

EMERGING INTERVENTIONS IN THE MANAGEMENT OF NEUROPATHIC PAIN

Shaifali Chowdhary¹, A.C Rana¹, Ramica Sharma¹, Shaveta Gangwani¹ Rayat Institute of Pharmacy, SBS Nagar, Ropar-144533, India

*Corresponding Author Email: Shaifali87@gmail.com

RECEIVED ON 30-08-2011

Review Article
ACCEPTED ON 23-09-2011

ABSTRACT

Neuropathic pain (NP) may arise as a consequence of a lesion or disease affecting the somatosensory system. It is one of the most incapacitating reasons for pain disorder in large number of population worldwide. The hallmark signs of NP are chronic allodynia and hyperalgesia. Various analgesics like opoids and NSAIDS lack antinociceptive effect in the management of NP but they are associated with various adverse effects and resistance. Hence the present review delineated with the recent targets for the management of NP along with their signalling pathways.

KEYWORDS: neuropathic pain, trauma, inflammation.

Introduction

Neuropathic pain (NP) is referred as "most terrible of all nerve tortures" [1], initiated by a primary lesion to somatosensory nervous system that involves an element of sensory dysfunction.^[2] It was estimated that about 37.6 million individuals suffered from NP across the seven major markets in 2005 and it is forecasted that its prevalence will increase to 39.1 million individuals by 2011. [3] Various studies indicate that 95% of patients with spinal cord injuries have NP. [4] The symptoms of NP includes burning, cramping, electric shock, numbness, shooting and stabbing. [5] NP characterized by disproportionate hypersensitivity to stimuli (hyperalgesia), abnormal pins and-needles or electric-shocklike sensations (hyperpathia) and finally nociceptive responses to non-noxious stimuli (allodynia). [6] Various factors are implicated in the pathogenesis and progression of NP like vascular lesions, avulsion injuries, traumatic injury, tumors^[7] and diabetes.^[8] Among all of them allodynia (pain perception in response to normally non painful stimuli) and hyperalgesia (exaggerated pain sensation to normally

painful stimuli) are regarded as the major symptoms of NP.[1] It has been well that during nerve documented injury numerous mediators are found to be up regulated. [9] Mast cells are found to play key role in the release of various inflammatory mediators [10] like are cytokines such as [interleukin-1beta (IL-1β), interleukin-6 (IL-6), factor-alpha tumor necrosis $(TNF\alpha)$], prostaglandin E2 (PGE2) and nitric oxide (NO). [9] During nerve injury, microglial cells causes up regulation of adenosine triphosphate (ATP) receptors (P2X4and P2X7) $^{[11]}$, chemokine receptor (CX3CR1) $^{[12]}$, TNF $\alpha^{[13]}$, IL-1 β and IL-6 $^{[14]}$ which mediate their effect via activation of mitogen-activated protein kinase (p38MAPK)^[15] and contributes to NP. [16] Moreover, it has been reported that NO which is a potent vasodilator too is responsible for the progression of NP^[17,18] via causing increase in the expression of inducible nitric oxide synthase (iNOS).[19,20] Further, during NP there is increased generation of reactive oxygen species (ROS) via activation of

International Journal of Pharmacy and Biological Sciences (eISSN: 2230-7605)



NADPH oxidase-2 (Nox2), which further plays a critical role in NP.^[21]

Biology of NP

As discussed earlier that NP may arise as a consequence of a lesions or disease affecting the somatosensory system. [22] factors are responsible in the progression of NP like trauma, metabolic abnormalities, chemotherapy, surgery, irradiation, neurotoxins, inherited neurodegeneration, nerve compression and inflammation. [23] NP is characterised by hyperalgesia (Provoked by heat stimulation) and chronic allodynia (Pain due to stimulus that does not normally provoke pain, it can be provoked by touch stimulation or cooling). [19,5] NP is a complex entity, it can be classified into various types such as trigeminal neuralgia, diabetic neuropathy (DPN) and postherpetic neuralgia (PHN). [23] Nerve injury in NP initiates either by central or peripheral mechanisms. [24] Around the site of injury, inflammation occurs, due to degranulation^[25,26] cell mast neutrophils^[27] of proliferation and macrophages^[28] which has been reported to trigger the release of various proinflammatory mediators such as histamine, cytokines and proteases. [29,30] In peripheral mechanisms, the pain arises due to noxious stimuli at nociceptive C-fibers and non nociceptive Aδfibers neurons.[31] These neurons become abnormally sensitive due to development of pathological spontaneous activity proinflammatory mediators.[32] Furthermore, there is increased expression of messenger RNA (mRNA) for sodium channel (Na⁺) present at these neurons^[33,34] and contribute to action potential threshold and hyperactivity. [35] Moreover, in peripheral nerve injury the nonneural glial cells too play a pivotal role and they have been reported to release various cytokines and excitatory amino acids (EAA)[36] EAA by acting on N-methyl-D-aspartate (NMDA) receptors and neuropeptide

substance P produces symptoms associated with NP. [37]

In central mechanism, microglial cells causes upregulation of ATP receptors and chemokine receptor^[38,39,40,12], which are elicited in progression of NP. TNF-α activates tumour necrosis factor receptor 1 (TNFR1) and TNFR2 receptors, which are upregulated during inflammation^[6] which further activates Na⁺, thus increases membrane potassium (K⁺) ion conductance which lead to overall neuronal hyperexcitability and causes NP. [13] IL-1β and IL-6 are too implicated in progression of NP by activating tyrosine kinases (TKs) and protein kinase C (PKC). [41,14] Further, various evidences indicates that during NP there is increased expression of Inos^[42] that causes the activation of cyclooxygenase (COX) and results in the marked increase in the level of prostaglandin E2 (PGE₂), which plays critical role in NP^[43] Moreover, peripheral nerve injury results in increase generation of ROS production via activation of Nox2, which further causes activation of spinal glial cells and cytokines. [21,19] proinflammatory Various evidences indicates that there is upregulation in activity of p38 MAPK which in return is responsible for activating various transcription factor like NF-κB, which lead to pain hypersensitivity^[44] Figure 1 indicates the pathogenesis of NP initiated by nerve injury.

Thus, by knowing various signalling mechanisms responsible in the progression of NP, various new safer, efficacious therapeutic interventions can be designed. Hence, the review deals with emerging drugs used in the management of neuropathic pain.

Emerging drugs management in NP

Capsaicin patches

Capsaicin is an agonist of the TRPV1 receptor and activates TRPV1 ligand-gated channels on nociceptive fibers results in depolariation and thus initiate action potential (AP). [45] The increase AP causes transmission of pain signals to the spinal cord. [46] Capsaicin at high doses deactivates the mechanism of vanilloid receptor. [47] Capsaicin containing creams are effective in PHN. Recently, the efficacy of a one-off application of high concentration (8%)

capsaicin patch (30, 60 or 120 min) applied to the painful area compared to low concentration (0.04%) has been demonstrated from weeks 2 to12 in PHN neuropathy. [48] Adverse effects include local capsaicin-related reactions at the application site pain, erythema, sometimes edema, and itching. [49,50]

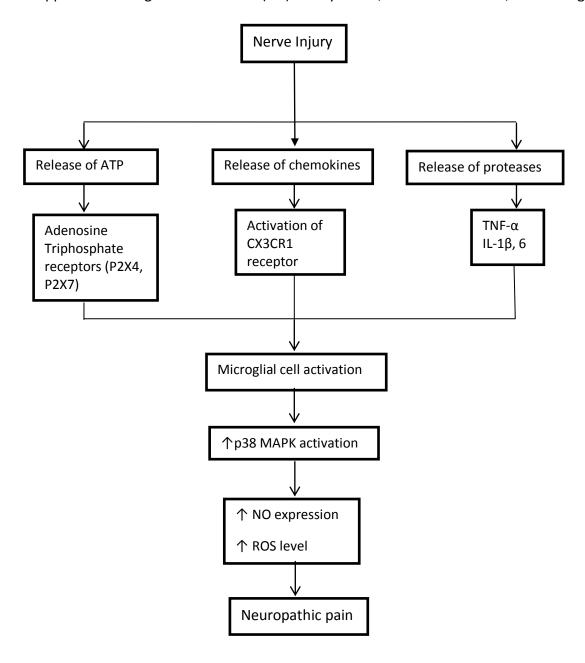


Figure 1: Pathophysiology of neuropathic pain occurs as a result of nerve injury.



Botulinum toxin A (BTX-A)

BTX-A, a potent neurotoxin used for the treatment of focal muscle hyperactivity, by acting on neurogenic inflammation. [51] Clinically subcutaneous injection of BTX-A is used in management of allodynia and diabetic polyneuropathy. [51,52] The drug has an excellent safety profile with no systemic side effects and only pain during injection.

Cannabinoids

Cannabinoids have therapeutic potential in chronic pain following the discovery of cannabinoid receptors and their endogenous ligands. [45] Cannabinoids acts as agonist TRPV1 receptor, leads to desensitization analgesia.[53] consequent behavioral Cannabinoids via acting on TRPV1 receptor channel help in increasing pain threshold. [54,55] Clinically, cannabinoids (2.7 mg delta-9tetrahydrocannabinol/ 2.5 mg cannabidiol) administered by oromucosal route effective in pain associated with multiple sclerosis^[56] and in refractory peripheral NP associated with allodynia [57]. Adverse reactions include dizziness, dry mouth, sedation, fatigue, gastrointestinal effects, and discomfort. [56,57] In long-term studies oromucosal cannabinoids, no tolerance was observed, but 90% of the patients experienced adverse reactions.[56]

NMDA antagonists

NMDA is a receptor for the excitatory neurotransmitter glutamate, which is released with noxious peripheral stimuli. The activation of NMDA receptors has been associated with hyperalgesia, neuropathic pain, thus results in increased spinal neuron sensitization, leading to neuropathic pain. Therefore, NMDA antagonist may have a role in these areas of pain management. [60]

Ketamine is a strong NMDA antagonist and is effective at doses of 2-4 mg/kg, *i.m.* and 0.2-0.75 mg/kg, *i.v.*^[61] Other antagonists include memantine, amantadine (200 mg, infused over 3 hr, *i.v.*), dextromethorphan (400mg/d) and methadone (2.5-10 mg, oral dose). [62,61]

TNF-α antagonists

TNF- α have a pivotal role in the genesis of mechanical allodynia and thermal hyperalgesia during inflammatory and neuropathic pain. [63] Etanercept is a potent antagonist of TNF- α . [64] It reduces tissue injury and improves motor recovery by enhanced axonal regeneration. [65] It is effective when i.p. injection 1 hr before and 6 hr after injury is given. [66] Preclinical studies demonstrated that aqueous extract of *Mangifera indica* L. act as antagonist of TNF- α . [3]

Conclusion

As NP is regarded as one of the major disorder associated with Hyperalgesia and Allodynia. But no evident management is available for this deadly disorder. Hence the present review opens the vista for the management of neuropathic pain via newer emerging drugs.

Acknowledgement

I express my thanks to Rayat and Bahra group and to the Chairman of the college S. Gurvinder Singh Bahra and S. Nirmal Singh Rayat for providing supportive and friendly atmosphere and excellent facilities in and around the region

References

[1] Kaur G., Jaggi A.S., Singh N., Exploring the potential effect of Ocimum santum in vincristine-induced neuropathic pain in rats. J BPPNI, 5:1-9, (2010).



- [2] Costigan M., Scholz J., Woolf C.J., Neuropathic Pain: A Maladaptive Response of the Nervous System to Damage. Ann Rev Neurosci, 32:1-32, (2009).
- [3] Garrido-Suarez B.B., Garrido G., Delgado R., Bosch F., Rabi M.D.C., A *Mangifera indica* L. Extract Could Be Used to Treat Neuropathic Pain and Implication of Mangiferin. Molecules, 15: 9035-9045, (2010).
- [4] Siniscalco D., Novellis V.D., Rossi F., Maione S., Neuropathic Pain: Is the End of Suffering Starting in the Gene Therapy?. Curr Dru Targ, 6: 75-80, (2005).
- [5] Beggs S. and Salter M.W., Neuropathic Pain: Symptoms, Models, and Mechanisms. Dru Dev Res, 67:287-301, (2006).
- [6] Leung L. and Cahill CM., TNF- α and neuropathic pain- a review. J Neuroinflam, 7:1-11, (2010).
- [7] Saxena A.K, and Azad R., Advances in the Mechanisms, Diagnosis and Management Of Neuropathic Pain: Current Opinions And Future Perspectives. Indian J Anaesth, 50:249-257, (2006).
- [8] Nagata K., Imai T., Yamashita T., Tsuda M., Tozaki-Saitoh H., Inoue K., Antidepressants inhibit P2X₄ receptor function: a possible involvement in neuropathic pain relief. Mole Pain, 5:1-12, (2009).
- [9] Thacker M.A., Clark A.K., McMahon S.B., Pathophysiology of peripheral Neuropathic Pain: Immune Cells and Molecules. Anesth Analg, 105:838-47, (2007).
- [10] Theoharides T.C. and Cochrane D.E., Critical role of mast cells in inflammatory diseases and the effect of acute stress. J Neuroimmunol, 146:1-12, (2004).

- [11] Tsuda M., Inoue K., Neuropathic pain and ATP receptors in spinal microglia. Brain Nerv, 59:953-959, (2007).
- [12] Abbadie C., Bhangoo S., Koninck Y.D., Malcangio M., Melik-Parsadaniantz S., White FA., Chemokines and pain mechanisms. Brain Res Rev, 60:125-34, (2008).
- [13] Jin X. and Gereau R.W., Acute p38-mediated modulation of tetradotoxin-resistant sodium channels in mouse sensory neurons by tumour necrosis factor-alpha. J Neurosci, 26:246-255, (2006).
- [14] Oliveira C.M.B., Sakata R.K., Issy A.M., Gerola L.R., Salomao R., Cytokines and Pain., Rev Bras Anestesiol, 61:255-265, (2011).
- [15] Baron R., Mechanism of Disease: neuropathic pain- a clinical perspective. Nat Clin Pract Neurol, 2:95-106, (2006).
- [16] Ji R.R. and Suter M.R., p38 MAPK, microglial signalling, and neuropathic pain. Mole Pain, 3:1-9, (2007).
- [17] Hokfelt T., et al., Messenger Plasticity in Primary Sensory Neurons Following Axotomy and Its Functional implications. Trends Neurosci, 17:22-30, (1994).
- [18] Noguchi K., et al., Axotomy Induced Preprotachykinin Gene Expression In A Subpopulation Of Dorsal Root Ganglion Neurons. J Neurosci Res, 37:596-603, (1994).
- [19] Naik A.K., Tandan S.K., Kumar D., Dudhgaonkar S.P., Nitric oxide and its modulators in chronic constriction injury-induced neuropathic pain in rats. Eur J Pharmacol, 530:59-69, (2006).
- [20] Levy D. and Zochodne D.W., NO Pain: Potential Roles of Nitric Oxide in Neuropathic Pain. Pain Pract, 4:11-18, (2004).



- [21] Kim D., You B., Jo E.K., Han S.K., Simon M.I., Lee S.J., NADPH oxidase 2-derived reactive oxygen species in spinal cord microglia contribute to peripheral nerve injury-induced neuropathic pain. PNAS, 1-6, (2010).
- [22] Treede R.D., Jensen T.S., Campbell J.N., et al., Neuropathic pain: redefinition and a grading system for clinical and research purposes. Neurology, 70:1630-1635, (2008).
- [23] Dworkin R.H, Backonja M., Rowbotham M.C., Allen R.R, Argoff C.R, et al., Advances in Neuropathic Pain, Diagnosis, Mechanisms and Treatment Recomendations. Arch Neurol, 60:1524-1534, (2003).
- [24] Ji R.R. and Wen Y.R., Neural-Glial Interaction in the Spinal Cord for the Development and Maintenance of Nerve Injury-Induced Neuropathic Pain. Dru Dev Res, 67:331-338, (2006).
- [25] Zuo Y., Perkins N.M., Tracey D.J., Geczy C.L., Inflammation and hyperalgesia induced by nerve injury in the rat: a key role of mast cells. Pain, 105:467-79, (2003).
- [26] Olsson Y., Degranulation of mast cells in peripheral nerve injuries. Acta Neurol Scand, 43:365-374, (1967).
- [27] Daemen M.A., Kurvers H.A., Kitslaar P.J., Slaaf D.W., Bullens P.H., et al., Neurogenic inflammation in an animal model of neuropathic pain. Neurol Res, 20:41-45, (1998).
- [28] Perry V.H., Brown H.C., Gordon S., The macrophage response to central and peripheral nerve injury. A possible role for macrophages in regeneration. J Exp Med, 165:1218-1223, (1987).
- [29] Galli S.J., Nakae S., Tsai M., Mast cells in the development of adaptive immune responses. Nat Immunol, 6:135-42, (2005).

- [30] Metcalfe D.D., Baram D., Mekori Y.A., Mast cells. Physiol Rev, 77:1033-79, (1997).
- [31] Baron R., Mechanism of Disease: neuropathic pain- a clinical perspective. Nat Clin Pract Neurol, 2:95-106, (2006).
- [32] Marchand F., et al., Role of the immune system in chronic pain. Nat Rev Neurosci, 6:521-532, (2005).
- [33] Ekberg J. and Adams D.J, Neuronal voltage-gated Sodium Channel Subtype Key roles in inflammatory and neuropathic pain. International J Biochem Cell Bio, 38:2005-2010, (2006).
- [34] Scadding J., Neuropathic pain. ACNR, 3:1-4, (2003).
- [35] Lai J., Gold M.S., Kim C.S., Bian D., Ossipov M.H., Hunter J.C., et al., Inhibition of neuropathic pain by decreased expression of the tetrodotoxin-resistant sodium channel, Nav1.8. Pain, 95:143-152, (2002).
- [36] Smith HS., Arachidonic Acid Pathways in Nociception. J Support Oncol, 4:277-287, (2006).
- [37] Luo Z.D., et al., Upregulation of dorsal root ganglion $\alpha 2\delta$ calcium channel subunit and its correlation with allodynia in spinal nerveinjured rats. J Neurosci, 21:1868-1875, (2001).
- [38] Tsuda M., Shigemoto-Mogami Y., Koizumi S., Mizokoshi A., Kohsaka S., Salter M., Inoue K., P2X4 receptors induced in spinal microglia gate tactile allodynia after nerve injury. Nature, 424:778-783, (2003).
- [39] Suzuki T., Hide I., Ido K., Kohsaka S., Inoue K., Nakata Y., Production and release of neuroprotective tumor necrosis factor by P2X7 receptor activated microglia. J Neurosci, 24:1-7, (2004).



- [40] White F.A., Jung H., Miller R.J., Chemokines and the pathophysiology of neuropathic pain. Proc Natl Acad Sci USA, 104:20151-20158, (2007)
- [41] McMahon S.B. and Malcangio M., Current challenges in glia Pain Biology. Neuron, 64:46-54, (2009).
- [42] Omoigui S., The biochemical origin of pain Proposing a new law of pain: The origin of all pain is inflammation and the inflammatory response. Part 1 of 3-A unifying law of pain. Med Hypotheses, 1-13, (2007).
- [43] Yang H.Y., Chang H.K, Lee J.W., Kim Y.S., Kim H., Lee M.H., Shin M.S., Ham D.H., Park H.K., Lee H., Kim C.J., Amygdalin suppressed lipopolysaccharide-induced expressions of cyclooxygenase-2 and inducible nitric oxide synthase in mouse BV2 microglial cells. Neurol Res, 29:S59-S64, (2007).
- [44] Raghavendra V. and DeLeo., The role of astrocytes and microglia in persistent pain. Adv Mole Cell Bio, 31:951-966, (2004).
- [45] Attall N. and Martinez V., Recent Development in the Pharmacological Management of Neuropathic Pain. Eur Neurological J, 1-6, (2010).
- [46] Wong G.Y. and Gavva N.R., Therapeutic potential of vanilloid receptor TRPV1 agonists and antagonists as analgesics: recent advances and setbacks. Brain Res Rev, 60:267-277, (2009).
- [47] Kova H.K. and Gallo M.P., Adjuvant Analgesics. Anesthesiolo Clin, 25:775-786, (2007).
- [48] Noto C., Pappagallo M., Szallasi A., NGX-4010, a high-concentration capsaicin dermal patch for lasting relief of peripheral neuropathic pain. Curr Opin Investig Drugs, 10:702-710, (2009).

- [49] Simpson D.M., Schifitto G., Clifford D.B., Murphy T.K., Cruz E.D.D., Glue P., Whalen E., Emir B., Scott G.N., Freeman R., 1066 HIV Neuropathy Study Group. Pregabalin for painful HIV neuropathy: a randomized, doubleblind, placebo-controlled trial. Neurology, 2:413-420, (2010).
- [50] Backonja M., Wallace M.S., Blonsky E.R., et al., NGX-4010, a high concentration capsaicin patch, for the treatment of postherpetic neuralgia: a randomised, double-blind study. Lancet Neurol, 7:1106-1102, (2009).
- [51] Ranoux D., Attal N., Morain F., Bouhassira D., Botulinum toxin a induces direct analgesic effects in neuropathic pain: a double blind placebo controlled study. Ann Neurol, 64:274-283, (2008).
- [52] Yuan R.Y., Sheu J.J., Yu J.M., et al., Botulinum toxin for diabetic neuropathic pain: a randomized double-blind crossover trial. Neurology, 72:1473-1478, (2009).
- [53] Palazzo E., Luongo L., Novellis V.D., Rossi F., Maione S., The Role of Cannabinoid Receptor in the Descending Modulation of Pain. Pharmaceuticals, 3:2661-2673, (2010).
- [54] Akopian A.N., Ruparel N.B., Jeske N.A., Patwardhan A., Hargreaves K.M., Role of ionotropic cannabinoid receptors in peripheral antinociception and antihyperalgesia. Trends Pharmacol Sci, 30:79-84, (2009).
- [55] Jordt S.E., Bautista D.M., Chuang H.H, McKemy D.D., Zygmunt P.M., Hogestatt E.D., Meng I.D, Julius D., Mustard oils and cannabinoids excite sensory nerve fibres through the TRP channel ANKTM1. Nature, 427:260-265, (2004).
- [56] Rog D.J., Nurmikko T.J., Young C.A., Oromucosal delta9-tetrahydro cannabinol cannabidiol for neuropathic pain associated with multiple sclerosis: an uncontrolled, open-



label, 2-year extension trial. Clin Ther, 29:2068-2079, (2007).

[57] Nurmikko T.J, Serpell M.G., Hoggart B., Toomey P.J., Morlion B.J., Haines D., Sativex successfully treats neuropathic pain characterised by allodynia: a randomised, double-blind, placebo-controlled clinical trial. Pain, 133:210-220, (2007).

[58] Chizh B.A. and Headley P.M., NMDA antagonists and neuropathic pain - multiple drug targets and multiple uses. *Curr Pharm Des*, 11:2977-2994, (2005).

[59] Bennett G.J., Update on the neurophysiology of pain transmission and modulation: focus on the NMDA-receptor. *J Pain Sympt Manage*, 19:S2-S6, (2000).

[60] DuPen A., Shen D., Ersek M., Mechanisms of opioid-induced tolerance and hyperalgesia. *Pain Manage Nursi*, 8:113-121, (2007).

[61] Jamero D., Borghol A., Vo N., Hawawini F., The Emerging Role of NMDA Antagonists in Pain Management. US Pharm, 36:HS4-HS8, (2011).

[62] Lucas L.K and Lipman A.G., Recent Advances in Pharmacotherapy for Cancer Pain Management. Can Pract, 10:S14-S20, (2002).

[63] Homma Y., Brule SJ., Zhang J.M., A comparison of chronic pain behavior following local application of tumor necrosis factor α to the normal and mechanically compressed lumbai ganglia in the rat. Pain, 95:239-246, (2002).

[64] Tracey D., Klareskog L., Sasso E.H., Saefeld J.G., Tak P.P., Tumor necrosis factor antagonist mechanisms of action: a comprehensive review. Pharmacol Ther, 117:244-279, (2008).

[65] Kato K., Kikuchi S., Shubayeb V.I, Myers R.R., Distribution and Tumor necrosis factor α isoform binding specificity of locally administered Etanercept into injured and uninjured rat sciatic nerve. Neurosci, 160:492-500, (2009).

[66] Tobinick E., Peripheral etanercept: a new therapeutic paradigm in neurology. Expert Rev Neurother, 10:985-1002, (2010).



*Address for the Correspondence:

Shaifali Chowdhary*

Rayat institute of pharmacy, Railmajra (Ropar), Punjab Contact no. 09780200799

Email: Shaifali87@gmail.com

 $98^{\rm egg}$