

Gangadhari Harika¹ and Gade Sammaiah^{*2}

^{1, 2} University College of Pharmaceutical Sciences (UCPSC) Kakatiya University, Hanamkonda, Warangal, Telangana.

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Abstract

A new, simple, rapid, precise, accurate and reproducible RP-HPLC method for estimation of Molnupiravir in bulk form and marketed formulation. Separation of Molnupiravir was successfully achieved on a Develosil ODS HG-5 RP C18, 5µm, 15cmx4.6mm i.e., column in an isocratic mode of separation utilizing Methanol: Phosphate buffer (0.02M, pH-3.6) in the ratio of 45:55% v/v at a flow rate of 1.0 mL/min and the detection was carried out at 255nm. The method was validated according to ICH guidelines for linearity, sensitivity, accuracy, precision, specificity and robustness. The response was found to be linear in the drug concentration range of 12-28mcg/mL for Molnupiravir. The correlation coefficient was found to be 0.9995 for Molnupiravir. The LOD and LOQ for Molnupiravir were found to be 5.004μ g/mL and 15.164μ g/mL respectively. The proposed method was found to be good percentage recovery for Molnupiravir, which indicates that the proposed method is highly accurate. The specificity of the method shows good correlation between retention times of standard solution with the sample solution. Therefore, the proposed method specifically determines the analyte in the sample without interference from excipients of pharmaceutical dosage forms. **Keywords**

Molnupiravir, RP-HPLC, Accuracy, Precision, Robustness, ICH Guidelines.

INTRODUCTION

Molnupiravir (EIDD-2801, MK-4482) is the isopropyl ester prodrug of [N4-hydroxycytidine]. With improved oral bioavailability in non-human primates, it is hydrolyzed in vivo, and distributed into tissues where it becomes the active 5'-triphosphate form. The active drug incorporates into the genome of RNA viruses, leading to an accumulation of mutations known as viral error catastrophe. Recent studies have shown Molnupiravir¹ inhibits replication of human and bat coronaviruses, including SARS-CoV-2, in mice and human airway epithelial cells. A [Remdesivir] resistant mutant mouse hepatitis virus has also been shown to have increased sensitivity to N4-hydroxycytidine. Molnupiravir was granted approval by the UK's Medicines and Health products Regulatory Agency (MHRA) on 4 November 2021 to prevent severe outcomes such as hospitalization and death due to COVID-19 in adults. Molnupiravir was also granted emergency use authorization by the FDA on December 23, 2021; however, it is not yet fully approved. Molnupiravir² is an orally bioavailable prodrug of EIDD-1931, the synthetic ribonucleoside derivative N4-hydroxycytidine and ribonucleoside



analog, with potential antiviral activity against a variety of RNA viruses. Upon oral administration, Molnupiravir, being a prodrug, is metabolized into its active form EIDD-1931 and converted into its triphosphate (TP) form. The TP form of EIDD-1931 is incorporated into RNA and inhibits the action of viral RNA-dependent RNA polymerase. This results in the

termination of RNA transcription and decreases viral RNA production, and viral RNA replication. The IUPAC Name of Molnupiravir³ is [(2R, 3S, 4R, 5R)-3, 4dihydroxy-5-[4-(hydroxy amino)-2-oxopyrimidin-1yl] oxolan-2-yl] methyl 2-methyl propanoate. The Chemical Structure of Molnupiravir is as following.



Fig-1: Chemical Structure of Molnupiravir

After performing an extensive literature review³¹⁻³², an attempt was made to develop a smooth plain sailing, unambiguous, valid, speedy, and decisive

strategy for estimating Molnupiravir in bulk form and marketed pharmaceutical dosage form by using RP-HPLC.

MATERIALS AND METHODS

Table-1: Instruments used.				
S.No.	Instruments and Glass wares	Model		
		WATERS Alliance 2695 separation		
1	HPLC	module, Software: Empower 2, 996		
		PDA detector.		
2	pH meter	Lab India		
3	Weighing machine	Sartorius		
4	Volumetric flasks	Borosil		
5	Pipettes and Burettes	Borosil		
6	Beakers	Borosil		
7	Digital ultra sonicator	Labman		

Table-2: Chefnicals used.						
S.No.	Chemical	Brand names				
1	Molnupiravir	Local Market				
2	Water and Methanol for HPLC	LICHROSOLV (MERCK)				
3	Acetonitrile for HPLC	Merck				
4	Ethanol	Sd fine-Chem ltd; Mumbai				
5	DMSO	Sd fine-Chem ltd; Mumbai				
6	DMF	Sd fine-Chem ltd; Mumbai				
7	Orthophosphoric Acid	Sd fine-Chem ltd: Mumbai				

HPLC METHOD DEVELOPMENT:

Preparation of Standard Solution:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7ml of Methanol and sonicate to dissolve and removal of air completely and make volume up to the mark with the same Methanol.

Further pipette 0.1ml of the above Molnupiravir stock solutions into a 10ml volumetric flask and dilute up to the mark with Methanol.

Preparation of Sample Solution:

Twenty capsules were taken, and the average weight was calculated as per the method prescribed in I.P. The weighed tablets were finally powdered and triturated well. A quantity of powder of Molnupiravir equivalent to 10mg were transferred to clean and dry 10 ml volumetric flask and 7 ml of HPLC grade methanol was added and the resulting solution was sonicated for 15 minutes. Make up the volume up to 10 ml with same solvent. Then 1 ml of the above



solution was diluted to 10 ml with HPLC grade methanol. One ml (0.1 ml) of the prepared stock solution diluted to 10 ml and was filtered through membrane filter (0.45µm) and finally sonicated to degas.

Procedure:

Inject the samples by changing the chromatographic conditions⁴ and record the chromatograms, note the conditions of proper peak elution for performing validation parameters as per ICH guidelines¹³⁻¹⁴.

Mobile Phase Optimization:

Initially the mobile phase⁵ tried was Methanol and Methanol: Water with varying proportions. Finally, the mobile phase was optimized to Methanol and Phosphate buffer (0.02M, pH-3.6) in proportion 45:55% v/v.

Optimization of Column:

The method was performed with various C18 columns like, X- bridge column, Xterra, and C18 column. Develosil ODS HG-5 RP C18, 5µm, 15cmx4.6mm i.d. was found to be ideal as it gave good peak shape and resolution at 1.0ml/min flow.

Preparation of Buffer and Mobile Phase:

Preparation of Potassium Dihydrogen Phosphate (KH2PO4) Buffer (0.02M) (pH-3.6):

Dissolve 2.72172g of potassium dihydrogen phosphate in 1000 ml HPLC water and adjust the pH 3.6 with diluted orthophosphoric acid. Filter and sonicate the solution by vacuum filtration⁶ and ultrasonication.

Preparation of Mobile Phase:

Accurately measured 450 ml (45%) of Methanol and 550 ml of Phosphate buffer (55%) were mixed and degassed in digital ultra sonicater for 15 minutes and then filtered through 0.45 μ filter under vacuum filtration.

Diluent Preparation:

The Mobile phase was used as the diluent.

METHOD VALIDATION PARAMETERS

System Suitability

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Molnupiravir stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

Procedure:

The standard solution was injected for five times and measured the area for all five injections in HPLC. The %RSD for the area of five replicate injections was found to be within the specified limits.

Specificity:

Preparation of Standard Solution:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Molnupiravir stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Preparation of Sample Solution:

Weight 10 mg equivalent weight of Molnupiravir sample into a 10mL clean dry volumetric flask and add about 7mL of Diluent and sonicate to dissolve it completely and make volume up to the mark with the same solvent.

Further pipette 0.1ml of Molnupiravir above stock solution into a 10ml volumetric flask and dilute up to the mark with diluent.

Procedure:

Inject the three replicate injections of standard and sample solutions and calculate the assay⁷ by using formula:

%ASSAY =

Weight of standard Dilution of Sample area sample Purity Weight of tablet

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		x	×			>	(10	C
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Standard area Dilution of standard Weight of sample 100 Label claim

Linearity and Range:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Preparation of Level – I (12ppm of Molnupiravir):

Take 0.12ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Preparation of Level – II (16ppm of Molnupiravir):

Take 0.16ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Preparation of Level – III (20ppm of Molnupiravir):

Take 0.2ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Preparation of Level – IV (24ppm of Molnupiravir):

Take 0.24ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.



Preparation of Level – V (28ppm of Molnupiravir):

Take 0.28ml of stock solution in to 10ml of volumetric flask and make up the volume up to mark with diluents and sonicate the solution for bubble entrapment using ultrasonicator.

Procedure:

Inject each level into the chromatographic system and measure the peak area.

Plot a graph of peak area versus concentration (on Xaxis concentration and on Y-axis Peak area) and calculate the correlation coefficient.

Precision

Repeatability

Preparation of Molnupiravir Product Solution for Precision:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Molnupiravir stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Procedure:

The standard solution was injected for six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits.

Intermediate Precision:

To evaluate the intermediate precision (also known as Ruggedness) of the method, Precision was performed on different days by maintaining same conditions.

Procedure:

Analyst 1:

The standard solution was injected for six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits.

Analyst 2:

The standard solution was injected for six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits.

Accuracy:

For Preparation of 80% Standard Stock Solution:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.08ml of the above Molnupiravir stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

For Preparation of 100% Standard Stock Solution:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.1ml of the above Molnupiravir stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

For Preparation of 120% Standard Stock Solution:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.12ml of the above Molnupiravir stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

Procedure:

Inject the Three replicate injections of individual concentrations (80%, 100%, 120%) were made under conditions. the optimized Recorded the chromatograms and measured the peak responses. Calculate the Amount found and Amount added for Molnupiravir and calculate the individual recovery⁸ and mean recovery values.

LIMIT LIMIT OF DETECTION AND OF **QUANTIFICATION (LOD & LOQ):**

Preparation of 5.004µg/ml Solution (For LOD):

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.05004ml of the above Molnupiravir stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Preparation of 15.164µg/ml Solution (For LOQ):

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7ml of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)

Further pipette 0.15164ml of the above Molnupiravir stock solutions into a 10ml volumetric flask and dilute up to the mark with diluents.

Robustness:

The analysis was performed in different conditions to find the variability of test results. The following conditions are checked for variation of results. .

For Preparation of Standard Solution:

Accurately weigh and transfer 10 mg of Molnupiravir working standard into a 10ml of clean dry volumetric flasks add about 7mL of Diluents and sonicate to dissolve it completely and make volume up to the mark with the same solvent. (Stock solution)





Further pipette 0.1ml of the above Molnupiravir stock solution into a 10ml volumetric flask and dilute up to the mark with diluents.

Effect of Variation of Flow Conditions:

The sample was analyzed at 0.9 ml/min and 1.1 ml/min instead of 1ml/min, remaining conditions are same. 20μ l of the above sample was injected and chromatograms were recorded.

Effect of Variation of Mobile Phase Organic Composition:

The sample was analyzed by variation of mobile phase i.e., Methanol: Phosphate Buffer was taken in the ratio and 50:50, 40:60 instead (45:55), remaining conditions are same. 20µl of the above sample was injected and chromatograms were recorded.

Forced Degradation Studies

The purpose of stability testing⁹⁻¹² is to provide evidence on how the quality of a drug substance or drug product varies with time under the influence of a variety of environmental factors such as temperature, humidity, and light, and to establish a re-test period for the drug substance or a shelf life for the drug.

RESULTS AND DISCUSSION Selection of Wavelength:

The detection wavelength was selected by dissolving the drug in mobile phase to get a concentration of $10\mu g/ml$ for individual and mixed standards. The resulting solution was scanned in U.V range from 200-400nm. The UV spectrum¹⁵ of Molnupiravir was obtained and the Molnupiravir showed absorbance's maxima at 255nm. The UV spectra of drug are follows:



Fig-2: UV Spectrum of Molnupiravir

Observation: While scanning the Molnupiravir solution we observed the maxima at 255nm. The UV spectrum has been recorded on T60-LAB INDIA make UV-Vis spectrophotometer model UV-2450.

Method Development

Several concurrent trials developed the proposed method to establish the preferred chromatographic conditions, which would be helpful to conduct a complete validation study. The optimal conditions considered for mobile phase were 45:55% v/v ratio of Methanol and 0.02M potassium dihydrogen orthophosphate buffer at pH 3.6 and Develosil ODS HG-5 RP C18, 5^[2]m, 15cmx4.6mm i.e., Column¹⁶ were used, which gave a sharp symmetric peak, minimum tailing factor with short run time for Molnupiravir at a flow rate of 1.0 mL/min and detection wavelength 255 nm was optimized. A sample of 20μL was injected into HPLC System¹⁷ and the different optimization of chromatographic parameters is depicted in Table 3. The retention time for Molnupiravir was found to be 3.254 minutes (Figure 3).

Optimized Chromatographic Method:

Table-3: Optimized Chromatographic Conditions

Mobile phase	Methanol: Phosphate buffer (0.02M, pH-3.6) = 45:55 v/v				
Column	Develosil ODS HG-5 RP C18, 5µm, 15cmx4.6mm i.d.				
Column Temperature	Ambient				
Detection Wavelength	255 nm				
Flow rate	1.0 ml/ min.				
Run time	07 min.				
Temperature of Auto sampler	Ambient				
Diluent	Mobile Phase				
Injection Volume	20µl				
Type of Elution	Isocratic				





Fig-3: Optimized Chromatographic Condition

Conclusion: This trial shows proper plate count, peak and baseline in the chromatogram. It's Pass all system suitability parameters. So, it's an optimized chromatogram.

Validation of Analytical Method

System Suitability: System suitability testing is an integral part of many analytical procedures. The tests

are based on the concept that the equipment, electronics, analytical operations and samples to be analyzed constitute an integral system that can be evaluated as such. The following system suitability test parameters¹⁸ were established. The data are shown in Table-4 & 5.

Table-4. Data of System Suitability Test									
S.No.	Injection No.	RT	Area	USP Plate Count	USP Tailing				
1	Injection 1	3.253	284568	7368	1.26				
2	Injection 2	3.254	285684	7295	1.25				
3	Injection 3	3.215	283659	7346	1.27				
4	Injection 4	3.297	284754	7394	1.29				
5	Injection 5	3.253	283695	7425	1.25				
6	Injection 6	3.213	284578	7385	1.27				
Mean			284489.7	7368.833	1.265				
S. D			752.5617						
%RSD			0.26453						

Table-4	Data	of Sv	vstem	Suitability	/ Test
1 0 0 1 5 - 4.	υαια	01.31	JSLEIII	Juitability	/ 1031

Table-5: System Suitability Results for Molnupiravir (Flow rate)						
	S.No.	Parameter	Limit	Result		
	1	Asymmetry	$T \leq 2$	Molnupiravir = 0.12		
	2	Theoretical plate	N > 2000	Molnupiravir = 7258		
	3	Tailing Factor	(Tf) < 2	Molnupiravir = 1.25		

Specificity:

Specificity¹⁹ can be determined by comparing the chromatograms obtained from the drugs with the chromatogram obtained from the blank solution. Blank solution was prepared by mixing the excipients in the mobile phase without drug. Drug solutions were prepared individually and the sample containing three drugs was also prepared. Now these mixtures were filtered by passing through 0.45 μ membrane filter before the analysis. In this observation no excipient peaks were obtained near the drug in the study run time. This indicates that the proposed method was specific. The chromatograms representing the peaks of blank, Molnupiravir and the sample containing the three drugs were shown in following figures respectively.

Observation: In this test method blank, standard solutions were analyzed individually to examine the interference. The above chromatograms show that the active ingredient was well separated from blank and their excipients and there was no interference of blank with the principal peak. Hence the method is specific.

Linearity: To evaluate linearity²⁰, serial dilution of analyte was prepared from the stock solution was diluted with mobile phase to get a series of concentration ranging from $0-28\mu$ g/ml for Molnupiravir. The prepared solutions were filtered through Whatman filter paper (No.41). From these solutions, 20µl injections of each concentration were injected into the HPLC system and chromatographed under the optimized conditions. Calibration curve²¹



was constructed by plotting the mean peak area (Yaxis) against the concentration (X-axis).

Plotting of Calibration Graphs: The resultant areas of linearity peaks are plotted against Concentration.



Fig-4: Standard Curve for Molnupiravir

Observation: Linearity range²² was found to be 0-28µg/ml for Molnupiravir. The correlation coefficient was found to be 0.9995, the slope was found to be

55283 and intercept was found to be 12871 for Molnupiravir.

а	able-6: Linearity Readings for Molnupira						
	CONC.(µg/ml)	MEAN AUC (n=6)					
	0	0					
	12	690316					
	16	910621					
	20	1121057					
	24	1328903					
	28	1554666					

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Linearity Plot:

The plot of Concentration (x) versus the Average Peak Area (y) data of Molnupiravir is a straight line. Y = mx + c

Slope (m) = 55283

Intercept (c) = 12871

Correlation Coefficient (r) = 0.9995

Acceptance/Validation Criteria: The response linearity is verified if the Correlation Coefficient is 0.99 or greater.

Conclusion: Correlation Coefficient (r) is 0.99, and the intercept is 12871. These values meet the validation criteria.

Accuracy:

Inject the three replicate injections of individual concentrations (80%, 100%, 120%) were made under conditions. Recorded the optimized the chromatograms and measured the peak responses. Calculate the Amount found, and Amount added for Molnupiravir and calculate the individual recovery and mean recovery values²³. Accuracy at different concentrations (80%, 100%, and 120%) was prepared and the % recovery was calculated.

	Concent	ration (µg/ml)					
Sample ID	Conc.	Conc.	Peak Area	%Recovery of Pure drug	Statistical Analysis		
	Found	Recovered					
S1: 80 %	8	8.064107	458679	99.867	Mean= 100.4113%		
S2:80 %	8	7.843532	446485	100.637	S.D. = 0.473694346		
S₃ : 80 %	8	8.19449	465887	100.73	% R.S.D.= 0.471753		
S4: 100 %	10	9.892661	559767	99.41	Mean=		
S5 : 100 %	10	9.978655	564521	100.868	100.6646667%		

Table-7. Accuracy results of Molnuniravir





S ₆ : 100 %	10	10.19623	576549	101.716	S.D. = 1.166369295
					R.S.D.= 1.158667
S7: 120 %	12	11.85907	668476	99.878	Mean= 100.4637%
S ₈ :120 %	12	12.16785	685546	100.69	S.D. = 0.51154309
S ₉ : 120 %	12	12.18644	686574	100.823	% R.S.D. = 0.509181

Observation: The mean recoveries were found to be 100.411, 100.664 and 100.463% for Molnupiravir. The limit for mean % recovery is 98-102% and as both the values are within the limit, it can be said that the proposed method was accurate.

Precision: The precision²⁴ of each method was ascertained separately from the peak areas obtained by actual determination of six replicates of a fixed

amount of drug Molnupiravir. The percentage relative standard deviations were calculated for Molnupiravir are presented in Table-8.

i) Repeatability

Obtained Six (6) replicates of 100% accuracy solution as per experimental conditions. Recorded the peak areas and calculated % RSD.

Table-8: Repeatability Results of Molnupiravir					
HPLC Injection Replicates	AUC for Molnupiravir				
Replicate – 1	285479				
Replicate – 2	284571				
Replicate – 3	286954				
Replicate – 4	283261				
Replicate – 5	285964				
Replicate – 6	284259				
Average	285081.3				
Standard Deviation	1318.666				
% RSD	0.462558				

Observation: The repeatability study which was conducted on the solution having the concentration of about 20μ g/ml for Molnupiravir (n=6) showed a RSD of 0.462558% for Molnupiravir. It was concluded that the analytical technique showed good repeatability.

ii) Intermediate Precision / Ruggedness

To evaluate the intermediate precision²⁵ (also known as Ruggedness) of the method, Precision was performed on different days by maintaining same conditions.

Procedure:

Analyst 1: The standard solution was injected six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits. **Analyst 2:**

The standard solution was injected six times and measured the area for all six injections in HPLC. The %RSD for the area of six replicate injections was found to be within the specified limits²⁹⁻³⁰.

Intra Day (Day-1)/Analyst-1:

	Table-9: Results of Ruggedness for Molnupiravir (Analyst-1)								
S.No.	Peak Name	RT	Peak Area	Theoretical Plates	Tailing Factor				
1	Molnupiravir	3.253	284568	7368	1.26				
2	Molnupiravir	3.254	285684	7295	1.25				
3	Molnupiravir	3.215	283659	7346	1.27				
4	Molnupiravir	3.204	286598	7457	1.22				
5	Molnupiravir	3.202	287965	7635	1.29				
6	Molnupiravir	3.297	285698	7459	1.28				
Mean			285695.3						
Std. Dev.			1508.898						
% RSD			0.528149						

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Table-10: Results of Ruggedness for Molnupiravir (Analyst-2)						
S.No.	Peak Name	RT	Peak Area	Theoretical Plates	Tailing Factor	
1	Molnupiravir	3.297	294754	7394	1.29	
2	Molnupiravir	3.253	293695	7425	1.25	
3	Molnupiravir	3.213	294578	7385	1.27	
4	Molnupiravir	3.297	296534	7584	1.23	
5	Molnupiravir	3.210	296571	7745	1.24	
6	Molnupiravir	3.254	298698	7658	1.25	
Mean			295805			
Std. Dev.			1819.334			
% RSD			0.615045			

Inter Day (Day -2/Analyst-2)

Robustness:

Robustness²⁶ is defined as the capacity of that method to be unaffected by even small deliberate changes that occur in the method parameters. The evaluation of robustness of a method is done by

varying the chromatographic parameters such as pH, temperature, flow rate, mobile phase proportions change, ionic strength etc., and determining any possible effect on the results obtained by that method.

|--|

Parameter used for Sample Analysis	Peak Area	Retention Time	Theoretical plates	Tailing factor
Actual Flow rate of 1.0 mL/min	283261	3.254	7258	1.25
Less Flow rate of 0.9 mL/min	315864	3.297	7569	1.29
More Flow rate of 1.1 mL/min	298542	3.212	7841	1.41
Less organic phase	279856	3.253	7965	1.27
More organic phase	306985	3.215	7458	1.28

LOD: The limit of detection (LOD) is the lowest concentration of analyte in a sample which can be detected, but not quantitated. LOD²⁷ is a limit test that specifies whether an analyte is above or below

a certain value. A signal-to-noise ratio of three-toone is used to determine LOD. L.O.D. = 3.3 (SD/S).

Where, SD = Standard deviation of the response S = Slope of the calibration curve

Table-12: Results of LOD				
LOD				
SD of Intercept	19518.16286			
Slope	55283			

Observation: The LOD was found to be $1.165 \mu g/ml$ for Molnupiravir.

LOQ: The Limit of Quantitation (LOQ) is defined as the lowest concentration of an analyte in a sample that can be determined with acceptable precision

and accuracy under the stated operational conditions of the method. A signal-to-noise ratio of ten-to-one is used to determine LOQ^{28} . L.O.Q. = 10 (SD/S)

Where, SD = Standard deviation of the response S = Slope of the calibration curve

Table-13: Results of LOQ				
LOQ				
SD of Intercept	19518.16286			
Slope	55283			

Observation: The LOQ was found to be $3.53 \mu g/ml$ for Molnupiravir.

Assay of Pharmaceutical Dosage form

Twenty tablets/Capsules were taken and the I.P. method was followed to determine the average

weight. Finally the weighed tablets are powdered and triturated well by using mortar and pestle. A quantity of powder which is equivalent to the 100mg of drugs were transferred to a clean and dry 100ml of volumetric flask and add 70 ml of mobile phase



and the resulted solution was sonicated for 15 minutes by using ultra sonicator, Then the final volume was make up to the mark with the mobile phase. The final solution was filtered through a selected membrane filter (0.45 μ m) and to sonicate to degas the mobile phase (Solvent system). From this above stock arrangement (1 ml) was exchanged to five distinctive 10 ml volumetric flagons and

ASSAY:

Assay %	=			
AT	WS	DT	Р	
	x -		-x	xx Avg. Wt = mg/tab
AS	DS	WΤ	100	

Where:

AT = Peak Area of drug obtained with test preparation

AS = Peak Area of drug obtained with standard preparation

WS = Weight of working standard taken in mg WT = Weight of sample taken in mg volume was made up to 10 ml with same dissolvable framework (Mobile stage).

The readied arrangements were infused in five repeats into the HPLC framework, and the perceptions were recorded.

A duplicate injection (Blank Solution) of the standard arrangement likewise infused into the HPLC framework and the chromatograms and peak zones were recorded and figured.

DS = Dilution of Standard solution

DT = Dilution of sample solution

P = Percentage purity of working standard

The assay was performed as explained in the previous chapter. The results which are obtained are following:

Table-14: Recovery	/ Data for	estimation	Molnupiravir	in Molflu	Capsule
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Brand name of Molnupiravir	Labelled amount of Drug (mg)	Amount (mg) found by the proposed method (n=3)	Assay %
Molflu Capsule (200mg) (Dr. Reddy's)	200mg	199.695mg	99.598%

Result & Discussion:

The amount of drug in Molflu Capsule was found to be 199.695 (± 0.789) mg/tab for Molnupiravir & % Purity²⁹ was 99.598 (± 0.695) %.

Forced Degradation Studies

Following protocol was strictly adhered to for forced degradation of Molnupiravir Active Pharmaceutical Ingredient (API). The API (Molnupiravir) was subjected to keep in some stress conditions in various ways to observe the rate and extent of degradation that is likely to occur in the course of storage and/or after administration to body. It is one type of accelerated stability studies of the drugs that

is used to help us to determining the total fate of the drug that is likely to happen after long time storage, within a very short time as compared to the real time or long-term stability testing. The different types of forced degradation pathways/studies are studied here are acid hydrolysis, basic hydrolysis, thermal degradation, and oxidative degradation.

Results of Degradation Studies: The results of the forced degradation studies³⁰ indicated the specificity of the developed method that has been developed. Molnupiravir were stable only in acidic, thermal, and basic stress conditions. The results of stability studies are given in the following Table-15.

Table-15: Results of Force Degradation Studies of M	Iolnupiravir API
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Stress Condition	Time	Assay of active	Assay of degraded	Mass Balance
Stress Condition	(hours)	substance	products	(%)
Acid Hydrolysis (0.1N HCl)	24Hrs.	91.326	8.674	100.00
Basic Hydrolysis (0.IN NaOH)	24Hrs.	83.215	16.785	100.00
Thermal Degradation (60 ⁰ C)	24Hrs.	90.311	9.689	100.00
UV (254nm)	24Hrs.	81.322	18.678	100.00
3% Hydrogen Peroxide	24Hrs.	73.514	26.486	100.00

SUMMARY AND CONCLUSION

To develop a precise, linear, specific & suitable stability indicating RP-HPLC method for analysis of Molnupiravir, different chromatographic conditions

were applied & the results observed are presented in previous chapters. Isocratic elution is simple, requires only one pump & flat baseline separation for easy and reproducible results. So, it was preferred



for the current study over gradient elution. In case of RP-HPLC various columns are available, but here Develosil ODS HG-5 RP C₁₈, 5µm, 15cmx4.6mm i.d. column was preferred because using this column peak shape, resolution and absorbance were good. Detection wavelength was selected after scanning the standard solution of drug over 200 to 400nm. From the U.V spectrum of Molnupiravir it is evident that most of the HPLC work can be accomplished in the wavelength range of 255 nm conveniently. Further, a flow rate of 1.0 ml/min & an injection volume of 20µl were found to be the best analysis. The result shows the developed method is yet another suitable method for assay which can help in the analysis of Molnupiravir in different formulations.

BIBLIOGRAPHY

- 1. https://go.drugbank.com/drugs/DB15661
- 2. https://pubchem.ncbi.nlm.nih.gov/compound/eidd-2801
- 3. https://en.wikipedia.org/wiki/Molnupiravir
- R. Snyder, J. Kirkland, L. Glajch, Practical HPLC 4. Method Development, john Wiley and sons' international publication, II Edn., 2011.
- 5. S. Ashutoshkar, Pharmaceutical Drug Analysis 2nd Edn, New Age International Private Limited Publishers, 452-474, 2005.
- 6. H. Beckett and J.B. Stenlake, Practical Pharmaceutical Chemistry, 4th End. C.B.S. Publishers and Distributors', New Delhi. 1-9, 157-167.
- 7. H.H. Williard, L.L. Merit, F.A. Dean, F.A. Settle, Instrumental Methods Of Analysis, 6th Edn, C.B.S. Publishers and Distributors, New Delhi.: 430-440, 495-504,529-545.
- B.K. Sharma, Instrumental Methods of Chemical 8. Analysis. GOEL Publishing House, Meerut: 286-300.
- Instant notes on analytical chemistry by D. Kealey 9. and P.J. Haines, © BIOS Scientific Publishers Limited, UK, 6-7, 2002.
- 10. Gurdeep R. Chatwal, Sham K. Anand, Instrumental methods of Chemical Analysis,5th edition, Himalaya Publishing House (Mumbai), P-2.566, 2005,
- 11. M. E. Swartz, Journal of liquid chromatography, 28(7/8), 1253-1263(2005).
- 12. Journal Analytical of Chromatography. Β. Technologies in the Biomedical and life Sciences. 2008 March 1; 863(2): 258-265. Published on Jan 18, 2008.
- Conference 13. International on Harmonization. Harmonized Tripartite Guideline. Validation of Analytical Procedures. Text and Methodology. Q2 (R1). November 2005.
- 14. International Conference on Harmonization (ICH). Validation of Analytical Methods: Definitions and Terminology. ICH Q2A. 1994.
- 15. J. M. Green, a practical guide to analytical method validation, anal. Chem. News & features, pp. 305a-309a, 1 May 1996.

- 16. P. A. Winslow and r. F. Meyer, defining a master plan for the validation of analytical methods, j. Validation technology, pp. 361-367, 1997.
- 17. Aoac peer-verified methods program, manual on policies and procedures, Arlington, Va., USA (1998).
- 18. R. Patil: J of Chromatographia, 67, 575, (2008).
- 19. Baht and Leena: J of Liq. Chrom., 30, 309, (2007).
- 20. H.H. Williard, L.L. Merit, F.A. Dean and F.A. Settle, Instrumental methods of analysis, 7th edition, C.B.S. Publishers, New Delhi, 2002.
- 21. GN Menon, LB White, Department of Analytical Research, Abbott Laboratories, (pub med-index for MEDLINE).
- 22. Food and Drug Administration (FDA), "Analytical Procedures and Methods Validation: Chemistry, Manufacturing and Controls Documentation;" Federal Register (Notices), Vol: 65 (169), 52776-52777, 2000.
- 23. Vibha G et al., Development and validation of HPLC method - a review. International Research Journal of Pharmaceutical and Applied Sciences. 2012, 2(4), 22-23.
- 24. Bliesner DM. In: Validating Chromatographic Methods. John Wiley & Sons Inc. 2006, 88-92.
- 25. Development and validation of HPLC method A Review, Vibha Gupta et al, International Research Journal of Pharmaceutical and Applied Sciences, 2012; 2(4):17-25.
- 26. A Review: HPLC Method Development and Validation, Santosh Kumar Bhardwaj *et al. International Journal of Analytical and Bioanalytical Chemistry, accepted 20 November 2015.
- 27. Method Development: A Guide to Basics Quantitative Qualitative HPLC, LC, & GC chromacademy.
- 28. Lalit V Sonawane* Bioanalytical Method Validation and Its Pharmaceutical Application- A Review Pharmaceutica Analytical Acta 2014, 5:3Center for Drug Evaluation and Research (CDER) Reviewer Guidance.
- 29. Validation of Analytical Procedures: Methodology. ICH-Guidelines Q2B, Geneva. 1996, 11. (CPMP/ICH/281/95).
- 30. ICH Topic Q 2 (R1) Validation of Analytical Procedures: Text and Methodology.
- 31. Tuba Reçber, Selin Seda Timur, Sevilay Erdoğan Kablan, Fatma Yalçın, Tutku Ceren Karabulut, R. Neslihan Gürsoy, Hakan Eroğlu, Sedef Kır, Emirhan Nemutlu, A stability indicating RP-HPLC method for determination of the COVID-19 drug Molnupiravir applied using Nano formulations in permeability studies, Journal of Pharmaceutical and Biomedical Analysis, Volume 214, 30 May 2022, 114693.
- 32. Jahnavi Bandla* and S. Ganapathy, Development and Validation of a Stability-indicating Method for the Simultaneous Estimation of Sofosbuvir and Ledipasvir by RP-HPLC, Indian J Pharm Sci 2018; 80(6):1170-1176.