



INVESTIGATIONAL STUDIES ON CARCINOMA IN MALE RATS

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ABSTRACT

Purslane (Portulaca oleracea L., Portulacaceae) has been traditionally used in folk medicine to afford protection against liver injury, although its actual efficacy remains uncertain. Objective: To investigate the hepatoprotective and antioxidant effect of purslane seeds extract against hepatocellular carcinoma (HCC) induced by Diethylnitrosamine (DEN). Materials and Methods: A total of 50 male Sprague–Dawley rats were randomly divided into five groups (10 rat/group) analyzed for a total experimental period 8 weeks. The first group was maintained as normal control, second group were treated with Purslane seeds extract (50 mg/kg of body weight) daily. Rats of remain groups were injected intraperitoneally with freshly dissolved in sterile 0.9% saline DENA (200 mg/kg body weight) and two weeks later, they received a subcutaneous injection of CCl₄ (3 ml/kg body weight/week) for 6 weeks. Rats of HCC + Purslane group were injected orally with Purslane seeds extract at a daily dose level of (50 mg/kg of body weight) two weeks after DENA injection while in Purslane + HCC group were injected orally with Purslane seeds extract before DENA injection. Serum and hepatic liver enzymes, blood reduced glutathione (GSH) and malondialdehyde (MDA) in red blood cells were determined and superoxide dismutase (SOD) enzyme also assayed. Caspases-3 was analyzed using flow cytometric analysis. Results: Serum liver enzymes, AFP, level of MDA and liver caspase 3 were increased in HCC group, while hepatic enzymes, tissue SOD and blood glutathione level were reduced. In groups treated with purslane seeds extract AFP, liver caspase 3, level of MDA and activity of liver enzymes were reduced, and the activity of the antioxidant parameters were increased. Conclusions: Purslane seeds extract enhanced innate antioxidant activity and ameliorate the DENA-induced HCC and therefore can be used as a hepatoprotective drug in the future.

KEYWORDS:

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Purslane, antioxidant, hepatoprotective drugs, DENA, carbon tetrachloride, HCC.

1. INTRODUCTION

Liver plays a key role in various pathological disorders such as fatty liver, hepatic virus infection, chemical hepatotoxins, and toxicity cases.[1, 2] Chronic liver diseases are worldwide health problems causing approximately 800,000 deaths per year.[2] Of these, liver fibrosis is caused by inflammation and the excessive accumulation of the extracellular matrix. Subsequently, cirrhosis occurs and can cause hepatocellular carcinoma (HCC). [3] HCC is the sixth most commonly diagnosed cancer with high mortality in the world.[4] The median survival period of HCC is less

than 12 months from diagnosis. [5] Several kinds of treatments might be beneficial for HCC, such as local ablative therapy, liver transplantation, resection, hepatic artery transcatheter treatment and surgical therapy. However, limited effective chemotherapy agent available for HCC patients has been found.[6] Thus, there is an urgent need to search for novel therapeutic strategies and agents. Several hepatotoxicants including polycyclic aromatic hydrocarbons, nitrosamines and carbon tetrachloride (CCl₄) are transformed into intermediate reactive oxygen species that have hepatotoxic effects in humans

and experimental animal models.[7] Carbon tetrachloride (CCl₄) is widely used as experimental model of liver damage. CCl₄ is activated in the liver to highly reactive trichloromethyl radicals that initiate the free radical-mediated lipid peroxidation of membrane phospholipids, causing functional and morphological changes in the cell membrane, which stimulate hepatotoxicity, fibrosis[8], cirrhosis and HCC in animal species.[9] Moreover, *Portulaca oleracea* L, is commonly known as purslane. It is a warm climate, annual and green shoot.[15] Research indicates that purslane offers better nourishment than the major cultivated vegetables due to its shoot that is a rich source of X9-3-fatty acids, α -tocopherols, ascorbic acid, β -carotene and glutathione.[15] These features contribute to the anti-oxidative properties of purslane which derive from the following pharmacologically active substances, including: 28% flavonoids; 8% terpenoids; 6–12% organic acids; and > 0.5% proanthocyanidins defined as flavonoid-based polymers.[16] Purslane is effective as an antioxidant agent as well as providing nourishment for the liver.[15] Other authors reported that the purslane contains many compounds, including alkaloids, omega-3 fatty acids, coumarins, flavonoids, polysaccharide, cardiac glycosides, anthraquinone glycosides and containing β -sitosterol.[17] Based on these issues and concerns, the present study was designed to investigate the preventive effect leaves extract of purslane on DENA mediated liver damage.

2. MATERIALS AND METHODS

2.1. Chemicals

Diethylnitrosamine (DENA) and carbon tetra chloride (CCl₄) were purchased from Sigma Chemical Co. DENA was freshly dissolved in sterile 0.9% saline and given to rats at a single dose of 200 mg/kg body weight.[18] CCl₄ was given to rats at a dose of 3 ml/kg/body weight/week to initiate the effect of DENA. [19, 20]

2.2. Plant materials

The seeds were dried in the shade and powdered by an electric grinder then, they were exhaustively extracted with 80% ethanol. The solvent was removed by evaporation under reduced pressure using Buchi Rotary Evaporator.[21]

2.3. Experimental animals

Fifty male Sprague–Dawley (SD) rats weighing 180-200 g were allowed acclimatization period of 14 days in an

ambient temperature of $25 \pm 32^{\circ}\text{C}$ on light/dark cycle of 12/12 hours. Animals received human care and the present study complies with the instruction's guidelines. The local committee approved the design of the experiments and the protocol conforms to the guidelines of the National Institutes of Health (NIH). All rats were kept in clean polypropylene cages and administered food and water ad libitum.

2.4. Hepatocellular rat's model

Experimental hepatocellular carcinoma rats were subjected by a single intraperitoneal injection of freshly prepared DENA (200 mg/kg body weight), then 2 weeks later received a subcutaneous injection of CCl₄ once every week (3 mL/kg body weight) for 10 weeks to promote the carcinogenic effect of DENA.[20]

2.5. Experimental design

At the end of the acclimatization period, the experimental 50 male Sprague–Dawley (SD) rats were divided into five groups (10 rats/group) analyzed for a total experimental period 8 weeks as follows: Group 1(Control): rats of this group did not receive any treatments; Group 2 (Purslane): rats of this group are treated with Purslane seeds extract at a daily dose level of (50 mg/kg body weight); Group 3 (HCC): rats treated with a single intraperitoneal injection of freshly dissolved in sterile 0.9% saline DENA (200 mg/kg body weight) and two weeks later, they received a subcutaneous injection of CCl₄ (3 ml/kg/week) for 6 weeks to promote the carcinogenic effect of DENA; Group 4 (HCC + Purslane): rats were injected orally with Purslane seeds extract at a daily dose level of (50 mg/kg/kg) two weeks after DENA injection and Group 5 (Purslane +

At the same time, a certain weight of liver tissue from each rat was washed with normal saline and then it was homogenized in ice cold phosphate buffer (50 mM, pH 7.5) and the resultant homogenate (10%, w/v) was centrifuged at 12000 rpm for 20 min at 4°C in a cooling centrifuge then the supernatant was collected and stored at -20°C for subsequent biochemical assays which include SOD and catalase.

2.7. Biochemical analysis

Alpha-fetoprotein (AFP) level was estimated by immunoenzymatic colorimetric method according to Acosta, (1983).[22] Alanine transaminase (ALT) and Aspartate transaminase (AST) activities in serum were assayed according to the method of Reitman and Frankel (1957)[23] while serum alkaline phosphatase (ALP) and GGT were determined by using kits obtained from Biodiagnostics according to the method of Belfield and Goldberg (1971)[24] and Szasz (1969)[25], respectively. Assay of superoxide dismutase (SOD) activity in liver was determined according to the method described by Nishikimi et al., (1972).[26] Assay of liver reduced glutathione (GSH) concentration in blood was done by the method of Beutler et al., (1963).[27] Lipid peroxidation was evaluated on the basis of Malondialdehyde (MDA) level, MDA in RBCs was determined using the method described by Stocks and Donnandy, (1971).[28] Other chemicals used throughout this investigation were of the highest analytical grade available.

2.8. Determination of Caspases-3.

Fresh liver tissue specimens were transported to the laboratory in isotonic saline and prepared according to the method described by Tribukait et al., (1975)[29] For the caspases-3 assay, liver cell suspensions were prepared with a PBS/BSA buffer. One hundred

microliters of the cell suspension were added to 7 µl of antibody (FITC Rabbit Anti- Active Caspase-3 (CPP32; Yama; Apopain, BD Bioscience), mixed well and incubated for 30 min at room temperature, the cells were washed with BD Perm/Wash (BD Bioscience), centrifuged at 400 rpm for 5 min and the supernatant was discarded, the cells finally were re-suspended in BD Perm/Wash and analyzed using flow cytometry (Belloc et al., 2000).[30]

2.9. Statistical analysis

The data were expressed as mean $\pm$ S.D and statistical analysis were performed by using ANOVA followed by post hoc test (Tukey). It is a parametric statistical analysis that compares between-and within-groups variance to measure differences between two or more groups. The P value less than 0.05 was considered as statistically significant. The data analysis was done with SPSS version 22.

3. RESULTS

3.1. Serum liver enzymes

As shown in Table 1, there was no significant difference ($P > 0.05$) in activities of serum AST, ALT, ALP and GGT enzymes in Purslane rats compared with control group. There were significant elevations of serum AST, ALT, ALP and GGT enzymes activities ($p < 0.0001$) in HCC rats compared with control group. In addition, there was decrease activity of serum AST, ALT, ALP and GGT in rats Purslane $\pm$ HCC (AST: $P < 0.0001$; ALT: $P < 0.001$; ALP: $P < 0.001$ and GGT: $P < 0.05$) and HCC $\pm$ Purslane (AST: $P < 0.0001$; ALT: $P < 0.001$; ALP: $P < 0.001$ and GGT: $P < 0.05$), respectively compared with HCC but the decrease in HCC $\pm$ Purslane more than Purslane $\pm$ HCC rats group with no significant difference between treated groups.

Table 1: Effect of Purslane administration on serum liver enzymes in rats infected with HCC alone and those treated with Purslane before/after HCC.

3.2. Hepatic liver enzymes

Table 2: Effect of Purslane administration on hepatic liver enzymes in rats infected with HCC alone and those treated with Purslane before/after HCC infection.

Groups	Parameters (U/g wet tissue)			
	AST	ALT	ALP	GGT
Control	22.8±4.0	56.4±17.8	30.8±18.5	23.5±4.3
Purslane	24.1±5.5	59.0±13.4	29.8±7.2	24.6±3.4
HCC	17.2±4.4	41.3±10.7	51.1±10.6	31.5±7.5
Purslane + HCC	19.5±3.8	50.3±10.0	36.7±16.0	25.8±6.7
HCC + Purslane	20.0±6.0	52.6±15.2	40.2±14.1	25.0±6.8
HCC vs Control	< 0.001	< 0.001	< 0.0001	< 0.01
HCC vs Purslane + HCC	< 0.05	< 0.05	< 0.05	> 0.05
HCC vs HCC + Purslane	< 0.05	< 0.05	< 0.05	< 0.05

Data are expressed as mean ± S.D. (n = 10 in each group). HCC: hepatocellular carcinoma. ALT: Alanine transaminase, AST: Aspartate transaminase, ALP: alkaline phosphatase and GGT: gamma-glutamyl transferase.

P > 0.05 considered not significant, * P < 0.05 considered significant, **P < 0.01 considered high significant, and ***P < 0.0001 considered extremely significant and (ns) not significant.

As shown in Table 2, there was no significant difference (P > 0.05) in activities of hepatic AST, ALT, ALP and GGT enzymes in Purslane rates compared with control group. There were significant elevations of hepatic AST, ALT, ALP and GGT enzymes activities (p < 0.0001) for all parameters compared with control group. In addition, there was decrease activity of hepatic AST, ALT, ALP and GGT in rats Purslane ± HCC (AST: P < 0.0001; ALT: P < 0.001; ALP: P < 0.001 and GGT: P < 0.05) and HCC ± Purslane (AST: P < 0.0001; ALT: P < 0.001; ALP: P < 0.001 and GGT: P < 0.05), respectively compared with HCC but the decrease in HCC ± Purslane more than Purslane ± HCC rats group.

3.3. Antioxidants profile

Table 3: Effect of Purslane administration on oxidate stress and antioxidant biomarkers in the liver of control and different treated rat groups (means ± SD).

Groups	SOD (% inhibition)	GSH (mMol/g Hb)	GST (mMol/gHb)	MDA (mg/dL)
Control	57.5±6.4	242.0±25.2	0.34±0.08	7.3±0.9
Purslane	55.2±6.5			

group compared with control group. In addition, there was decrease ($P < 0.0001$) in rats treated with Purslane before and after HCC infection compared with HCC alone but in rats treated with Purslane seeds extract after more decrease than before.

3.5. Flow cytometric hepatic caspase 3

Figure 1 showed hepatic apoptotic marker % (caspase 3) in control and different treated rat groups. In the case

of HCC rats group, there was a significant increase in liver caspase 3 compared with control rats group. In contrast the treatment of HCC rats showed a significant decrease in liver caspase 3 but not significant in the case of Purslane administration in caspase 3 if comparing to HCC rats group

Fig. 1: Flow cytometric hepatic caspase-3 (as an apoptotic marker) analysis showing in control and different treated rat groups.

4. DISCUSSION

There has been growing interest in dietary bioactive compounds obtained from natural sources which have a therapeutic effect against various diseases including chemoprotective properties against cancer.[31] HCC is a common disease, being the third leading cause of death worldwide.[32] The current study suggests that treatment with DENA and CCl₄ is a good model for the induction of HCC in rats.[33] In the present study, we observed elevation of serum AST, ALT and ALP in HCC rats. These findings may be due to damage to hepatocytes caused by exposure to DENA resulting in hepatic dysfunction and subsequent leakage of these enzymes from the neoplastic cell into circulation.[34] Elevated serum aminotransferase activity is more specific for liver injury due to damage to the liver cell membrane.[35] As well, alkaline phosphatase is used as a specific tumor marker for making diagnoses in the early detection of cancer.[35] This enzyme is involved in the transport of metabolites across cell membranes, in protein synthesis, secretory activities and glycogen metabolism. It is a membrane-bound enzyme and its

alteration is likely to affect the membrane permeability that produces derangement in the transport of metabolites.[37] The observed increases of serum and liver ALP in HCC rat groups may be due to altered gene expression.[38] Upon treatment with purslane extract, the levels of these markers were restored to near normal or were only slightly elevated, indicating protection against liver damage. Such a restoration of increased serum levels of hepatic enzymes to the normal range reflects protection against hepatic damage caused by hepatotoxins.[39] The obtained data were in line with those of Venukumar and Latha, (2002)[40] whom mentioned that the raise in the activities of ALT, AST and ALP in rats' serum was a sign of hepatocellular damage of liver similar to acute viral hepatitis. On the other hand, Ahmida (2010) [41] reported that the hepatoprotective effective effect of purslane was due to the phytochemical present in it, included omega-3-fatty acids, Bcarotene, flavonoids and alkaloids.

The hepatotoxicity of CCl₄ is due to reductive dehalogenation products, such as trichloromethyl

($\text{CCl}_3\cdot$) and trichloromethyl peroxy ($\text{CCl}_3\text{O}_2\cdot$) radicals (Brattin et al., 1985).[42] These radicals can bind to proteins and lipids or remove a hydrogen atom from unsaturated fatty acids, thereby initiating lipid peroxidation and contributing to liver injury.[43] Hepatocyte injury initiates the activation of Kupffer cells which secrete potent mediators of the early inflammatory response, such as reactive oxygen species (ROS), especially superoxide anions that accounted for the formation of peroxynitrites and hydrogen peroxides (H_2O_2) therefore oxidative stress can be occur.[44] The antioxidants could attenuate this oxidative damage caused by free radicals indirectly by enhancing natural defenses of cell and/or directly by scavenging the free radicals.[45] Antioxidants such as superoxide dismutase (SOD) can scavenge the superoxide anions whereas the glutathion reduced (GSH) is responsible to remove H_2O_2 through the action of glutathione peroxidase.[46] Also, H_2O_2 is consumed by the action of catalase.[47] In this sense, the antioxidant action plays an important role in protecting against CCl_4 -induced liver injury.

In the current study, the HCC rats group suffered from severe oxidative stress, achieved by elevation of MDA level and depletion of antioxidant enzymes. This may be due to the conversion of cellular poly-unsaturated fatty acids to the toxic product MDA which has a cytotoxicity and inhibitory action on cellular protective enzymes.[48] HCC caused by carcinogenic DENA generally reflects instability of liver metabolism associated with free radicals species (ROS) generation, which leads to oxidative stress and alterations in antioxidant defense mechanisms. .

Moreover, HCC causes depletion of SOD and GSH contents. Such studies support the current findings, as the current study showed a significant decrease in the activities of antioxidant enzyme in the liver of animals treated with carcinogen. Antioxidants are substances that either directly or indirectly protect cells against adverse effects of xenobiotics, drugs, carcinogens and toxic radical reactions. The observed decrease in SOD activity in liver suggests the inactivation of antioxidant enzymes; this is possibly due to increased superoxide radical production or to an inhibition by H_2O_2 as a result of corresponding decrease in the activity of catalase which selectively degrades H_2O_2 . The decreased GSH and SOD observed in the HCC group of rats may be due to accumulation of lipid peroxidation that was seen to increase during carcinogenesis. In conclusions, Purslane

seeds extract enhanced innate antioxidant activity and ameliorate the DENA-induced liver injury and therefore can be used as a hepatoprotective drug in the future.

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