



SEASONAL VARIATION OF PHYTOCHEMICALS

Rubaya Sultan^a, Neelofar Majid^b, Saduf Nissar^b and Aabid M. Rather^{*b}

^a Department of Botany, S. P. College Srinagar, 190001, J & K, India

^b Plant Reproductive Biology, Genetic Diversity and Phytochemistry Research Laboratory, Department of Botany, University of Kashmir, Srinagar, 190006, J&K, India

*Corresponding Author Email: abid.bot@gmail.com

ABSTRACT

Plant kingdom is a potential source of pharmaceutically active substances. Medicinal plants depict a tremendous variation in active constituents during different seasons; which have been widely attributed to variations in environmental variables such as temperature and rainfall. Aspects of synergistic effects due to UV radiation, humidity and hot temperatures can explain seasonal trends of plant secondary metabolite accumulation. Since active principles of some plants vary quantitatively in different seasons of the year so therapeutic efficacy of medicinal plants is also likely to vary during different seasons of the year and the majority of plant materials are usually best collected during season when the herbs are at peak maturity and concentration.

KEY WORDS

Active constituents, Medicinal plants, Seasons, Therapeutic efficacy, UV radiation

INTRODUCTION

The plant kingdom is a treasure house of potential drugs and in the recent years there has been an increasing awareness about the importance of medicinal plants. Medicinal plants are of great local significance and also global importance. In Ayurveda about 2000 plant species are labeled as a source of medicinal value, while, in Chinese Pharmacopoeia 5700 traditional medicines are listed (Chopra *et al.*, 1956), most of which are still used in conventional medical practice though the whole plants are rarely used (Todarwal *et al.*, 2011). Medicinal plants have been used in traditional treatments for numerous human diseases for thousands of years and they continue to be an important therapeutic aid for alleviating the ailments of human kind (Momin and Kadam, 2011). Among the 120 active compounds currently isolated from the higher plants and widely used in modern medicine, today 80% show a positive correlation between their modern therapeutic use and the traditional use of the plants from which they are

derived (Fabricant and Farnsworth, 2001; Pandith, 2012).

Drugs from the plants are easily available, less expensive, safe, and efficient and rarely have side effects (Yadav and Agarwala, 2011). Traditional knowledge of medicine has long been used since ages for curing various human ailments. About 60-80% of world populations still rely on plant-based medicines (Santhi *et al.*, 2011). Though the traditional Indian system of medicine has a long history of use, yet they lack adequate scientific documentation, particularly in light of modern scientific knowledge (Shrivastava and Leelavathi, 2010).

The medicinal value of plant lies in the bioactive phytochemical constituents of the plant showing various physiological effects on human body. So, through phytochemical screening one could detect the various important compounds which could be used as the base of modern drugs for curing various diseases (Sheikh *et al.*, 2013). Phytochemical screening of various plants has been reported by many workers (Mojab *et al.*,

2003; Parekh and Chanda, 2008). These studies have revealed the presence of numerous chemicals including alkaloids, flavonoids, steroids, phenols, glycosides and saponins.

Plants being valuable source of a wide range of secondary metabolites are used as pharmaceuticals, agrochemicals, flavors, fragrances, colors, biopesticides and food additives. Over 80% of the approximately 30,000 known natural products are of plant origin (Fowler and Scragg, 1988). Secondary metabolites perform no direct metabolic function but fulfill specific ecological functions such as maintaining aesthetics of the plant, attracting insects for pollen transfer and animals for consumption of fruits, as defense mechanism in conditions of stress, wounding or pathogen attack and as natural pesticides (Heldt, 2005). Despite advancements in synthetic chemistry, biological sources are usually preferred for a number of secondary metabolites including pharmaceuticals due to lesser side effects and better biodegradability (Pezzuto, 1995; Vimala *et al.*, 2014). Previously the crude drugs/extracts prepared from plants were identified by comparison only with the standard descriptions available in the literature, but recently due to advancement in the field of pharmacognosy, various techniques have been followed for the standardization of crude drugs. Phytochemical screening is one of the techniques to identify new sources of therapeutically and industrially important compounds like alkaloids, flavonoids, phenolics, steroids, tannins, saponins etc. present in the plant extracts. These compounds can be derived from any part of the plants like bark, leaves, flowers, seeds, etc.

Knowledge of the chemical constituents of plants is desirable because such information will be of value for the synthesis of new bioactive compound/s for treating the specific disease (Geetha and Geetha, 2014).

IMPACT OF SEASONS ON PHYTOCHEMICALS

Season has impact on availability of active principles in medicinal plants. According to principles of western herbal medicine, therapeutic efficacy varies during different times or seasons of the year. The constituents and active principles vary quantitatively at different seasons of the year and the majority of plant materials are usually best collected during season when the herbs are at peak maturity and concentration (Singh, 2008; Jayanthi *et al.*, 2013).

Biosynthesis of these secondary metabolites is not only controlled genetically, but is also strongly affected by different biotic and abiotic stresses (Naghdi *et al.*, 2004). Number of factors such as climate, altitude, rainfall and other conditions may affect growth of plants which in turn affect the quality of herbal ingredients present in a particular species even when it is produced in the same country. These conditions may produce major variations in the bioactive compounds present in the plants (Kokate *et al.*, 2004; Geetha and Geetha 2014).

Many studies on medicinal plants indicated that chemical content composition of these plants may vary substantially with the developmental stage of the plants. For this reason, investigations on ontogenetic variation of secondary metabolites from different classes have received considerable interest from plant scientists over several decades (Cirak *et al.*, 2007). The analytical determination of taxanes in extracts from the bark of *T. baccata* indicates that the taxane content depends significantly on the season of the year. It appears likely that the choice of the right season for harvesting plant material, along with optimization of other parameters (e.g. improved isolation techniques) can lead to the availability of taxanes in higher yields (Vesela, 1999).

Secondary metabolites are only produced in large quantities when they are needed, at a particular stage in lifecycle, at certain seasons or in those tissues that require most protection (Salminen *et al.*, 2001). Climatic conditions, such as time of day, precipitation and outside temperature, have a significant influence on the physical qualities, chemical composition of the medicinal plants. Sunshine duration, the average height of rainfall, average temperature and thermal amplitude between day and night also influence the physiological and biochemical activity of plants. It is important to predetermine all of these factors (Endrias, 2006). Studies have shown that in Brassicaceae concentrations and profiles of GLS show considerable variation within species and that they vary with environmental conditions and developmental stage (Poelman *et al.*, 2008; Hanson *et al.*, 2009).

Alkaloid content changes constantly in a plant throughout the growth period, the maximum stocks of alkaloids in leaves are accumulated before flowering (Maknickiene *et al.*, 2013). The average contents of vitamin C, glucosinolate, total phenol, and total flavonoid and antioxidant activities were significantly

higher in florets in the spring than in the fall in broccoli (Bhandari and Kwak, 2014).

The concentration of a particular phenolic compound within a plant tissue is dependent on season and may also vary at different stages of growth and development (Ozyigit *et al.*, 2007). Stefkov *et al.* (2009) reported in *Teucrium polium* that the content of total flavonoids are the highest in the period from May to July, which could be recommended as the most convenient period in the season for its collection. Contents of phenolics and flavonoids as well as anti-oxidant activity of daisy flowers vary to a relatively small extent during the year (Siatka and Kasparova, 2010).

According to Generalic *et al.* (2012), *Salvia officinalis* (extract) collected in the month of the May is richest in total flavonoids and showed the best anti-oxidant properties and the highest anti-microbial activity. Thus, collection of these plants during May seems the best choice for further use in the pharmaceutical industry. In *Melilotus indicus*, total flavonoid and phenolic content increases from colder to hotter seasons when plants receive stronger sunlight for a longer duration. Antioxidant activities and lipid peroxidation inhibitory activities of methanolic extracts and their fractions also showed variation with the seasonal change (Ahmed *et al.*, 2012). The active components in *Momordica charantia* fruit which are responsible for its anti-diabetic and hypolipidemic activity vary quantitatively and/or qualitatively during different seasons of the year and reach the peak during spring (Kolawole and Ayankunle, 2012).

Essential oils extracted from *Inga laurina* in the rainy season showed a better inhibition of the bacterial growth when compared to the oils of the dry season, particularly with respect to aerobic microorganisms (Furtado *et al.*, 2014). In *Microtus oeconomus* concentration of flavonoids, condensed tannins and total phenols being lowest in the month of June and gradually increasing from July to August/September (Dai *et al.*, 2014). Firdusi *et al.* (2015) observed that total alkaloids, flavonoids and phenolic compounds showed increasing value in monsoon than pre-monsoon in *Lasia spinosa*.

REFERENCES

- Ahmed D, Baig H and Zara S (2012). Seasonal variation of phenolics, flavonoids, antioxidant and lipid peroxidation inhibitory activity of methanolic extract of *Melilotus indicus* and its sub-fractions in different solvents. *International Journal of Phytomedicine*, 4: 326-332.
- Bhandari SR and Kwak J (2014). Seasonal variation in phytochemicals and antioxidant activities in different tissues of various *Broccoli* cultivars. *African Journal of Biotechnology*, 13: 604-615.
- Chopra RN, Nayar SL and Chopra IC (1956). *Glossary of Indian Medicinal Plants*, CSIR, New Delhi, India, 152-162.
- Cirak C, Radusiene J, Ivanauskas L and Janulis (2007). Variation of bioactive secondary metabolites in *Hypericum origanifolium* during its phenological cycle. *Acta Physiologiae Plantarum*, 29: 197 – 203.
- Dai X, Han M, Liu Q, Shang G, Yin B, Wang A, Dean BE, Wei W and Yang S (2014). Seasonal changes in the concentrations of plant secondary metabolites and their effects on food selection by *Microtus oeconomus*. *Mammalian Biology-Zeitschrift fur Säugetierkunde*, 79: 215-220.
- Endrias A (2006). Bio-raffinage des plantes aromatiques et médicinales appliquent à l'*Hibiscus sabdariffa* L. et à l'*Artemisia annua*. École doctorale science des procédés. Spécialité sciences des Agroressource. Docteur de l'institut national polytechnique. Toulouse, 15p.
- Firdusi SN, Neog N and Barooah MS (2015). Seasonal variation in biochemical contents of *Lasia spinosa*. *Medicinal Plants-International Journal of Phytomedicines and Related Industries*, 7: 55-59.
- Fowler MW and Scragg AH (1988). Natural products from higher plants and plant cell culture. In: Pais MSS, Mavituna F, Novais JM, editors. *Plant cell biotechnology*. NATO ASI Series. Berlin: Springer-Verlag, 18: 165-77.
- Furtado FB, de Aquino FJT, Nascimento EA, Martins Cde M, de Moraes SAL, Chang R, Cunha LCS, Leandro LF, Martins CHG, Martins MM, da Silva CV, Machado FC and de Oliveira A (2014). Seasonal variation of the chemical composition and antimicrobial and cytotoxic activities of the essential oils from *Inga laurina* (Sw.) Willd. *Molecules*, 19: 4560-4577.
- Geetha TS and Geetha N (2014). Phytochemical Screening, Quantitative Analysis of Primary and Secondary Metabolites of *Cymbopogon citratus* (DC) stapf. leaves from Kodaikanal hills, Tamilnadu. *International Journal of Pharm Tech Research*, 6: 521-529.
- Generalic I, Skroza, D, Surjaka J, Mozina SS, Ljubenkovic I, Katalinic A, Simate V and Katalinic V (2012). Seasonal variations of phenolic compounds and biological properties in Sage (*Salvia officinalis* L.). *Chemistry & Biodiversity*, 9: 441-457.
- Hanson P, Yang R and Chang L (2009). Contents of carotenoids, ascorbic acid, minerals and total glucosinolates in leafy brassica pakchoi (*Brassica rapa* L. *chinensis*)

- as affected by season and variety. *Journal of the Science of Food and Agriculture*, 89: 906–914.
- Heldt HW and Heldt F (2005). *Plant biochemistry* 3rd ed. USA: Academic Press.
- Jayanthy A, Upadhaya PK and Remashree AB (2013). Seasonal and Geographical Variations in Cellular Characters and Chemical Contents in *Desmodium gangeticum* (L.) DC. – An Ayurvedic Medicinal Plant. *International Journal of Herbal Medicine*, 1: 34-37.
- Kokate CK, Purohit AP and Gokhale SB (2004). *Practical Pharmacognosy* 2nd edition. Vallabh Prakashan, New Delhi, pp. 466-470.
- Kolawole OT and Ayankunle AA (2012). Seasonal variation in the anti-diabetic and hypolipidemic effects of *Momordica charantia* fruit extract in rats. *European Journal of Medicinal Plants*, 2: 177-185.
- Maknickiene Z, Asakaviciute R, Baksiene E and Razukas A (2013). Alkaloid Content Variations in *Lupinus luteus* L. and *Lupinus angustifolius* L. *Archives of Biological Science Belgrade*, 65: 107-112.
- Mojab F, Kamalinejad M, Ghaderi N, and Vahidipour HR (2003). Phytochemical screening of some species of Iranian plants. *Iranian Journal of Pharmaceutical Research*, 2: 77-82.
- Momin RK and Kadam VB (2011). Determination of ash values of some medicinal plants of genus sesbania of marathwada region in maharashtra. *Journal of Phytology* 3: 52-54.
- Naghdi BH, Yazdani D, Mohammad AS and Nazari F (2004). Effects of spacing and harvesting on herbage yield and quality\quantity of oil in thyme. *Industrial Crops and Products*, 19: 231-236.
- Ozyigit II, Kahraman MV and Ercan O (2007). Relation between explant age, total phenols and regeneration response of tissue cultured cotton (*Gossypium hirsutum* L.). *African Journal of Biotechnology*, 6: 3-8.
- Pandith JI (2012). Phytochemical Screening of certain plant species of Agra city. *Journal of Drug Delivery & Therapeutics*, 2: 135-138.
- Parekh J and Chanda SV (2007). In vitro antimicrobial activity and phytochemical analysis of some Indian medicinal plant. *Turkish Journal of Biology*, 31: 53-58.
- Pezzuto JM (1995). Natural product cancer chemoprotective agents. In: Arnason J. T., Mata, R., Romeo, J. T. (eds.). Recent advances in phytochemistry. *Phytochemistry of Medicinal Plants*. New York: Plenum Press, 29: 19-45.
- Poelman EH, Galiart RJ, Raaijmakers CE, Van Loon JJ and Van Dam NM (2008). Performance of specialist and generalist herbivores feeding on cabbage cultivars is not explained by glucosinolate profiles. *Entomologia Experimentalis et Applicata*, 127: 218-228.
- Salminen JP, Ossipov V, Haukioja E and Pihlaja K (2001). Seasonal variation in the content of hydrolysable tannins in leaves of *betula pubescens*. *Phytochemistry*, 57: 15-22.
- Santhi R, Lakshmi G, Priyadarshini AM and Anandaraj L (2011). Phytochemical screening of Nerium oleander leaves and Momordica Charantia leaves. *International Research Journal of Pharmacy*, 2: 131-135.
- Sheikh N, Kumar Y, Misra AK and Pfoze L (2013). Phytochemical screening to validate the ethnobotanical importance of root tubers of *Dioscorea* species of Meghalaya, North East India. *Journal of Medicinal Plants Studies*, 1: 62-69.
- Shrivastava S and Leelavathi S (2010). Preliminary Phytochemical evaluation of leaf extracts of *Catunaregum spinosa* Thumb. *International Journal of Pharmaceutical Sciences Review and Research*, 3: 114-118.
- Siatka T and Kasparova M (2010). Seasonal variation in total phenolic and flavonoid contents and DPPH scavenging activity of *Bellis perennis* L. flowers. *Molecules*, 15: 9450-9461.
- Singh A (2008). A note on variation of active principles in Indian Medicinal Plants and TIM formulations, *Ethnobotanical Leaflets*, 12: 603- 606.
- Stefkov G, Karapandzova M, Stefova M and Kulevanova S (2009). Seasonal variation of flavonoids in *Teucrium polium* L. (Lamiaceae). *Macedonian Pharmaceutical Bulletin*, 55: 33-40.
- Todarwal A, Jain P and Bari S (2011). *Abelmoschus manihot* Linn: ethnobotany, phytochemistry and pharmacology / *Asian Journal of Traditional Medicines*, 6: 1-7.
- Vesela D, Saman D, Valterova I and Vanek T (1999). Seasonal Variations in the Content of Taxanes in the bark of *Taxus baccata* L. *Phytochemical Analysis*, 10: 319–321.
- Vimala V, Mathew R, Sankar PD and Kalaivani T (2014). Phytochemical analysis in *Ocimum* Accessions. *International Journal of Pharmacy and Pharmaceutical Science*, 6: 555-557.

Received:06.05.18, Accepted: 08.06.18, Published:01.07.2018

*Corresponding Author:

Aabid M. Rather*

Email: abid.bot@gmail.com