Pectinases, Catalase, Penicillinase, Glucosidases etc. Mycotoxins like Aflatoxin, Zearalenone, Ochratoxin, Citrinin, T-2 toxin, Fumonisins etc. Pigments like qunonine, phynolic group, and also synthesizes the Nanoparticles like Silver (Ag), Gold (Au), Platinum (Pt), Copper (Cu) <sup>2,1,5</sup>. Biological synthesis of nanoparticles has gained more attention by the researchers for its potential applications<sup>15;5</sup>. There are also have been several reports on the biosynthesis of AgNPs using fungi, including Fusarium oxysporum <sup>3</sup> Fusarium acuminatum <sup>10</sup>, Penicillium fellutanum <sup>12</sup>Aspergillus clavatus <sup>10</sup>, F. solani <sup>11</sup>, Aspergillus niger <sup>10</sup>, Alternaria alternata <sup>9</sup> etc. have been successfully used for the synthesis of silver nanoparticles. Silver ions and silver-based compounds are highly toxic to living organisms because when it is a higher concentration in the cell death occurs due to more reactive silver ions, but when it is in small concentration i.e. in nanoparticle which is useful to living organisms.

In this article, the cell filtrate of this fungus *Trichoderma harzianum* was used for the synthesis of silver nanoparticles. Silver nanoparticles were observed within 1hr after incubation with AgNo3 solution in to cell filtrate. However, to study the effect of biosynthesized silver nanoparticles on germination of oil seeds was also done.

# MATERIALS AND METHODS

#### **Collection of Materials:**

The present study was undertaken with a view to study the biosynthesis of silver nanoparticles in T. harzianum fungi was isolated from agriculture soil and maintained on potato dextrose agar (PDA) medium at 28C°. The isolated fungus was identified by lacto phenol cotton blue mounting by morphological and microscopic

observation. Pure culture was maintained on potato dextrose agar slants at 28C°. Healthy sunflower and soybean seed were collected from market to check the effect of silver nanoparticles on seed germination.

#### **Biosynthesis of Silver Nanoparticles:**

Glucose nutrient broth medium (GNB) was used for biomass preparation of *T. harzianum*. 25gm of clean fresh fungal biomass was again inoculated in 100 mL of double distilled water for 3 days at 30°C and agitated again at 120 rpm. The cell filtrate was obtained by filtering it through Whatman filter paper No. 1 and the cell free filtrate was collected for experiment. The 10 mL filtrate was treated with 10 mL of 1 mM AgNo3 solution in an Erlenmeyer flask and incubated at room temperature in dark. Control containing cell-free filtrate without silver nitrate solution was run simultaneously as standard with the experimental flask. All experiments were done in duplicate.

#### **Characterization of Silver Nanoparticles:**

### **UV-visible spectroscopy analysis**

Color of the cell free filtrate changes after the incubation of silver nitrate solution was visually observed. Silver ion bio-reduction was monitored by sampling of aliquots (1 mL) at different time intervals. Absorption measurements were carried out on UV-visible spectrophotometer (Cystronics UV-Vis spectrophometer 117) and absorbance was measured between 300-600 nm.

Transmission electron microscope (TEM) Synthesized AgNPs drop was placed on the carbon coated copper grids and kept for dry. After dryness of sample grid loaded on to a specimen holder. TEM images of the sample were taken using the Morgagni 268D TEM instrument.

# Effect of biosynthesized silver nano-particles on seed germination:

Seeds of soybean and safflower were surface sterilized with 1% mercuric chloride solution for 1 min. and rinsed several times in sterile distilled water. Clean seeds were per-soaked in 3 days old silver nanoparticles solution of T. harzianum for varying period of time (2hr and 4hr) in undiluted solution. Control containing water in which seeds were presoaked was run simultaneously as standard with the experimental flask.



#### **RESULTS**

### **Biosynthesis of AgNPs:**

When cell-free filtrate of *T. harzianum* isolates was incubated with silver nitrate salt, the color of cell filtrate was exhibited a gradual change to brown color

under dark condition. The color of the culture filtrate with silver nitrate salt changed to intense brown after 24hr of incubation whereas the control (without silver nitrate salt) did not exhibit any color change.



Figure-1. Color of sample A) 1 mM Silver nitrate solution B) Crude cell filtrate of *T. harzianum* before immersion of AgNo3 C) after immersion of AgNo3.

#### **Characterization of AgNPs:**

The UV-visible spectra of *T. harzianum* fungal cell filtrate of treated with the silver nitrate solutions showed a characteristic surface plasmon absorption band at 440nm which are nearby similar to result of reported an intense peak at 410nm<sup>14</sup>. It is reported that the absorption spectrum of spherical silver nanoparticles presents a maximum between 420nm and 450nm <sup>13&6</sup>. Banu and Rathod, (2011) indicating the synthesis of silver nanoparticles and the maximum

color intensity was obtained after three days. Beyond three days

of incubation, no further increase in intensity was recorded indicating complete reduction of silver ions by the fungal cell filtrate. Synthesized AgNPs was extremely stable at room temperature, without agglomeration was monitored regularly by UV-visible spectrophotometer. This indicated that the nanoparticles were well dispersed in the solution without aggregation (Figure-2).

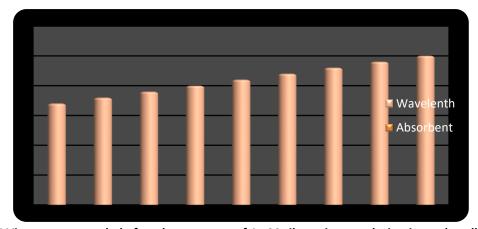
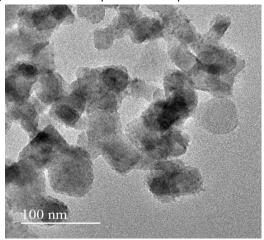


Figure-2. UV-Vis spectra recorded after the exposure of 1mM silver nitrate solution in crude cell filtrate of *T. harzianum*.



TEM & SEM micrograph provided detailed morphology of silver nanoparticles. The data obtained from micrograph showed distinct shape and size of polydisperse nanoparticles. Mostly particles were spherical, but some are ellipsoidal in shape and 20-60

nm and average size 30 nm in size without significant agglomeration (Figure-3). TEM provided confirmation of presence of silver nanoparticles with detailed size and shape which are similar to the results <sup>04& 07</sup>.



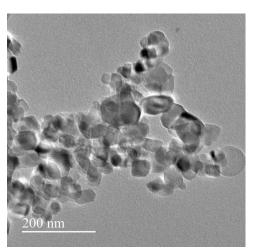


Figure-3. TEM micrograph (200nm and 50nm) of silver nanoparticles synthesized by T. harzianum.

# Effect of silver nanoparticles on germination status of Rice and Brinjal:

From Table no-1 it is clear that silver nanoparticle solution of T. harzianum found optimistic effect on germination of soybean and sunflower as compared to control. Silver nanoparticles solution showed maximum effect on germination status of soybean and sunflower. As compered soybean, sunflower shows germination irrespective maximum of silver nanoparticles solution. Results indicates that increasing the soaking period of silver nanoparticles solution, will increase the germination of soybean and sunflower which indicates that's seed germination is directly proportional to soaking period of silver nanoparticles solution (Figure-3).

# Effect of *Silver nanoparticles* on Brinjal Crop. Effect on Shoot length:

The shoot length (in cm) were recorded in the *Silver nanoparticles* treated and Control Brinjal crop at different time interval viz., 45, 55, 65, and 75 days after transplanting. The Brinjal crops were found to have a good shoot length in compared to the control crop.

## **Effect on Root length:**

After 45 days of *Silver nanoparticles treated* the root length was taken in the Brinjal crop as well as in control. The Silver nanoparticles Brinjal crops were found to have a good root length in compared to the control Crop Fig 4.

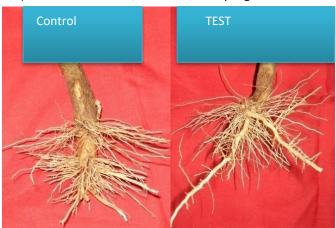


Fig.4: Root length of Brinjal at harvesting stage for different treatments of Control and Silver nanoparticles treated.



#### **Effect on Yield:**

The yield of Brinjal crop was found to be higher in *Silver nanoparticals* treated field as compared to the control one (Table 1).

Table 1: Effect of Silver nanoparticals Treatment on Brinjal crop.

Control						TEST						
Shoot Length in different Days (cm) Root length (cm)				length	Yield (kg)	Shoot Length in different Days (cm)				Root length (cm)	Yield (kg)	
45 <sup>th</sup>	55 <sup>th</sup>	65 <sup>th</sup>	75 <sup>th</sup>			45 <sup>th</sup>	55 <sup>th</sup>	65 <sup>th</sup>	75 <sup>th</sup>			
<b>day</b> 20.5	<b>day</b> 25.00	<b>day</b> 33.03	<b>day</b> 40.1	6	50	day	day	day	day	9	58	
						23.5	27.9	36.6	43.16			

# Effect of Silver nanoparticles treated and on Rice (Samba) Crop.

### **Effect on Shoot length:**

The shoot length (in cm) was recorded in the *Silver* nanoparticles treated and Control Rice (Samba) crop at different time interval viz., 45, 55, 65, and 75 days after transplanting. The *Silver nanoparticles* treated Rice

crops were found to have a good shoot length in compared to the control crop.

#### Effect on Root length:

After 45 days of *Silver nanoparticles* treatment the root length was taken in the Rice (Samba) as well as in control. The *Silver nanoparticles* treated Rice was found to have a good root length in compared to the control crop Fig 5.



Fig.5: Root length of Rice (Samba) at harvesting stage for different treatments of Control and Silver nano particals treated.

#### Effect on Yield:

The yield of Rice crop (Samba) was found to be higher in *Silver nanoparticals treated* field as compared to the control treated one (Table 2).

Table 2: Effect of Silver nanoparticals Treatment on Rice (Samba) crop.

Control						TEST						
Shoot Length in different Days (cm)				Root length (cm)	Yield (kg)	Shoot Length in different Days (cm)				Root length (cm)	Yield (kg)	
45 <sup>th</sup>	55 <sup>th</sup>	65 <sup>th</sup>	75 <sup>th</sup>			45 <sup>th</sup>	55 <sup>th</sup>	65 <sup>th</sup>	75 <sup>th</sup>			
day	day	day	day	6	38	day	day	day	day	10	47	
21.3	25.00	30.03	41.1			23.33	27.36	35.56	42.6			



#### **DISCUSSION:**

From above result it can be concluded that *T. harzianum* secrete large amount of proteins and enzymes so there is no need of reducing agent and stabilizer like as chemical method. Thus, the present study has reported the biological process for the synthesis of silver nanoparticles using *Trichoderma harzianum*.

Characterization was made by UV-Visible absorption spectroscopy which shows maximum absorption at 440nm, Transmission Electron Microscope (TEM) revealed the formation of spherical and ellipsoid nanoparticles with size ranging between 19-63 nm and average size 34.77 nm with no agglomeration.

#### **CONCLUSION:**

Biosynthesized silver nanoparticles have optimistic effect of seed germination. Thus, results conclude that maximum soaking period of silver nanoparticles will allow the maximum soaking of silver nanoparticles through seed coat and scutellum which ultimately effect on physiology of the seed. Therefore, biosynthesized silver nanoparticles have biological assay used in agricultural purposes to increases the viability of seeds.

### **REFERENCES**

- Ahmad, A., Mukherjee P., Senapati, S., Mandal, D., Khan, M., Kumar R., Sastry M., 2003. Extracellular Biosynthesis of silver nanoparticles using the fungus Fusarium oxysporium. Colloid Surface, 28, 313,
- Ahmad, A., Mukherjee, P., Mandal, D., Senapati, S., Khan, M., Kumar, R., Sastry, M. 2002. Enzyme Mediated Extracellular Synthesis of CdS Nanoparticles by the fungus, *Fusarium oxysporum*. Journal of American Chemical Society. 124; 120108-12109.
- Ahmad, A., Mukherjee, P.Senapati, S., Mandal, D., Khan, M.I., Kumar, R. and Sastry, M. 2003. Extracellular biosynthesis of silver nanoparticles using the fungus Fusarium oxysporum, Colloids and Surfaces B: Biointerfaces 28:313-318.
- Anand KR. Thakur and Nanda Ghosh. 2014. Correlation between ecological factors and diversity of Agylla remelana, moore (lepidoptera: Noctuidae) at Bariyatu, Ranchi, Jharkhand, India. Biolife. 2(2);415-419.
- Bansal, V., Rautaray, D., Ahmed, A. and Sastry, M. 2004. Biosynthesis of Zirconia nanoparticles using fungus *Fusarium oxysporum*. J. of Materials Chemistry. 14:3303-3305.

- Banu A and Rathod V. 2011. Synthesis and characterization of silver nanoparticles by *Rhizopus* stolonifer, International Journal of Biomedical and Advance Research, 2: 5.
- Estari Mamidala, Rajendra Prasad Gujjeti, Sainath Namthabad, 2014. Calotropis gigantea flowers extracts with HIV-1 reverse transcriptase (RT) inhibitory activity. World Journal of Pharmacy and Pharmaceutical Sciences. 3 (9), 1016-1022.
- Gade A., Bonde P., Ingle A., Marcato P., Duran N., Rai M. 2008. Exploitation of *Aspergillus niger* for synthesis of silver nanoparticles," J. Biobased. Mater. Biener, 2,243.
- Gajbhiye M., Kesharwani J, Ingle A., Gade A., Rai, M. 2009. Fungus mediated synthesis of silver nanoparticles and their activity against pathogenic fungi in combination with fluconazole," Nanomedicine: Nanotech. Bio. And Med. 5, 383.
- Ingle A., Gade A., Pierrat S., Sonnichsen C., Rai M. 2008.
  Mycosynthesis of silver nanoparticles using the fungus Fusarium acuminatum and its activity against some human pathogenic bacteria, Curr. Nanosci. 4, 141.
- Ingle I., Gade A., Bawaskar M., Rai M. (2011). Fusarium solani a novel biological agent for the extracellular synthesis of silver nanoparticles. J. Nanopart. Res. 11, 2079.
- Kathiresan K., Manivannan S., Nabeel M.A., Dhivya B. 2009. Studies on silver nanoparticles synthesized by a marine fungus, *Penicillium fellutanum* isolated from coastal mangrove sediment," Colloid surface B. 71, 13.
- 13. Maliszewska J, Szewczyk K and Haszak K. 2008. Biological Synthesis of Silver Nanoparticles. Journal of Physics: Conference Series, 146. 012025.
- 14. Mukherjee P, Roy M, Mandal B, Dey G, Mukherjee P, Ghatak J, Tyagi A, Kale S. 2008. Green synthesis of highly Stabilized nanocrystalline silver particles by a non-pathogenic and agriculturally important fungus *T. asperellum*. IOP publishing Nanotechnology journal, 19: 075103(7pp).
- Narayanan K. and N. Sakthivel, 2008. Coriander leaf mediated biosynthesis of gold nanoparticles, Materials Letters, 62, 4588.
- Philip D. 2010. Green synthesis of gold and silver nanoparticles using *Hibiscus rosa sinensis*. Physica E: Low-dimensional Systems and Nanostructures, 42, 5, 1417.
- Verma V., Karwar R. and Gange A. 2010. Biosynthesis of antimicrobial silver nanoparticles by the endophytic fungus Aspergillus clavatus. Nanomedicine (Lond), 5(1), 33.