

International Journal of Pharmacy and Biological Sciences-IJPBS™ (2025) 15 (2): 39-53
Online ISSN: 2230-7605, Print ISSN: 2321-3272

Review Article | Biological Sciences | OA Journal | MCI Approved | Index Copernicus

A Comprehensive Review on Ayurvedic and Pharmacological Aspects of *Picrorhiza kurroa* Royle ex Benth

Varesh Parihar^{1*} and Banshidhar Behera²

¹PG Scholar (Ayu), Department of Dravyaguna Vigyan, A and U Tibbia College & Hospital Karol Bagh, New Delhi-110005

²Associate Professor (Ayu), Department of Dravyaguna vigyan, A &U Tibbia College and Hospital Karol Bagh, New Delhi-110005

Received: 26 Jan 2025 / Accepted: 20 Mar 2025/ Published online: 01 April 2025 *Corresponding Author Email: vareshparihar@gmail.com

Abstract

Picrorhiza kurroa Royal ex Benth, Kutki (P. kurroa) is an important medicinal plant, which has been traditionally recommended and used in Ayurveda for thousands of years with some precautions. The last few decades have witnessed a significant revival of interest in its pharmacology, pharmacognosy and phytochemistry. Kutki is a small perennial herb and rapidly decaying high-value medicinal plant that grows in the north-western Himalayan region of India, Pakistan and hilly parts of Nepal. The leaf, bark and underground parts of the plant, mainly the rhizome, are used in Ayurveda for their medicinal properties. Evidence of its hepatoprotective activity in several experimental and clinical studies has led to the correlation of specific phytochemical components of P. kurroa (e.g., iridoid glycosides, especially picrosides) with precise pharmacological activities, and the efficacy of P. kurroa in many diseases is done. Kutki is mainly used for liver disorders like jaundice as it protects the liver from cell damage caused by free radicals due to its antioxidant and hepatoprotective properties. The present review study is an attempt to provide detailed information about traditional uses, phytochemistry and pharmacological potential of P. kurroa, an important medicinal herb of the western Himalayas.

Keywords

Picrorhiza kurroa, Kutki, Katuka, Pharmacological activities

1 INTRODUCTION

The word "Picrorhiza" comes from the Greek word "Picros," which means "bitter," and "rhiza," which means "roots." The genus Picrorhiza, which belongs to the Plantaginaceae family, contains two species: Picrorhiza kurroa Royle ex Benth and Picrorhiza scrophulariiflora Pennell. The majority of species can be found in natural settings like cliffs, mountainside ledges, and crevices. The primary habitat of Picrorhiza kurroa Royle ex Benth is at an elevation of 3000 and 5000 m in the northwestern Himalayan region of India Pakistan and the mountainous areas of Nepal [1]. This plant grows in the Himalayan region (Garhwal to Bhutan) on moist, organic soils and rocky slopes [2]. In Ayurveda, the leaf, bark, and

underground parts of the plant, particularly the rhizome, are utilised for their therapeutic properties. *Picrorhiza kurroa* Royle ex Benth (Family: Plantaginaceae, previously classified as *Scrophulareaceae*) is a highly regarded medicinal herb, commonly referred to as "*Kutki*," "Kurro," or 'Indian gentian. The perennial herb *kutki* has a long history of use in the Indian ayurvedic system to treat respiratory and liver disorders.

The active compounds in *kutki* include kutkoside and iridoid glycosides (IGs), specifically picroside I and II, and they are mainly located in the rhizome and roots. No significant adverse effects have been noted for this plant. The various pharmacological properties of



P.kurroa, an endangered plant of medicinal importance, are noteworthy.

The aim of this review article was to assess the pharmacological effects of P. kurroa on various disorders. P. kurroa could serve as a natural substitute for conventional treatments aimed at addressing emerging diseases. This study is designed to serve as a reference for upcoming basic and clinical research. It is considered valuable due to its pharmacological effects, which include being hepatoprotective [3], anti-inflammatory immunomodulatory [5], antimicrobial [6], antitumor [7], anti-carcinogenic [8], antioxidant [9], antineoplastic [10], anti-arthritic [11], anti-leishmanial [12], anti-asthmatic [13], anti-diabetic [14], antifungal [15] and cardioprotective [16]. It is also helpful for gastrointestinal and urinary disorders, scorpion stings, snake bites, leukoderma and inflammatory conditions.

2 Vernacular Name

Hindi : Kutki English : Hellebore

Sanskrit : Kandaruha, Matsyashakala, Katurohini,

Chakrangi, Rohini, Tikta, Katumbhara

Gujrati : Kadu, Katu
Marathi : Kutki, Kalikutki
Punjabi : Karru, kaur
Assamese : Katki, Kutki

Oriya: Katuki

Tamil: Katuka rohini, Katuku rohini, Kadugurohini

Telugu: Karukarohini

Urdu: Kutki

'Kutki is used by various synonyms in classical literature, such as in Bhavprakash Nighantu. These synonyms describe biological activity or typical traits, i.e Kandaruha (The plant can grow from stem cuttings), Chakrangi (When cut, the stem appears like a wheel), Matsyashakala (Rhizome resembles fish scales), Ashoka (Relieves grief), Rohini (Protects organs with regenerative ability), Katurohini (Has regenerative action), Krushnabheda (Can cause black stools), Tikta (Has a very bitter taste) Katumbhara (Full of bitter principles) Matsyapitta (Bitter taste like fish bile and similar action). To demonstrate some biological effects and clinical uses associated with Kutki that pertain to this review are Bhedaneeya (promoting excretion / downwords expelled of solid or liquid purisha), Deepaniya (facilitate appetite), Rechani (laxative action), Aruchihara (improving Yakrutdalyodar-nashak appetite), enlargement), Shotha-nashak (anti-inflammatory), Kamala-nashak (alleviating jaundice), Jalodarnashan (useful in ascites), Pramehaghna (useful in prediabetic and diabetic states), and Hrudya (favourable for the heart) [17].

3 BOTANICAL AND MORPHOLOGICAL DESCRIPTION [18].

Feature	Description					
Plant Type	Small perennial herb					
Height	10-20 cm					
Stem	Small, leafy, weak, creeping; erect at flowering					
Roots	6-10 inches long, hard, wrinkled, greyish to brown					
Rhizomes	2.5–12.0 cm long, 0.3–1.0 cm thick, cylindrical, irregularly curved, jointed nodes w					
	branching and rooting					
Rhizome	Externally greyish-brown					
Appearance						
Aerenchyma	Found in leaves, aerial stem, and rhizome					
Presence						
Leaves	Basal, alternate, 5-15 cm long, oval, acuminate, serrate, stalked, winged,					
	oblanceolate/narrowly spathulate, coarsely toothed					
Leaf Hairs	Two types of glandular hairs present					
Flowers	White or pale purple, bisexual, on long terminal spikes					
Calyx	5 equal parts					
Corolla	4-5 lobed, actinomorphic					
Stamens	4, slightly didynamous, inserted on corolla tube					
Stigma	Capitate					
Fruits/Capsules	Acute, ovoid, swollen, 6-10 mm, tapered at the top, splits into 4 valves					
Seeds	Numerous, pale brown, 1 x 0.8 mm, ellipsoid, thick transparent seed coat					





5 TAXONOMY OF PICRORHIZA KURROA (Table no. 1)

Table No. 1

TUDIC ITOL I					
Kingdom	Plantae				
Subkingdom	Tracheobionta				
Super-division	Spermatophyta				
Division	Magnoliophyta				
Class	Mannoliopsida				
Subclass	Asteridae				
Order	Lamiales				
Old Family	Scrophulariaceae				
New Family	Plantaginaceae				
Genus	Picrorhiza				
Species	P. kurroa				

6 Phytochemistry of P. kurroa

Picrorhiza has been extensively studied for its chemical properties, resulting in the extraction of numerous physiologically active compounds from its rhizomes, roots, seeds, stems, and leaves. This plant has produced over 65 secondary metabolites. The rhizomes and roots of P. kurroa yield a crystalline compound known as "Picroliv" or "Kutkin," which consists of two C9-IGs (C9-Iridoid Glycosides) referred to as kutkoside. Picroside-II and Picroside-I occur in a ratio of 2:1 [19]. Using HPLC (highperformance liquid chromatography) methods, the IGs (Iridoid Glycosides) Picrosides I, II, and III, cucurbitacins, and kutkoside were identified in P. kurroa rhizomes [20]. P. kurroa possesses a variety of bioactive compounds with pharmacological and therapeutic potential, including glycosides, iridoids, alkaloids, phenolics, terpenes, and cucurbitacins [21-26]. Apart from its purgative and emaciating qualities, picrorhiza also possesses cholagogue, digestive qualities, appetizing qualities, and actions that stimulate the liver and heart. Kutkin, the active ingredient in P. kurroa, is a combination of picroside and kutkoside. Pharmacologically, kutkin (picrosides

and kutkosides) has demonstrated hepatoprotective qualities. Two additional bioactive compounds that were separated from this plant are drosin and apocyanin. One catechol that might inhibit neutrophils' oxidative response is apocynin [27]. P. kurroa contains cucurbitacins B, D, and R, which are well known for their cytotoxic and anti-tumorous qualities. Apocynin has anti-inflammatory and antioxidant qualities and is a strong inhibitor of **NADPH** (nicotinamide adenine dinucleotide phosphate) oxidase. There is an anti-asthmatic effect of androsin [28]. The 70 percent hydroalcoholic fraction has also been found to contain flavonoids like apocynin and vanillic acid, kutokoside, pikurosides, and picrosides I, II, III, and IV, according ESI-MS/MS (liquid chromatography-LC electrospray ionization/multi-stage spectrometry) techniques [29]. Other significant substances found in P. kurroa include the carbohydrate D-mannitol and aromatic acids like ferulic acid, cinnamic acid, and vanillic acid [30,31]. A variety of P. kurroa secondary metabolites are displayed in Figure [32,33].

Picroside I (C₂₄H₂₈O₁₁₎

Picroside II (C₂₃H₂₈O₁₃₎

H³CO HO OH OH OH

Picroside III (C₂₅H₃₀O₁₎



Figure: Various bioactive principles from P. kurroa

Cinnamic acid (C₆H₅CH=CHCOOH

P. kurroa has undergone a physicochemical analysis in certain investigations. The following were reported: moisture content (12.95% v/w), insoluble acid ash (2.14% w/w), water-soluble ash (3.35% w/w), total ash (5.92% w/w), and pH of 10% aqueous solution (12.95) [34–36]. P. kurroa roots were found to contain vanillic acid, the primary phenolic component, at a rate of 161.2 mg/100 g dry weight [37]. In a different study, the total phenolic content of aqueous and methanolic extracts at different concentrations (50, 100, P. kurroa has undergone a physicochemical analysis in certain investigations.

Total phenolics were more abundant in the methanolic extract (14.11–35.36 g GAE) than in the aqueous extract (10.79–24.87 g GAE) [38]. The total phenolic content of roots was found to be 3.14 mg/100 g of dry weight based on HPLC analysis [39]. In a related study, the total phenolics and flavonoids in a 70% ethanol extract of dry root were found to be 222 μg GAE/mg and 197 μg QE/mg. It was determined that the hydro methanolic extract of P. kurroa contained 400 mg RE of flavonoids and 544 mg GAE/q of total phenolics [40].

7 Classical views of kutki (P. kurroa)

Picein (C₁₄H₁₈O₇₎

Picrorhiza kurroa, commonly known as Kutki, has been recognized as a medicinal plant in the three major Ayurvedic treatises, namely Charaka Samhita, Sushruta Samhita, and Ashtanga Hridaya. [41-44]

Charaka Samhita: Bhedaneeya, Lekhaneeya and Stanyashodhana Mahakashaya, Tikta Skandha.

Sushruta Samhita: Patoladi, Pippalyadi, and Mustadi gana.

Ashtanga Hridaya: Patoladi, Pippalyadi, and Mustadi gana.

Ashtanga Sangraha: Stanyashuddhikaragana, Patoladi, Pippalyadi, and Mustadi gana.

- Furthermore, the continuous and extensive clinical use of Kutki (P. kurroa) is reflected in its inclusion in most of the medieval compendia, such as Nighantus:
 - Bhavaprakasha Nighantu (Hartikyadi varga) [17]
 - Madanapala Nighantu (Abhayadi varga)[45]
 - Raja Nighantu (Pippalyadi varga) [46]

Int J Pharm Biol Sci.



Dhanvantari Nighantu (Guduchyadi varga) [47]

Kaiyadeva Nighantu (Aushadhi varga) [48]

> Ashtanga Nighantu (Mustadi gana) [49]

Madanadi Nighantu (Panchadasha gana) [50]

> Sodhala Nighantu (Guduchyadi varga) [51]

Paryayaratnamala [52]8 Properties and Properties [17]

Rasa: Tikta (Bitter) Vipak : Katu (Pungent)

Guna: Ruksa, Laghu (Light to digest)

Veerya: Shita (Cold potency)

Karma: Hrudya, Deepaniya, Bhedini, Pittahara

In the Ayurvedic texts (Samhitas) and Nighantus, the properties and uses of *Kutki* are described as follows: (**Table no. 2**)

S. No.	Roghaghnata (Indication)	C.S. [42]	S.S. [43]	A.H. [44]	A.S. [45]	B.N. [18]	M.N. [46]	R.N. [47]	D.N. [48]	K.N. [49]	M.N [51]
1	Asrajit (useful in bleeding disorders)					+		+	+		
2	Dahajit (relieves burning sensation)					+	+	+	+	+	+
3	Aruchinashak (Anti- anorexic/Appetizer)		+	+	+			+	+		
4	Vishamajvara (useful in chronic recurrent fever)								+		+
5	Bhedani, (piercing, causes purgation)					+				+	
6	Rechani (Purgative)							+			+
7	Deepani (Appetizer)		+			+				+	
	Hrudya/Hrudyaroga (good for										
8	heart and useful in heart diseases)					+		+			+
9	Jwraghna/Kapha Pitta Jwarapaha (Antipyretic/ useful in fever of Kapha and Pitta			+	+	+	+	+		+	+
10	origin) <i>Pramehahara</i> (useful in urinary										
10	tract disorders, diabetes) Shvasahara (useful in asthma,					+				+	
11	dyspnoea)					+	+	+		+	
12	Kasa (useful in cough, cold)					+				+	
13	Kushtahara (useful in skin disorders)			+	+					+	
14	Chardinashana (Anti-emetic)		+	+	+						
15	Hikkanashana (Useful in										
15	Hiccups)		+								
16	Kamalanashak (Alleviating jaundice)			+	+						+
17	Vishaghna (Detoxicant)		+	+							+
18	Krumighna (Antimicrobial)					+	+			+	
19	Sheetapittaghna (Anti-urticarial) Stnyashodhana/Stnyadoshahara							+			
20	(Improving the quality of breast milk)	+	+	+	+						
21	Yonidoshahara (Vaginal disorders)		+	+	+						
22	Pratishyaya roga (Nasal disorders)		+	+	+						
23	Arsha roga (Hemorrhoids)									+	
	7.1.3.1.4.1.0gu (Helliottilolus)									•	

9 Formulations of Picrorhiza kurroa



Arogya vardhini, Tiktaka Ghruta, Mahatiktaka Ghruta, Sarivadyasava, Mahayograj Guggulu, Sudarshan choorna, Katukadyavaleha, Tiktadi kwath, Sarvajvarahara Lauha are some of the multiingredient formulations mentioned in the samhitas, containig Kutki (P. Kurroa) as one of the components, which could be present in a significant or minor amount.

10 Pharmacological Activities of P. kurroa

Pharmacological Characteristics of *P. kurroa* Rhizomes

Pharmacological Activity	Action	Concentration	Study Model	Reference
Hepatoprotective	Prevents biochemical alterations in the liver and serum of galactosamine-intoxicated animals	12 mg/kg/day for 7 days	Rats	[111]
Anti-diabetic	Reduces blood glucose levels under normal and high glucose conditions	75 mg/kg extract of body weight	Rats	[14]
Anti-diabetic	Assesses glucose tolerance in normal, non-diabetic rats	100 mg/kg – 200 mg/kg, p.o.	Rats	[72]
Anti-diabetic	Increases GLUT-4 levels in the soleus muscle membrane fractions	100 mg/kg/day – 200 mg/kg/day	Rats	[73]
Anticancer	Provides protection against hypoxic damage	-	Hep3B and glioma cells	[90]
Anticancer	Suppresses FMuLv- induced erythroleukemia	-	BALB/c mice	[86]
Anticancer	Counteracts DMH-induced liver carcinogenesis Prevents	40 mg/kg and 200 mg/kg	Rats	[87]
Anticancer	Hepatocarcinogenesis caused by N- Nitrosodiethylamine	200 mg/kg body weight	Rats	[88]
Anticancer	Inhibits sarcoma and papilloma formation induced by specific carcinogens	100 mg/kg – 200 mg/kg, p.o.	BALB/c mice	[89]
Anticancer	Induces cell toxicity and reduces levels of MMP-1, MMP-13, MMP-2, and MMP-9	50 μg/mL and 100 μg/mL	MCF-7 cell lines (Human breast cancer)	[85]
Anticancer	Demonstrates maximum cytotoxic effects	50 mg/kg for 10 days	Ehrlich ascites carcinoma tumor- bearing mice	[84]
Antitumor & Anti- carcinogenic	Inhibits yeast topoisomerase I and II enzymes	250 μg/mL	Tumor-bearing mice	[7]
Anticarcinogenic	Affects serum and tissues of tumor-bearing animals	150 mg/kg – 750 mg/kg body weight	Rats	[78]
Cytoprotection	Reduces adriamycin- induced cardiomyopathy	50 mg/kg body weight	Rats	[53]
Anti-HBsAg Activity	Exhibits hepatoprotective properties	-	Serum	[91]
Anti-anaphylaxis	Suppresses passive cutaneous anaphylaxis	25 mg/kg p.o.	Mice/Rats/Guinea Pig	[106]



Antileishmanial	Boosts lymphocyte proliferation and enhances antileishmanial activity	20–5 mg/kg dose for 12 days and 10 mg/kg for further experiments	Leishmania donovani/Hamster	[106]
Antifungal Activity	Inhibits the growth of dermatophytic fungi	5% and 10% alcoholic solvent extracts of root and rhizome	Solidified agar plates	[15]
Immunomodulatory	Increases cytokine levels (IFN-γ, IL-4) and promotes lymphocyte proliferation	12.5 mg/kg, 25 mg/kg, and 50 mg/kg body weight for 14 days	Balb/c mice	[5]
Immunomodulatory	Enhances both cell- mediated and humoral immune responses	-	Mice and rats	[97]
Immunomodulatory	Modulates hypersensitivity reactions Exhibits antioxidant	100 mg/kg body weight	Albino mice	[96]
Antioxidant & Anti- neoplastic	effects by scavenging free radicals and inhibiting lipid peroxidation	-	Hep3B, PC-3, MDA- MB-435S	[62]
Anticonvulsant	Reduces seizure intensity and mortality in various chemically and electrically induced convulsions Decreases joint	100 mg/kg	Mice	[108]
Anti-arthritic	inflammation and downregulates inflammatory cytokines (IL-1β, IL-6, TNF-R1, VEGF) in arthritis models	Picrorhiza kurroa rhizome extract (PKRE)	Rats	[11]

10.1 Hepatoprotective

In comparison to hepatoprotective substances like catapol, silymarin, and rographolide, picroliv, which was derived from *P. kurroa*, has been evaluated for its effects against ethanol-induced toxicity in rats. The findings indicated a minimal hepatoprotective effect, as evidenced by the reduced levels of marker enzymes such as alkaline phosphatase, glutamicoxaloacetic transaminase, glutamic pyruvic transaminase, and aldehyde dehydrogenase [3].

A methanol extract derived from the rhizomes of *Picrorhiza kurroa Royle ex Benth*. (Plantaginaceae) showed hepatoprotective properties against liver damage in mice induced by D-galactosamine (D-GalN)/lipopolysaccharide (LPS) [55].

The active component of the plant *Picrorhiza kurroa*, known as picroliv, was evaluated for its protective effects on the liver in rats suffering from ethanol-induced liver damage. Administration of alcohol (3.75 g/kg for 45 days) altered liver markers (lipid, glycogen, and protein) and certain serum markers

(AST, ALT, and ALP) by 20–114%. Furthermore, the treatment reduced the viability of ex vivo isolated hepatocytes (by 44–48%) as measured by oxygen uptake rate and Trypan blue exclusion [56].

In the liver and kidneys of the test rats, Picroliv exhibited hepatoprotective effects against AFB1 toxicity, comparable to that of standard silymarin [57].

The liver-protecting properties of Enliv, an herbal formulation that includes *P. kurroa*, were enhanced by counteracting the decline in glutathione levels induced by paracetamol and elevating lipid peroxidation levels in liver tissue [58].

10.2 Antioxidant

Kant et al. (2013) investigated the antioxidant potential of *P. kurroa* leaves, aiming to discover a new natural source of antioxidants. Various solvent extracts, including ethanol, butanol, and ethyl acetate, along with an isolated compound, luteolin-5-O-glucopyranoside, were evaluated for their



antioxidant activities using DPPH and ABTS assays. luteolin-5-0results showed that glucopyranoside exhibited an IC50 value comparable to that of standard ascorbic acid, indicating that P. kurroa leaves may serve as a natural source of antioxidants [9].

The antioxidant activities of two Picrorhiza species, P. kurroa and P. scrophulariiflora, were compared at a concentration of 0.1 mg/ml, and the results showed that P. scrophulariiflora had higher radical scavenging activity (34.30%) than P. kurroa (37.70%) [59].

The 70% hydroalcoholic extract of P. kurroa was found to exhibit antioxidant activity by inhibiting DNA damage and lipid peroxidation induced by hydrogen peroxide, with IC50 values of 75.16 and 55.50 µg/ml in DPPH and metal chelating antioxidant assays [60].

Rajaprabhu et al. (2007) found that P. kurroa extract exhibited significant antioxidant and protective effects when administered orally to rats (50 mg/kg body weight/day for 15 days), countering the detrimental effects of adriamycin [61].

Extracts of P. kurroa exhibited exceptional antioxidant potential in scavenging radicals and inhibiting lipid peroxidation, as evidenced by assays such as DPPH, FRAP, and TBA [62]

Choi et al. (2008) conducted a study to evaluate the antioxidant activity of enzymatic extracts of P. kurroa against hydroxyl radical-induced DNA damage using ESR spectroscopy, and their findings suggest that P. kurroa may emerge as a valuable natural source of antioxidants [63].

10.3 Anti-inflammatory

Rohit Kumar et al. (2016) found that PKRE, a hydroalcoholic extract of P. kurroa rhizome, exhibited anti-inflammatory effects by attenuating paw oedema, granuloma formation, proinflammatory mediator production in macrophages, achieved by inhibiting the iNOS and NF-κB signalling pathway [64].

The anti-inflammatory effect of Picrorhiza Kurroa extract (PK) was confirmed to be mediated by βadrenergic blockade, suggesting that PK treatment alters cell-surface biology. Furthermore, the blockade of protein synthesis by cycloheximide pretreatment reduced the effect of PK, indicating that protein mediation plays a role. Additionally, the metabolic inhibitor dinitrophenol equally inhibited inflammatory edema in both control and PK-treated animals, leading to the conclusion that PK's effect is masked [65].

B.L. Pandey et al. investigated anti-inflammatory action of P. kurroa extract (PK) in albino rats. Research showed that the anti-inflammatory effect of PK treatment was reversed at specific time intervals (1 hour, 3 hours, and 5 hours) following the depletion of mast cells, neutrophils, macrophages, respectively [66].

Apocynin, a key constituent of *P. kurroa* roots, has been found to exhibit anti-inflammatory properties by inhibiting thromboxane A2 formation, stimulating prostaglandin release, and ultimately preventing bovine platelet aggregation in arachidonic acidinduced inflammation [67].

Kutkin, a bioactive compound present in the alcoholic root extract of P. kurroa, has been identified as the primary contributor to its antiinflammatory effects in animal models [68].

10.4 Anti-asthmatic

Studies have shown that androsin, a constituent of P. kurroa, significantly reduced allergen-induced bronchial obstruction in guinea pigs. The plant extract also inhibited histamine release in vitro [70]. Additionally, an iridoid glycoside fraction isolated from P. kurroa roots and rhizomes demonstrated anti-allergic properties by inhibiting passive cutaneous anaphylaxis and preventing mast cell degranulation in rats and mice [70,71].

10.5 Anti-diabetic

Research has found that P. kurroa extract reduces serum lipid peroxide levels and blood urea nitrogen in diabetic rats induced by alloxan, while also inhibiting weight loss and leukopenia. This suggests that P. kurroa extracts may mitigate metabolic damage caused by alloxan [14].

Moreover, studies have provided in vivo evidence that standardized extracts of P. kurroa exhibit considerable antidiabetic activity in rats with type-2 diabetes [72].

Another study has elucidated the mechanism of antidiabetic activity of P. kurroa extract, revealing its ability to enhance insulin-mediated glucose uptake in skeletal muscle [73].

Nirja et al. investigated the antidiabetic properties of P. kurroa extract in a male Wistar rat model. The results of the study demonstrated a significant reduction in blood glucose levels, as well as improvements in lipid profile levels. These findings suggest that P. kurroa extract may have a beneficial effect on glucose metabolism and lipid profiles. Furthermore, the extract showed promising results in both in-vitro and in-vivo studies, indicating its potential as a natural remedy for the treatment and management of diabetes [74].

The methanolic extract of *Picrorhiza kurroa* has been found to possess high total phenolic content (TPC) and total flavonoid content (TFC), as well as exceptional antioxidant potential. This is attributed



to its high scavenging capacity, as demonstrated by its performance in DPPH, ABTS, and FRAP assays. Furthermore, the extract's significant α -amylase and α -glucosidase inhibitory activities suggest its potential as a natural therapeutic agent for the treatment and management of diabetes [75].

A study found that administering an herbal formulation containing *P. kurroa* to streptozotocininduced diabetic rats resulted in decreased levels of malondialdehyde and NADPH-oxidase dependent superoxide formation in the diabetic kidney. This indicates that *P. kurroa* may play a protective role in diabetic nephropathy by mitigating oxidative stress and inflammation [76].

A study found that KT fraction of PKE enhanced adipogenesis, improved lipid metabolism, and increased insulin sensitivity in adipocytes. KT activated PPAR γ , PI3K/Akt signalling, and upregulated adiponectin, while reducing TNF α expression. Additionally, KT promoted insulinmediated glucose uptake and activated PI3K/Akt axis even in the presence of a PI3K inhibitor [104].

10.6 Anti-arthritic

R. Kumar et al. evaluated the anti-arthritic activity of *Picrorhiza kurroa* (PK) in rat models of formaldehyde and adjuvant-induced arthritis (AIA). The study found that administration of *Picrorhiza kurroa* rhizome extract (PKRE) resulted in significant reductions in joint inflammation and inflammatory cytokine expression, including IL-1 β , IL-6, TNF-R1, and vascular endothelial growth factor, in AIA-induced arthritic rats. These results suggest that PKRE may exert its anti-arthritic effects by suppressing the expression of inflammatory cytokines and modulating the inflammatory response in arthritic joints [11].

10.7 Anti-cancer

A study investigated the antineoplastic and antioxidant properties of methanolic extracts and aqueous extracts of *P. kurroa* rhizomes. The extracts were tested for their cytotoxic effects on human breast carcinoma, human hepatocellular carcinoma, and human prostate cancer cell lines using the XTT assay. The results demonstrated that both extracts exhibited significant antineoplastic activity by inducing apoptosis in all three cancer cell lines, suggesting their potential as natural anticancer agents [77].

Rathee et al. (2013) found that iridoid glycosides, including picroside-I, kuthkoside, and kutkin, isolated from *P. kurroa*, exhibited anti-invasive and antimigratory effects on MCF-7 breast cancer cells by reducing the activity of matrix metalloproteinases (MMP-2, 9, and MMP-1,13). These compounds also

suppressed the expression of MMPs at both protein and mRNA levels, leading to the inhibition of inflammatory mediators and the suppression of cancer cell invasion and migration [85].

Additionally, Mallick et al. (2015) found that the dichloromethane fractions of *P. kurroa* rhizome exhibited significant anticancer effects, including a reduction in tumor cell count and restoration of haematological parameters, which were attributed to the presence of cucurbitacin B, cucurbitacin E, picroside-I, picroside-II, betulinic acid, and apocynin [84].

The extract of *P. kurroa* was found to significantly inhibit hepatocarcinogenesis induced by N-nitrosodiethylamine, suggesting its potential as a chemopreventive agent against liver cancer [78].

Kong et al. demonstrated that Cucurbitacin E (CuE) inhibits the growth of human breast cancer cells in vitro by inducing apoptosis and cell cycle arrest. The study suggested that CuE's anti-cancer effects are mediated through the inhibition of STAT3 function. Additionally, the results showed that low-dose CuE enhances the efficacy of cisplatin in inhibiting breast cancer cell growth, highlighting its potential as an adjunctive therapy for breast cancer [79].

Another study revealed that combining cucurbitacin B (CuB) at a relatively low concentration with either Docetaxel (DOC) or Gemcitabine (GEM) exhibits significant antiproliferative activity against breast cancer cells, without increasing toxicity [80].

Another study's findings strongly suggest that picroliv is a beneficial agent for reducing damage caused by chemotherapy and radiation [81].

Jun-shan zhu et al. discovered that CuB exhibits antitumor activity in Jurkat cells, inhibiting their growth in a dose-dependent manner. Treatment with CuB arrested cell cycle progression at the G2/M phase, inducing both apoptotic cell death and autophagy [82].

Duangmano et al. demonstrated that cucurbitacin B possesses strong antiproliferative properties against breast cancer cells, inhibiting their growth in a dose-dependent manner. The mechanism of action involves disruption of microtubule polymerization and nucleophosmin/B23 translocation, resulting in cell cycle arrest at the G2/M phase and apoptosis. These results indicate that cucurbitacin B may be a promising agent for breast cancer therapy [83].



10.8 Anti-microbial

Methanol extract of P. kurroa rhizomes exhibited significant antimicrobial activity, comparable to the standard antibiotic Ciprofloxacin, as determined by the cup plate method. Additionally, the aqueous extract demonstrated antifungal activity against fungal strains, similar to the standard antifungal agent Fluconazole [92].

A study has demonstrated that acetone and methanol extracts of dried stolons of P. kurroa exhibit potent antimicrobial activity against a broad range of microorganisms, including bacteria and fungi. The extracts showed dose-dependent inhibitory effects against Bacillus subtilis, Erwinia chrysanthemi, Escherichia coli, Gloeocercospora sorghi, Fusarium oxysporum, Rhizoctonia solani, and Sporisorium scitamineum. Significantly, the extracts demonstrated greater antibacterial activity against B. subtilis than the standard antibacterial agent used in the study, highlighting their potential as natural antimicrobial agents [93].

A study on the leaf extracts of Picrorhiza kurroa revealed significant antibacterial and antioxidant activity. The acetone extract demonstrated notable antibacterial potency against S. aureus, S. mutans, E. coli, K. pneumoniae, and P. aeruginosa. Although the antioxidant activity of the extracts was moderate compared to ascorbic acid, the findings suggest that Picrorhiza kurroa leaf extracts may be a valuable source of natural antimicrobial and antioxidant agents [94].

Rathee et al. conducted a study to evaluate the antimicrobial activity of methanolic and aqueous extracts of P. kurroa against various fungi and bacteria. The study involved in vitro testing against two fungal strains (Aspergillus niger and Candida albicans) and five bacterial strains (Escherichia coli, Pseudomonas aeruginosa, Staphylococcus aureus, Micrococcus luteus, and Bacillus subtilis). The results showed that the methanolic extract exhibited significant activity against P. aeruginosa and S. aureus, and moderate activity against E. coli, B. subtilis, and M. luteus. In contrast, the aqueous extract had no activity against any of the tested fungi

10.9 Immunomodulatory

A study conducted by Hussain et al. demonstrated the immunomodulatory properties of Picrorhiza kurroa in an in vivo model of immunosuppressed mice. Cyclophosphamide was employed to induce immunosuppression, and the results showed that the alcoholic extract of the plant significantly augmented immunostimulant activities [95].

A study evaluated the immunomodulatory properties of RLJ-NE-205, a biopolymeric fraction isolated from P. kurroa, in a murine model. The results demonstrated a statistically significant increase in cytokine levels (IFN-y and IL-4) and lymphocyte proliferation at a dose of 50 mg/kg, suggesting that RLJ-NE-205 may function as a biological response modulator, influencing the immune response in a dose-dependent manner [5]. Research has shown that a 50% ethanolic extract of P. kurroa leaves possesses immunomodulatory effects, enhancing both cellular and humoral immune components, as well as phagocytic activity, in animal models. Specifically, the extract was found to augment humoral immune responses in rats and mice, and increase phagocytic activity in the reticuloendothelial system of mice, indicating its potential as an immunoenhancing agent [97].

This study demonstrated the efficacy of AEV01 in treating COVID-19 patients. The results showed significant improvements in oxygen saturation and cough relief, with a reduced recovery time of 4.5 days compared to 9.1 days in the placebo group. Additionally, AEV01 treatment led to improvements in key biomarkers, including ESR, LDH, serum ferritin, and inflammatory cytokines. Overall, the study suggests that AEV01 is a promising treatment for COVID-19, offering rapid and improvements in clinical outcomes [98].

10.10 Cardioprotective

A study examined the cardioprotective effects of P. adriamycin-induced kurroa rats with cardiomyopathy. Adriamycin treatment resulted in elevated plasma marker enzymes, lipid peroxidation, and decreased glutathione levels and antiperoxidative enzymes in cardiac tissue. Conversely, oral administration of P. kurroa effectively counteracted these changes, returning the rats to a normal state [53]

Research has shown that an ethanolic extract of P. kurroa's rhizome and roots effectively reversed myocardial stress induced by isoproterenol in mice, highlighting its potential as a cardioprotective agent. [54]

10.11 Anti-ulcer

A study assessed the gastroprotective effects of an ethanol extract of P. kurroa roots and rhizomes against ethanol/HCl-induced gastric ulceration in rats. The extract's effects on gastric mucosal pepsin, antioxidant status, glycoproteins, and proteins were evaluated. The results showed that oral treatment with the extract for 10 days resulted in a significant reduction in ulceration, suggesting its potential as a gastroprotective agent [99].

A study investigated the gastroprotective effects of the methanol extract of P. kurroa rhizomes on indomethacin-induced gastric ulcers in mice. The



results showed that the extract accelerated ulcer healing by modulating key physiological processes, including mucin secretion, oxidative stress reduction, prostaglandin production enhancement, and cyclooxygenase enzyme expression upregulation, thereby demonstrating its potential as a therapeutic agent for gastric ulcer treatment [100].

10.12 Hypolipidemic Effect

Lee et al. assessed the hepatoprotective and hypolipidemic potential of *P. kurroa* root extract (PKRE) in a murine model of high-fat diet-induced hyperlipidaemia. The results demonstrated that PKRE supplementation significantly reduced serum levels of total cholesterol, LDL cholesterol, triglycerides, and liver enzymes (ALT and AST), while also decreasing liver weight. Conversely, PKRE did not exert any significant effects on serum HDL cholesterol levels, suggesting a neutral impact on HDL metabolism [101].

A study investigated the hepatoprotective and hypolipidemic effects of the water extract of *P. kurroa* (PR) in mice with hyperlipemia induced by poloxamer (PX)-407. The efficacy of PR was compared to that of simvastatin, a standard lipid-lowering agent. The results demonstrated that PR extract and simvastatin treatment resulted in dose-dependent reductions in serum triglycerides, LDL cholesterol, and total cholesterol levels [102].

A study on *Picrorhiza kurroa* plant extract showed it significantly reduced cholesterol and triglyceride levels in hyperlipidaemic rats, with high-dose (200mg/kg) extract being most effective, comparable to standard drug Atorlip-20 [103]

10.13 Antileishmanial activity

The synergistic potential of Picroliv in enhancing the antileishmanial efficacy of conventional drugs has been elucidated in various studies. Specifically, the combination of Picroliv with ketoconazole and miltefosine resulted in a significant augmentation of antileishmanial activity, with efficacy increasing from 72% to 82% against visceral leishmaniasis [12].

Picroliv also showed significant enhancement of antileishmanial effects when combined with paromomycin and miltefosine [105-106].

10.14 Antimalarial activity

An in vivo study evaluated the antimalarial efficacy of the ethanolic extract of *Picrorhiza kurroa* roots and leaves against Plasmodium berghei in a murine model. The results demonstrated that the root extract possessed significant antimalarial activity, with a more pronounced inhibitory effect on malaria parasites compared to the leaf extract [107].

10.15 Anticonvulsant effect

An in vivo study evaluated the anticonvulsant activity of the ethanolic root extract of *Picrorhiza kurroa* in a

mouse model of maximal electroshock-induced seizures and pentylenetetrazole, picrotoxin-induced seizures. The findings revealed that administration of the extract at a dose of 100 mg/kg significantly attenuated convulsive seizures and mortality, thereby confirming its anticonvulsant properties [108]

11 TOXICITY STUDY

A study was conducted to evaluate the oral toxicity of *Picrorhiza kurroa* root extract (PKRE) in Wistar rats. A single high dose of 2000 mg/kg was administered, and the rats were observed for 14 days. The results showed no signs of toxicity, and the rats exhibited weight gain, indicating a high LD50 value [109].

The results of the acute oral toxicity study indicated that the maximum non-lethal dose of *Picrorhiza kurroa* (*P. kurroa*) in rats is less than 2000 mg/kg of body weight per day, administered in two fractional doses. Moreover, the study showed that *P. kurroa* exhibits thermal stability, retaining its properties when stored at temperatures up to 50°C for a duration of three months [110].

Despite its diverse applications, *Picrorhiza kurroa* has been subjected to limited toxicity studies. Consequently, further toxicological investigations are warranted to elucidate the potential adverse effects associated with its use. These studies are essential for ensuring the safe handling, production, and utilization of its bioactive extracts and secondary metabolites.

12 CONCLUSION & FUTURE PERSPECTIVES

This comprehensive review aims to provide an updated overview of the versatile applications of Picrorhiza kurroa, an herb characterized by its diverse pharmacological properties and widespread inclusion in herbal preparations. As a cornerstone of traditional Indian medicine, Picrorhiza kurroa presents a promising alternative to conventional therapeutic modalities. Its profound ethnomedical significance underscores the need for further research to delineate the bioactive compounds and molecular mechanisms of action responsible for its therapeutic efficacy. A substantial body of evidence from in vitro and in vivo studies has substantiated the pharmacological relevance of the bioactive secondary metabolites present in Picrorhiza kurroa. However, a critical knowledge gap persists due to the scarcity of human clinical trials to validate these preclinical findings. Thus, there is an urgent need for well-designed human clinical trials to confirm the efficacy and safety of *P. kurroa*, which will ultimately



inform the development of novel therapeutic applications.

REFERENCES

- Chettri N, Sharma E, Lama SD. Non-timber forest produces utilization, distribution and status in a trekking corridor of Sikkim, India. Lyonia. 2005;8: 89-101.
- Dutta IC. Non-Timber Forest Products of Nepal: Identification, Classification, Ethnic Uses and Cultivation. Pokhara, Nepal: Inst. For.; 2007.
- Visen PKS, Saraswat B, Patnaik GK, Agarwal DP, Dhawan BN. Protective activity of Picroliv Isolated from *Picrorhiza Kurroa* against ethanol toxicity in isolated rat hepatocytes. *Indian J Pharmacol*. 1996;28: 98-101.
- 4. Gupta BD, et al. Anti-inflammatory activity of the iridoids kutkin, picroside-1 and kutkoside from *Picrorhiza kurroa*. *Phytother Res.* 1993;7: 402-407.
- Gupta A, Khajuria A, Singh J, Bedi KL, Satti NK, Dutt P, Suri KA, Suri OP, Qazi GN. Immunomodulatory Activity of Biopolymeric Fraction RLJ-NE-205 from Picrorhiza kurroa. Int Immunopharmacol. 2006;6: 1543-1549.
- Rathee D, Rathee P, Rathee S. Phytochemical screening and anti-microbial activity of *Picrorrhiza* kurroa, an Indian traditional plant used to treat chronic diarrhoea. Arabian J Chem. 2016;9(Suppl 2): S1307-S1313.
- 7. Joy KL, Rajeshkumar NV, Kuttan G, Kuttan R. Effect of *Picrorrhiza kurroa* Extract on Transplanted Tumours and Chemical Carcinogenesis in Mice. *J Ethnopharmacol*. 2000;71: 261-266.
- Soni D, Grover A. Picrosides" from Picrorhiza kurroa as potential anti carcinogenic agents. Biomed Pharmacother. 2019;109: 1680-1687.
- Kant K, Walia M, Agnihotri VK, Pathania V, Singh B. Evaluation of antioxidant activity of *Picrorhiza kurroa* (leaves) extracts. *Indian J Pharm Sci.* 2013;75: 324-329.
- Rathee D, Thanki M, Bhuva S, Anandjiwala S, Agrawal R. Iridoid glycosides Kutkin, Picroside I, and Kutkoside from *Picrorhiza kurroa Royle Ex Benth* inhibits the invasion and migration of MCF-7 breast cancer cells through the down regulation of matrix metalloproteinases. *Arabian J Chem.* 2013;6: 49-58.
- 11. Kumar R, Gupta YK, Singh S, Arunraja S. *Picrorhiza kurroa* inhibits experimental arthritis through inhibition of pro-inflammatory cytokines, angio genesis and MMPs. *Phytother Res.* 2016;30: 112-119.
- Gupta S, Ramesh, Sharma SC, Srivastava VML. Efficacy of picroliv in combination with miltefosine, an orally effective antileishmanial drug against experimental visceral leishmaniasis. Acta Tropica. 2005:94: 41-47.
- Sehgal R, Chauhan A, Gilhotra UK, Aindry Gilhotra. invitro and in-vivo evaluation of anti-asthmatic activity of *Picrorhiza kurroa* plant. *IJPSR*. 2013;4: 3440-3443.
- 14. Joy KL, Kuttan R. Anti-Diabetic Activity of *Picrorrhiza Kurroa* Extract. *J Ethnopharmacol*. 1999;67: 143-148.

- 15. Shubha KS, Sumana K, Lakshmidevi L. Antifungal Activity of Solanum Xantocarpum Sch and Wend and *Picrorhiza Kurroa Royle Ex Benth* against Some Clinical Dermatophytes. *Int J Curr Microbiol Appl Sci*. 2016;5: 236-244.
- Nandave M, Ojha SK, Kumari S, Nag TC, Mehra R, Narang R, Arya DS. Cardioprotective effect of root extract of *Picrorhiza kurroa* (*Royle ex Benth*) against isoproterenol-induced cardiotoxicity in rats. *Indian J Exp Biol*. 2013;51: 694-701.
- 17. Chunekar KC. Bhavprakash Nighantu. Varanasi, India: Chaukhamba Bharti Academy; 2010.
- Kumari I, Kaurav H, Chaudhar G. Ethnobotanical Significance of *Picrorhiza kurroa* (*Kutki*), a Threatened Species. *Int J Res Rev*. 2021;8: 363-368.
- Dhawan BN. Picroliv—A New Hepatoprotective Agent from an Indian Medicinal Plant, *Picrorhiza Kurroa*. Med Chem Res. 1995;5: 595-605.
- Nisha S, Vijaylata P, Bikram S, Raghbir C. Intraspecific Variability of Main Phytochemical Compounds in Picrorhiza Kurroa Royle Ex Benth. From North Indian Higher Altitude Himalayas Using Reversed-Phase High-Performance Liquid Chromatography. J Med Plants Res. 2012;6: 3181-3187.
- 21. Katoch M, Fazli IS, Suri KA, Ahuja A, Qazi GN. Effect of Altitude on Picroside Content in Core Collections of *Picrorhiza kurroa* from the North Western Himalayas. *J Nat Med*. 2011;65: 578-582.
- 22. Stuppner H, Mueller E, Wagner H. Cucurbitacins from *Picrorhiza kurroa*. Phytochem. 1991;30: 305-310.
- 23. Stuppner H, Wagner H. New cucurbitacin glycosides from *Picrorhiza kurroa*. *Plan Med*. 1989;55: 559-563.
- 24. Stuppner H, Wagner H. HPLC und DC-Untersuchungen von iridoids, Cucurbitacin and Phenol-Giykosiden aus *Picrorhiza kurroa*. *Sci Pharm*. 1992;60: 73-85.
- Sturm S, Stuppner H. Analysis of cucurbitacins in medicinal plants by high pressure liquid chromatography-mass spectrometry. *Phytochem Anal*. 2000;11: 121-127.
- 26. Ernst E. Major Herbs of Ayurveda. Focus Altern Complement Ther. 2010;8: 98.
- Simons JM, Hart BA, Ip Vai Ching TRAM, Van Dijk H, Labadie RP. Metabolic Activation of Natural Phenols into Selective Oxidative Burst Agonists by Activated Human Neutrophils. Free Radic Biol Med. 1990;8: 251-258.
- 28. Salma U, Kundu S, Gantait S. Phytochemistry and Pharmaceutical Significance of *Picrorhiza kurroa Royle Ex Benth*. In: Phytochemistry and Pharmacology of Medicinal Herbs. Delhi, India: Lenin Media Private Limited; 2017.
- Krupashree K, Hemanth Kumar K, Rachitha P, Jayashree GV, Khanum F. Chemical Composition, Antioxidant and Macromolecule Damage Protective Effects of *Picrorhiza kurroa Royle Ex Benth*. S Afr J Bot. 2014;94: 249-254.
- 30. Kumar N, Kumar T, Sharma SK. Phytopharmacological Review on Genus Picrorhiza. *Int J Univ Pharma Bio Sci.* 2013;2: 334-347.



- 31. Zhu TF, Chen JJ, Sun QW, Yan ZH. Chemical Constituents from Picrorhiza Scrophulariiflora. Chinese Tradit Herb Drugs. 2017;48: 263-265.
- 32. Kumar V, Bansal A, Chauhan RS. Modular Design of Picroside-II Biosynthesis Deciphered through NGS Transcriptomes and Metabolic Intermediates Analysis in Naturally Variant Chemotypes of a Medicinal Herb, Picrorhiza kurroa. Frontiers in Plant Science. 2017;8: Article 564.
- 33. Basu K, Dasgupta B, Ghosal S. Structure of kutkin, the bitter glucoside of Picrorhiza kurroa. J Org Chem. 1970;35: 2971-2975.
- 34. Pathan D, Memon S, Parkar H. Pharmacognostical and Phytochemical Evaluation of Picrorhiza Kurroa. J Curr Pharma Res. 2018;9: 2561-2566.
- 35. Kumar KNS, Padhi MM, Babu R. Evaluation of Phamacognostic and Physicochemical parameters of Picrorrhiza kurroa Royle ex Benth. Int J Ayurvedic Med. 2010;1: 41-47.
- 36. Mall AK, Chaubey S, Tiwari RC, Kour G. Pharmacognostical study of kutaki (Picrorhiza kurroa royle ex. benth). Int J Res Ayurveda Pharm. 2016;7: 36-40.
- 37. Guleria S, Tiku AK, Singh G, Koul A, Gupta S, Rana S. In Vitro Antioxidant Activity and Phenolic Contents in Methanol Extracts from Medicinal Plants. J Plant Biochem Biotechnol. 2013;22: 9-15.
- 38. Rajkumar V, Guha G, Ashok Kumar R. Antioxidant and Anti-Neoplastic Activities of Picrorhiza kurroa Extracts. Food Chem Toxicol. 2011;49: 363-369.
- 39. Surveswaran S, Cai YZ, Corke H, Sun M. Systematic Evaluation of Natural Phenolic Antioxidants from 133 Indian Medicinal Plants. Food Chem. 2007;102: 938-953.
- 40. Mohiuddin R, Mir RH, Wani TU, Shah AJ, Mir PA, Jan R, Faroog S, Mohiuddin I, Banday N. Phytochemical and Pharmacological Properties of Picrorhiza kurroa. In: Edible Plants in Health and Diseases. Singapore: Springer; 2022. p. 399-423.
- 41. Charak Samhita, Vidyotini hindi vyakhya by Kashinath Shashtri and Gorakhnath Chaturvedi, Chaukhamba bharti academy, edition 2009, Sutrasthan4, pg no.72,84, Nidansthan,4/47 pg no.640 Vimansthan 8/143page no. 792.
- 42. Sushrut Samhita, Ayurveda Tattava Sandipika- Hindi Vyakhya, Purvardh, kaviraj Dr. Ambikadutta Shastri, reprint 2009. Sutrasthan 38/22, 38/33, 38/54, N 6/5-6 Nidansthan pg. no. 184-187, 326.
- 43. Gupta KA ed. Astangahrdayam Sutrasthan 15/15, 15/40 Nidansthana, 10/7, 10/38-39; Chaukhamba Prakashan, Varanasi reprint 2022, p. 141, 144, 345 ,349.
- 44. Vagbhata. Ashtanga Samgraha with Hindi Commentary (Chapters 15-16). Edited by Kaviraj Atrideva Gupta. Varanasi: Chaukhamba Orientalia; 2011.
- 45. Sastry JLN. Madanpal Nighantu. Chaukhamba Orientalia; 2010.
- 46. Sankhyadhar SC. Raja Nighantu. Chaukhamba Orientalia; 2012.
- 47. Singh AP. Dhanwantari Nighantu. Chaukhamba Orientalia; 2008.

- 48. Sharma GP. Kaiyadev Nighantu. Chaukhamba Orientalia; 2009.
- 49. Vahata. Astanga Nighantu. Edited by PV Sharma, 1st Edition. Madras: Kuppuswamy Shastri Research Institute; 1973.
- 50. Chandranandana. Madanadi Nighantu. Edited by P Visalakshi. Thiruvananthapuram: Oriental Research Institute & Manuscripts Library, University of Kerala; 1998.
- 51. Sodhala. Sodhala Nighantu; Edited by Priya Vrit Sharma, 1st Edition. Baroda: Oriental Institute; 1978.
- 52. Madhavakara. Paryaya Ratnamala. Edited by Tarapada Chowdhury. Patna: Patna University Press; 1947.
- 53. Rajaprabhu D, Rajesh R, Jeyakumar R, Buddhan S, Ganesan B, Anandan R. Protective effect of Picrorhiza kurroa on antioxidant defense status in adriamycininduced cardiomyopathy in rats. J Med Plant Res. 2007;1:85.
- 54. Hari Senthil Kumar S, Anandan R, Devaki T, Santhosh Kumar M. Cardioprotective Effects of Picrorrhiza Kurroa against Isoproterenol-Induced Myocardial Stress in Rats. Fitoterapia. 2001;72: 402-405.
- 55. Sakamoto Y, Inoue N, Nakanishi Y, Ninomiya K, Yoshikawa M, Muraoka O, Manse Y, Morikawa T. Hepatoprotective Principles from the Rhizomes of Picrorhiza kurroa. Chem Pharm Bull (Tokyo). 2023;46: 848-855.
- 56. Saraswat B, Visen PKS, Patnaik GK, Dhawan BN. Ex vivo and in vivo investigations of picroliv from Picrorhiza kurroa in an alcohol intoxication model in rats. J Ethnopharmacol. 1999;66: 263-269.
- 57. Rastogi R, Srivastava AK, Srivastava M, Rastogi AK. Hepatocurative effect of Picroliv and silymarin against aflatoxin B1 induced hepatotoxicity in rats. Planta Med. 2000;66: 709-713.
- 58. Bhar MK, Das SK, Chakraborty AK, Mandal TK, Roy S. Hepatoprotective effect of Enliv on paracetamolinduced liver damage in broiler chicks. Indian J Pharmacol. 2005;37: 257.
- 59. Tiwari SS, Pandey MM, Srivastava S, Rawat A. TLC densitometric quantification of picrosides (picroside-I and picroside-II) in Picrorhiza kurroa and its substitute Picrorhiza scrophularuflora and their antioxidant studies. Biomed Chromatogr. 2012;26: 61-68.
- 60. Krupashree K, Hemanth Kumar K, Rachitha P, Jayashree GV, Khanum F. Chemical composition, antioxidant and macromolecule damage protective effects of Picrorhiza kurroa Royle ex Benth. South African J Bot. 2014;94: 249-254.
- 61. Rajaprabhu D, Rajesh R, Jeyakumar R, Buddhan S, Ganesan B, Anandan R. Protective effect of Picrorhiza kurroa on antioxidant defense status in adriamycininduced cardiomyopathy in rats. J Med Plant Res. 2007; 1:85.
- 62. Rajkumar V, Guha G, Ashok Kumar R. Antioxidant and anti-neoplastic activities of Picrorhiza kurroa extracts. Food Chem Toxicol. 2011;49: 363-369.
- 63. Choi SH, Kim EK, Lee SJ, Jeon YJ, Moon SH, Lee CH, Park SH. ESR spectroscopy investigation of antioxidant activity and protective effect on hydroxyl



- radical-induced DNA damage of enzymatic extracts from Picrorhiza kurroa. J Food Biochem. 2008;32:
- 64. Kumar R, Gupta Y, Singh S, Raj A. Anti-inflammatory Effect of *Picrorhiza kurroa* in Experimental Models of Inflammation. Planta Med. 2016;82: 1403-1409.
- 65. Pandey BL, Das PK. Immunopharmacological studies on Picrorhiza kurroa Royle-ex-Benth. Part IV: Cellular mechanisms of anti-inflammatory action. Indian J Physiol Pharmacol. 1989.
- 66. Pandey BL, Das PK. Immunopharmacological studies on Picrorhiza kurroa Royle-Ex-Benth. Part V: antiinflammatory action: relation with cell types involved in inflammation. Indian J Physiol Pharmacol. 1988.
- 67. Engels F, Renirie BF, 't Hart BA, Labadie RP, Nijkamp FP. Effects of apocynin, a drug isolated from the roots of Picrorhiza kurroa, on arachidonic acid metabolism. FEBS Lett. 1992;305: 254-256.
- 68. Smit H, Kroes B, van den Berg AJ, van der Wal D, van den Worm E, Beukelman C, Labadie R. Immunomodulatory and anti-inflammatory activity of Picrorhiza scrophulariiflora. J Ethnopharmacol. 2000;73: 101-109.
- 69. Dorsch W, Stuppner H, Wagner H, Gropp M, Demoulin S, Ring J. Anti-asthmatic effects of Picrorhiza kurroa: Androsin prevents allergen- and PAF- induced bronchial obstruction in guinea pigs. Int Arch Allergy Immunol. 1991;95: 128-133.
- 70. Baruah CC, Gupta PP, Nath A, Patnaik LGK, Dhawan BN. Anti-allergic and anti-anaphylactic activity of picroliv - A standardised iridoid glycoside fraction of Picrorhiza kurroa. Pharmacol Res. 1998;38: 487-492.
- 71. Dorsch W, Müller A, Christoffel V, Stuppner H, Antus S, Gottsegen A, Wagner H. Antiasthmatic acetophenones — an in vivo study on structure activity relationship. Phytomedicine. 1994;1: 47-54.
- 72. Husain GM, Singh PN, Kumar V. Antidiabetic Activity of Standardized Extract of Picrorhiza Kurroa in Rat Model of NIDDM. Drug Discov Ther. 2009;3: 88-92.
- 73. Husain GM, Rai R, Rai G, Singh HB, Thakur AK, Kumar V. Potential Mechanism of Anti-Diabetic Activity of Picrorhiza Kurroa. TANG [HUMANITASMEDICINE]. 2014:4: 27.1-27.5.
- 74. Nirja, et al. Evaluation of Antidiabetic Activity of Picrorhiza Kurroa Royle Ex. Benth. - In-Vitro and In-Vivo. IJPSR. 2022:13: 2405-2411.
- 75. Jaweria Nisar et al. Phytochemical Screening, Antioxidant, and Inhibition Activity of Picrorhiza kurroa Against α -Amylase and α -Glucosidase. Biomolecules. 2022;20: 15593258221095960.
- 76. He LJ, Liang M, Hou FF, Guo ZJ, Xie D, Zhang X. Ethanol extraction of Picrorhiza scrophulariiflora prevents renal injury in experimental diabetes via anti-inflammation1 action Running title: EPS inhibits inflammation in diabetic kidney. J Endocrinol. 2009;200: 347-355.
- 77. Rajkumar V, Guha G, Ashok Kumar R. Antioxidant and Anti-Neoplastic Activities of Picrorhiza Kurroa Extracts. Food Chem Toxicol. 2011;49: 363-369.
- 78. Jeena KJ, Joy K, Kuttan R. Effect of Emblica officinalis, Phyllanthus amarus and Picrorrhiza kurroa on N-

- nitrosodiethylamine induced hepatocarcinogenesis. Cancer Lett. 1999;136: 11-16.
- 79. Kong Y, Chen J, Zhou Z, Xia H, Qiu MH, Chen C. Cucurbitacin e Induces Cell Cycle G2/M Phase Arrest and Apoptosis in Triple Negative Breast Cancer. PLoS ONE. 2014;9: e103760.
- 80. Aribi A, Gery S, Lee DH, Thoennissen NH, Thoennissen GB, Alvarez R, Ho Q, Lee K, Doan NB, Chan KT, et al. The Triterpenoid Cucurbitacin B Augments the Antiproliferative Activity of Chemotherapy in Human Breast Cancer. Int J Cancer. 2013;132: 2730-2737.
- 81. Bigoniya P, Warathe A, Singh C. Protective Action of Picroliv Isolated from Picrorhiza Kurroa against Radiation Clastogenecity on Mice Cyclophosphamide-Induced Cytotoxicity in Allium Cepa Root. J Radiat Cancer Res. 2019; 10:58.
- 82. Zhu JS, Ouyang DY, Shi ZJ, Xu LH, Zhang YT, He XH. Cucurbitacin B Induces Cell Cycle Arrest, Apoptosis and Autophagy Associated with G Actin Reduction and Persistent Activation of Cofilin in Jurkat Cells. Pharmacology. 2012;89: 348-356.
- 83. Duangmano S, Sae-lim P, Suksamrarn A, Domann FE, Patmasiriwat P. Cucurbitacin B Inhibits Human Breast Cancer Cell Proliferation through Disruption of Microtubule Polymerization Nucleophosmin/B23 Translocation. **BMC** Complement Altern Med. 2012;12.
- 84. Mallick MN, Singh M, Parveen R, Khan W, Ahmad S, Najm MZ, et al. HPTLC analysis of bioactivity guided anticancer enriched fraction of hydroalcoholic extract of Picrorhiza kurroa. BioMed Res Int. 2015;2015: 513875.
- 85. Rathee D, Thanki M, Bhuva S, Anandjiwala S, Agrawal R. Iridoid glycosides Kutkin, Picroside I, and Kutkoside from Picrorhiza kurroa Royle Ex Benth inhibits the invasion and migration of MCF-7 breast cancer cells through the down regulation of matrix metalloproteinases. Arabian J Chem. 2013;6: 49-58.
- 86. Harikumar KB, Kuttan G, Kuttan R. Inhibition of Progression of Erythroleukemia Induced by Friend Virus in BALB/c Mice by Natural Products-Berberine, Curcumin and Picroliv. J Exp Ther Oncol. 2008;7: 275-284.
- 87. Rajesh kumar NV, Kuttan R. Modulation of Carcinogenic Response and Antioxidant Enzymes of Rats Administered with 1,2-Dimethylhydrazine by Picroliv. Cancer Lett. 2003;191: 137-143.
- 88. Rajeshkumar NV, Kuttan R. Inhibition of N-Nitrosodiethylamine-Induced Hepatocarcinogenesis by Picroliv. J Exp Clin Cancer Res. 2000;19: 459-465.
- Rajeshkumar NV, Kuttan R. Protective Effect of Picroliv, the Active Constituent of Picrorhiza Kurroa, against Chemical Carcinogenesis in Mice. Teratog Carcinog Mutagen. 2001;21: 303-313.
- 90. Gaddipati JP, Madhavan S, Sidhu GS, Singh AK, Seth P, Maheshwari RK. Picroliv—A Natural Product Protects Cells and Regulates the Gene Expression during Hypoxia/Reoxygenation. Mol Cell Biochem. 1999;194: 271-281.



- 91. Mehrotra R, Rawat S, Kulshreshtha DK, Patnaik GK, Dhawan BN. In Vitro Studies on the Effect of Certain Natural Products against Hepatitis B. Virus. Indian J Med Res Sect B Biomed Res Other Than Infect Dis. 1990;92: 133-138.
- 92. Sharma SK, Kumar N. Antimicrobial Screening of Picrorhiza kurroa Royle ex Benth rhizome. Int J Curr Pharm Rev Res. 2012;3: 60-65.
- 93. Laxmi V, Preeti C. Antimicrobial Activity of Dried Stolon Extracts of Picrorhiza Kurroa Royle Ex Benth— An Endemic and Endangered Himalayan Herb. ENVIS Cent Himal Ecol. 2015;23: 127-132.
- 94. Singh G, Bisht D, Arya RKK, Kumar S, Kumar D, Dash AK. Identification and Quantification of six natural compounds from Picrorhiza kurroa leaf extract and their Antibacterial and Antioxidant activity. Res J Pharm Technol. 2022;15.
- 95. Hussain A, Shadma W, Maksood A, Ansari SH. Protective effects of Picrorhiza kurroa on cyclophosphamide-induced immunosuppression in mice. Pharmacognosy Res. 2013;5: 30-35.
- 96. Siddiqui NA, Singh S, Siddiquei MM, Khan TH. Immunomodulatory Effect of Withania somnifera, Asparagus racemosus and Picrorhiza kurroa Roots. Int J Pharmacol. 2012;8: 108-114.
- 97. Sharma ML, Rao CS, Duda PL. Immunostimulatory Activity of Picrorhiza kurroa Leaf Extract. J Ethnopharmacol. 1994;41: 185-192.
- 98. Balan AK, Murugesan SB, Kannan R, Sekhar MM, Murali kumar V. A randomized, placebo controlled, double blinded, parallel group clinical study to evaluate the efficacy and safety of AEV01 along with standard care for elderly patients with mild COVID-19. J Ayurveda Integr Med. 2023;15: 100767.
- 99. Anandan R, Deepa Rekha R, Saravanan N, Devaki T. Protective Effects of Picrorrhiza Kurroa against HCI/Ethanol-Induced Ulceration in Rats. Fitoterapia. 1999;70: 498-501.
- 100. Banerjee D, Maity B, Nag SK, Bandyopadhyay SK, Chattopadhyay S. Healing Potential of Picrorhiza (Scrofulariaceae) Rhizomes Indomethacin-Induced Gastric Ulceration: A Mechanistic Exploration. BMC Complement Altern Med. 2008; 8:3.
- 101. Lee HS, Yoo CB, Ku SK. Hypolipemic Effect of Water Extracts of Picrorhiza Kurroa in High Fat Diet Treated Mouse. Fitoterapia. 2006;77: 579-584.

- 102. Lee HS, Ahn HC, Ku SK. Hypolipemic Effect of Water Extracts of Picrorhiza Rhizoma in PX-407 Induced ICR Mouse Model Hyperlipemic Hepatoprotective Effects: A Prevention Study. J Ethnopharmacol. 2006;105: 380-386.
- 103. Singh V, Chaudhary A. Anti-hyper lipidemic activity of Picrorhiza kurroa Royle ex Benth roots. Int J Drug Dev Res. 2012.
- 104. Sinha K, Kumar S, Rawat B, Singh R, Purohit R, et al. Kutkin, iridoid glycosides enriched fraction of Picrorhiza kurroa promotes insulin sensitivity and enhances glucose uptake by activating PI3K/Akt signaling in 3T3-L1. Phytomedicine. 2022.
- 105. Shakya N, Sane SA, Gupta S. Antileishmanial efficacy of fluconazole and miltefosine in combination with an immunomodulator - Picroliv. Parasitol Res. 2011;108: 793-800.
- 106. Sane SA, Shakya N, Gupta S. Immunomodulatory effect of picroliv on the efficacy of paromomycin and miltefosine in combination in experimental visceral leishmaniasis. Exp Parasitol. 2011;127: 376-381.
- 107. Banyal Hs, Rani, Devi N. Picrorhiza kurrooa Royal Ex Benth Exhibits Antimalarial Activity against Plasmodium berghei Vincke and Lips, 1948. Asian J Biol Sci. 2014;7: 72-75.
- 108. Pathan D, Ambavade S. Anticonvulsant activity ethanolic extract of Picrorhiza Pharmacophore. 2014;5: 141-146.
- 109. Krishna AB, Manikyam HK, Sharma VK, Sharma N. Single Dose Oral Toxicity Study of Picrorhiza Kurroa Rhizome Extract in Wistar Rats. Fundam Toxicol Sci. 2016;3: 9-12.
- 110. Vaidya AB, Sirsat SM, Doshi JC, Antarkar DS. Selected Medicinal Plants and Formulations as Hepato-Biliary Drugs: An Over-View. Indian J Clin Pharmacol Ther. 1996.
- 111. Dwivedi Y, Rastogi R, Garg NK, Dhawan BN. Picroliv and Its Components Kutkoside and Picroside I Protect Liver Against Galactosamine-Induced Damage in Rats. Pharmacol Toxicol. 1992;71: 383-387.
- 112. Baruah CC, Gupta PP, Nath A, Patnaik GK, Dhawan BN. Anti-allergic and anti-anaphylactic activity of Picroliv—A standardised iridoid glycoside fraction of Picrorhiza kurroa. J Ethnopharmacol. 1998;60: 167-174.