



ISOLATION AND GCMS CHARACTERIZATION OF CERTAIN NON-POLAR COMPOUNDS FROM *SPILANTHES CILIATA*

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ABSTRACT

Spilanthes ciliata, commonly known as toothache plant, is a traditional medicinal plant. In the present study an attempt was made to analyze the oil obtained after solvent extraction of flowers of the plant. GCMS of the extract revealed presence of 15 different compounds in the oil. 11-tridecene-1-ol (18.72%), 2R-acetoxymethyl-1,3,3-trimethyl-4T-(3-methyl-2-buten-1-yl)-1T-cyclohexanol (12.66%), Pentadecanoic acid (8.36%), 1-hexacosanol (8.32%), Nonadecane (7.35%) and Hexatriacontane (7.014%) were found to be the major constituents and the remaining were in trace amounts. Most of the compounds identified are found associated with various biological activities. The study presents a large-scale isolation of major compounds for their characterization and medicinal application.

KEY WORDS

GCMS; Fatty acids; sesquiterpenoid; *Spilanthes ciliata*

1. INTRODUCTION

Plants have been used as a reliable source of clinically important compounds that confer health benefits to humans since a long time. The plants continued to serve as cheap and best source of biomedicine, supplements and chemical entities to serve as source of drugs. Their mechanism of action generally imparts synergistic effects on human body making them efficient and safer as compared to the drugs that are synthetic ones. Recent reports from WHO estimate reveals nearly 80% of population rely on herbal medicines as primary healthcare in developing countries [1]. Genus *Spilanthes* is one such plant known since antiquity for several medicinal properties. Members of this genus, commonly known as toothache plants contain various bioactive molecules including alkaloids, sesquiterpenoids, flavonoids, tannins, coumarins, saponins, glycosides, phenolics, lignans, steroids etc. [2,3]. There are 300 species of *Spilanthes*, reported to be distributed widely in tropics and subtropics among which 6 species namely *S. calva*, *S. paniculata*, *S.*

radicans, *S. ciliata*, *S. uliginosa* and *S. oleracea* are found in India [4,5]. Reports on occurrence of *Spilanthes ciliata* (syn: *Acmella ciliata*) indicate their distribution in Chhattisgarh, Kerala and some parts of Tamil Nadu [6]. Often the whole plant is used in the treatment of dysentery and rheumatism, while flowers in particular served as spice in Japanese food habits [7]. Though the phyto-constituents of many of the species of *Spilanthes* is described, the information on *Spilanthes ciliata* has not been completely described so far. Here we present our results on GCMS of the hexane fraction of the flowers extract of this important medicinal plant.

2. MATERIALS AND METHODS

2.1 Plant Material

Fresh flower heads of *Spilanthes ciliata* were collected from the plants that we maintain in the experimental garden of our department. The flower heads harvested were dried at 40°C in hot air oven until consecutive similar dry weight was achieved. The dry flowers were

made into powder in a warring blender to yield *S. ciliata* floral powder.

2.2 Preparation of hexane fraction

200g of floral powder was extracted with methanol (2L, 100%) by incubating for 72h at $28\pm 2^{\circ}\text{C}$ temperature. The methanol solution was then filtered through Whatman No.1 filter paper and the filtrate obtained was rotary evaporated to obtain the crude extract. The crude extract was then subjected to normal phase chromatography using silica (200-400) gel column with hexane as mobile phase. The hexane extract was evaporated, and a yellow oil was obtained. An aliquot of the oil was mixed with fresh hexane and subjected to GCMS.

2.3 GCMS Analysis

The mass analysis of the extracted oil was performed using Perkin Elmer GCMS (Model: Clarus 680). GC used in the analysis employed a fused silica column (30m x 0.25mm, ID: 250 μm) packed with Elite-5MS (5% biphenyl 95% dimethylpolysiloxane) and the components were separated using helium as carrier gas set at a constant flow of 1 ml/min. The injector temperature was set at 260°C during the chromatographic run. 1 μL of the sample was injected into the instrument and oven temperature varied from 60°C (2 min) to 300°C with increment set at $10^{\circ}\text{C}/\text{min}$ and a hold of 6 min at 300°C . The mass detector parameters include transfer line and ion source temperature both set at 240°C , ionization mode electron impact at 70 eV and a scan time 0.2 sec with a scan interval of 0.1 sec. The spectrums of the components were compared with the database of spectrum of known compounds available in the GCMS NIST (2008) library.

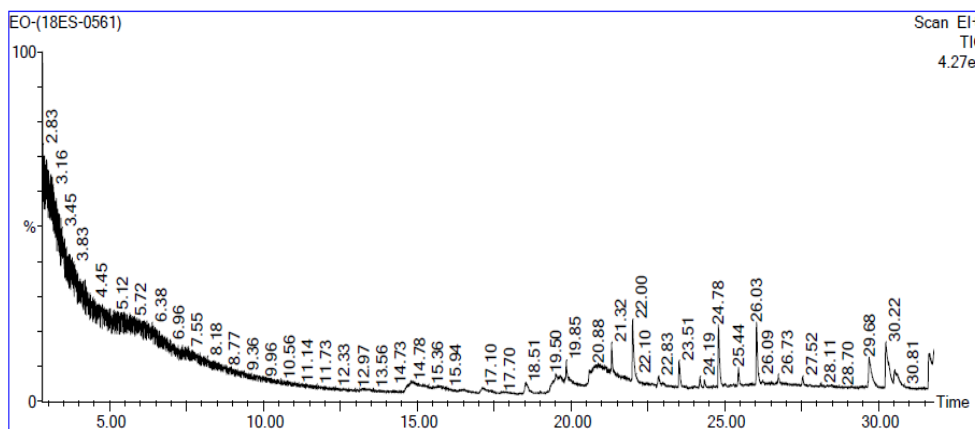
3. RESULTS AND DISCUSSION

A picture of the plant used in the current study is presented (Figure 1). The plant has several features common to its close relatives *Spilanthes paniculata* and *Spilanthes calva* [8]. Several species cultivated under the name of *Spilanthes* were differentially identified. For example based on phenotypic details and description, *S. ciliata* has been used as synonym for *S. acmella*, *S. mauritiana* and *S. calva* [8]. Similarly, *S. oleracea* is used as a synonym for *A. oleracea* and *S. acmella* [9]. We propose a systematic study of all the species under *Spilanthes/Acmella* at molecular and phytochemical level is needed for fine tuning the species confirmation. Nevertheless, the crop of the cultivated *Spilanthes* species is mostly used as toothache plants and the determination of phytoconstituents of the plant and their biological properties is a subject of extensive research in many labs around the globe [10,11].

Figure 1: *Spilanthes ciliata* crop at flowering stage growing in the experimental garden of Department of Biotechnology, Pondicherry University



Figure 2: GC chromatogram showing peaks for different compounds present in hexane fraction of flower extract of *S. ciliata*



A picture of chromatogram obtained after GC analysis is presented (Figure 2). An observation of the chromatogram reveals presence of many distinct peaks indicating presence of several compounds in the hexane fraction of the flower extract. Using GCMS coupled with NIST library search a total of 15 different compounds could be identified. The fragmentation pattern of the individual compounds along with their deduced structure is presented (Figure 3). The compounds identified by GCMS followed by library search with name, molecular formula, molecular weight and their potential biological properties is presented (Table 1). Among the compounds identified there were a number of fatty acids, esters, fatty alcohols, higher alkanes and

terpenoids were present in the extract. Barring eicosanoic acid all other compounds are new report for *S. ciliata*. 11-tridece-1-ol constituted the most abundant (18.72%) component in the extract. This was followed by 2R-acetoxymethyl-1,3,3-trimethyl-4T-(3-methyl-2-buten-1-yl)-1T-cyclohexanol, Pentadecanoic acid, 1-hexacosanol, Nonadecane, Hexatriacontane, 4,4,6a,6b,8a,11,11,14b-Octamethyl-1,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,14,14a,14b-octadecahydro-2H-picen-3-one, Sulfurous acid, 2-propyl tridecyl ester, 3-O-Acetyl-6-methoxycycloartenol and 2,4,4-Trimethyl-3-hydroxymethyl-5A-(3-methyl-but-2-enyl)-cyclohexene while others were in trace amount. This study revealed 14 new compounds

in *S. ciliata* of which 12 were oxygenated. Earlier in *S. acmella*, there were 20 compounds identified in the essential oil with only terpen-4-ol being oxygenated [12]. Later on, in *S. acmella* 44 compounds and in *A. oleracea* 71 compounds were identified [8,9]. In another study the hexane fraction of *A. oleracea* obtained following a similar procedure as ours was found to contain several fatty acids as is reported here [13]. But there were differences in number and type of compounds identified in the present and other studies cited above which might be attributed to the difference in isolation procedures followed. We infer that applying different procedures for isolation and fractionation would facilitate identification of diverse array of compounds with some of them becoming potential lead for developing new drugs. Often hydro-distillation method [7,8, 14-16] or gradient elution [13] have been used. The new compounds identified in our study

appear to be associated with antineoplastic, anti-inflammatory, anti-microbial, anti-cancerous and other biological properties (Table 1). The sesquiterpenoid (2R-acetoxymethyl-1,3,3-trimethyl-4t-(3-methyl-2-buten-1-yl)-1tcyclohexanol) found in the oil is a potential anticancer agent [17]. Another compound identified in the oil sample namely 2,4,4-Trimethyl-3-hydroxymethyl-5A-(3-methyl-but-2-enyl)-cyclohexene satisfies the rule of five and indicates favorable bioavailability as determined by molinspiration calculation and is predicted to be a substrate for Cytochrome P₄₅₀ 2J (CYP2J) enzyme which are expressed in heart tissues [18,19]. Moreover, similar esters have also been also reported in members of Malvaceae [20]. Thus, our study paves the way for large scale isolation of these compounds and subjecting them to validation for clinical application.

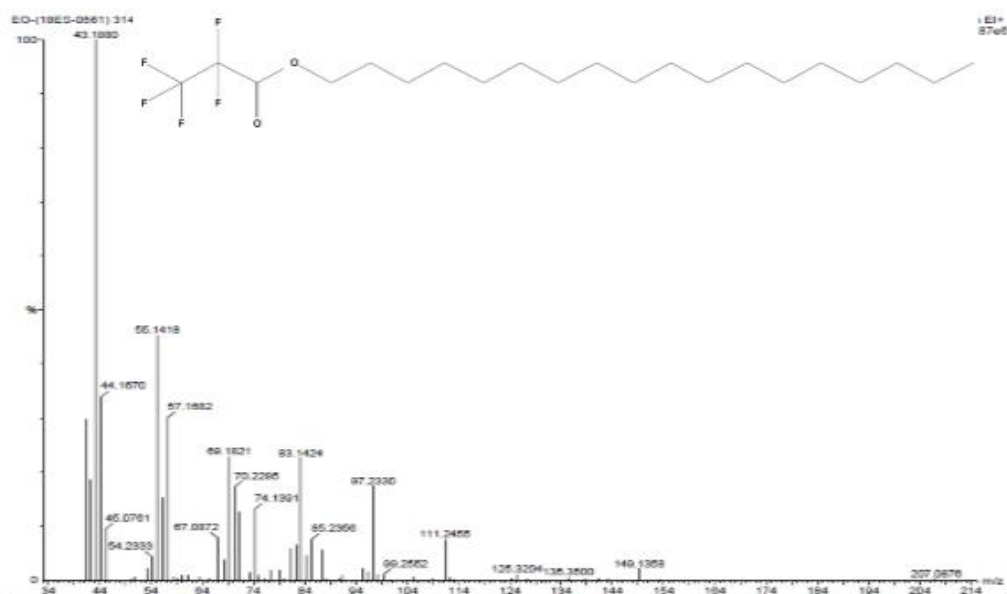
Table 1: List of compounds identified from flower extract of *S. ciliata* and their associated biological properties

S.No	RT	Area (%)	Compound Name	Chemical Nature	Molecular Formula	Molecular Weight	Bioactivity Properties	Reference
1	18.53	2.720	Pentafluoropropionic acid, octadecyl ester	Ester	C ₂₁ H ₃₇ O ₂ F ₅	416	NR	NA
2	19.4	1.850	Eicosanoic acid	Fatty acid	C ₂₀ H ₄₀ O ₂	312	Peroxisome proliferator activated receptor delta, alpha and gamma (PPAR-d/a/g) agonist	21
3	19.5	8.368	Pentadecanoic acid	Fatty acid	C ₁₅ H ₃₀ O ₂	242	Peroxisome proliferator activated receptor delta (PPARd) agonist	22
4	19.84	8.320	1-hexacosanol	Fatty alcohol	C ₂₆ H ₅₄ O	382	Neuromuscular regeneration	23
5	19.94	2.446	Lauroyl peroxide	Peroxide	C ₂₄ H ₄₆ O ₄	398	Bleaching agent, Polymerization catalyst, Plasticizer	24
6	20.58	2.849	Tetratetracontane	Higher alkane	C ₄₄ H ₉₀	618	Anti-microbial	25

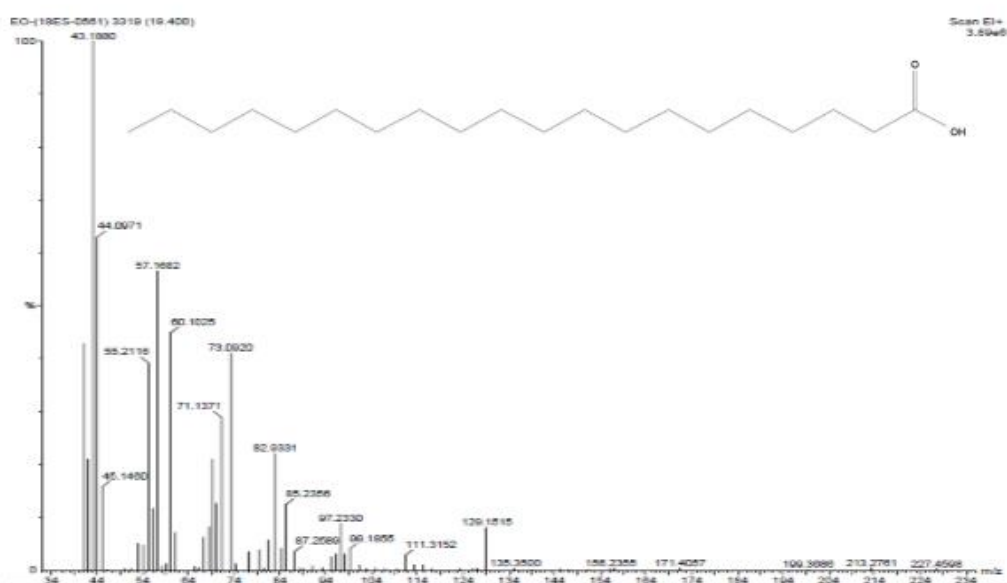
S.No	RT	Area (%)	Compound Name	Chemical Nature	Molecular Formula	Molecular Weight	Bioactivity Properties	Reference
7	20.88	18.724	11-tridecen-1-ol	Fatty alcohol	C ₁₃ H ₂₆ O	198	Detergents, Cosmetics, Leather processing	26
8	21.99	7.358	Nonadecane	Alkane	C ₁₉ H ₄₀	268	Source of n-parafilms	27
9	23.50	2.853	Sulfurous acid, 2-propyl tetradecyl ester	Ester	C ₁₇ H ₃₆ O ₃ S	320	NR	NA
10	24.78	7.014	Hexatriacontane	Higher alkane	C ₃₆ H ₇₄	506	Anti-microbial	28
11	26.02	6.608	Sulfurous acid, 2-propyl tridecyl ester	Ester	C ₁₆ H ₃₄ O ₃ S	306	NR	NA
12	29.68	6.850	4,4,6a,6b,8a,11,11,14b-Octamethyl-1,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,14,14a,14b-octadecahydro-2H-picen-3-one	Triterpenoid	C ₃₀ H ₄₈ O	424	Anti-neoplastic	29
13	30.22	12.662	2R-acetoxymethyl-1,3,3-trimethyl-4T-(3-methyl-2-buten-1-yl)-1T-cyclohexanol	Sesquiterpenoid	C ₁₇ H ₃₀ O ₃	282	Anti-cancerous	30
14	30.52	6.327	3-O-Acetyl-6-methoxy-cycloartenol	Triterpenoid	C ₃₃ H ₅₄ O ₃	498	Anti-inflammatory, anti-convulant	31
15	31.62	5.051	2,4,4-Trimethyl-3-hydroxymethyl-5A-(3-methyl-but-2-enyl)-cyclohexene	Organic Compound	C ₁₅ H ₂₆ O	222	Antibacteria	32

• NR – Not Reported, NA – Not Available

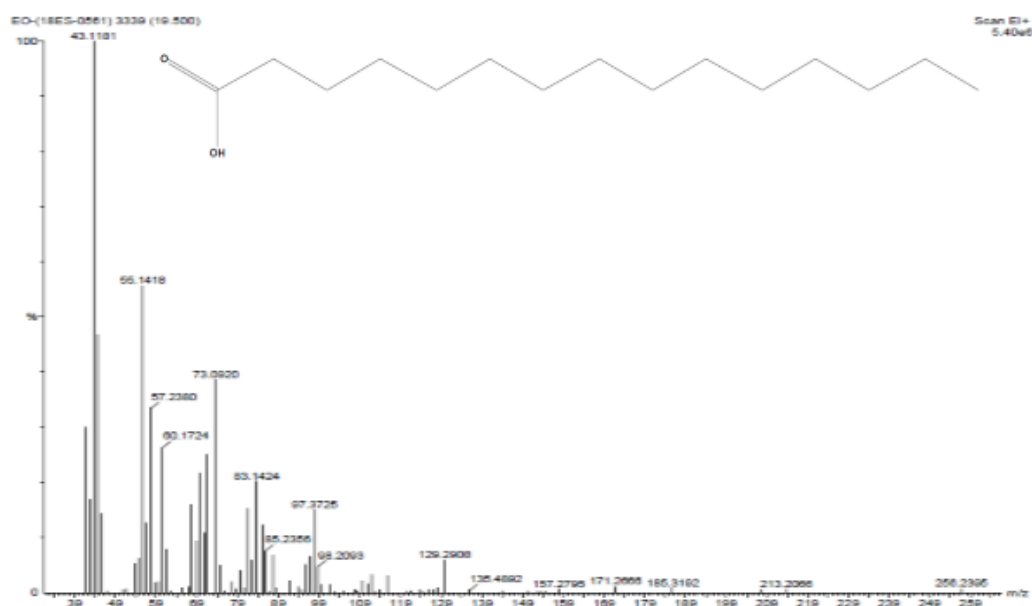
Figure 3: GCMS fragmentation pattern and deduced chemical structure of the identified compounds



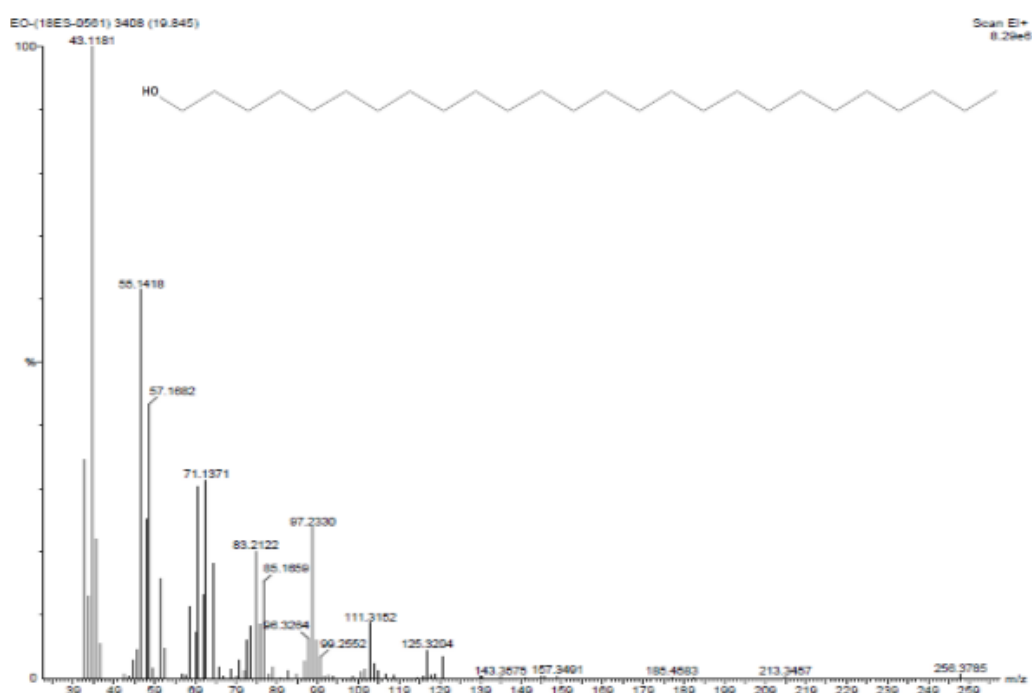
1. Pentafluoropropionic acid, octadecyl ester



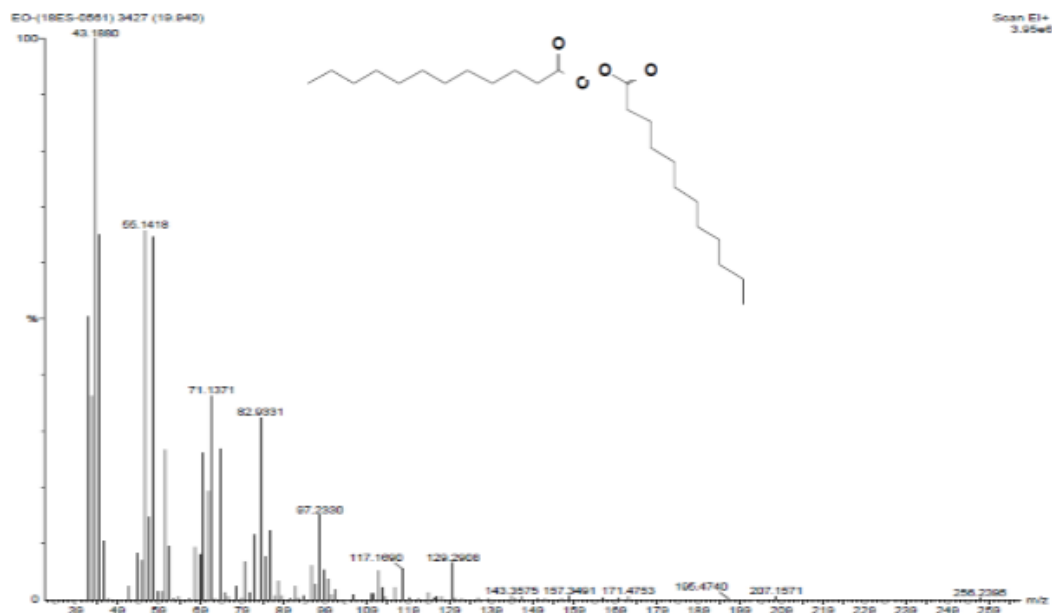
2. Eicosanoic acid



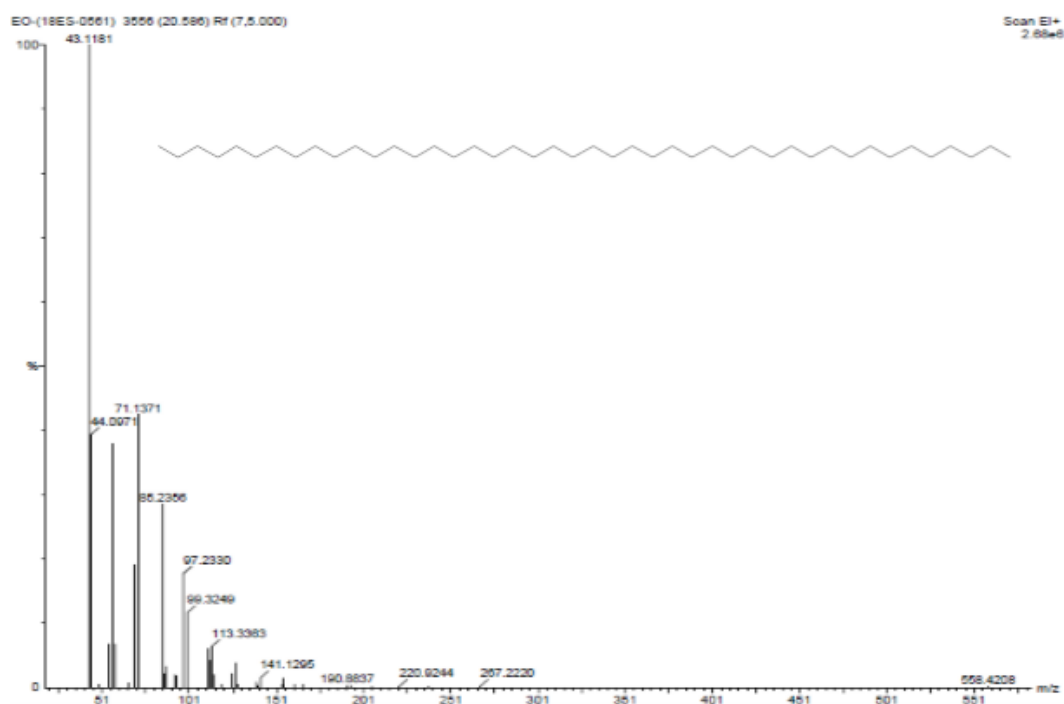
3. Pentadecanoic acid



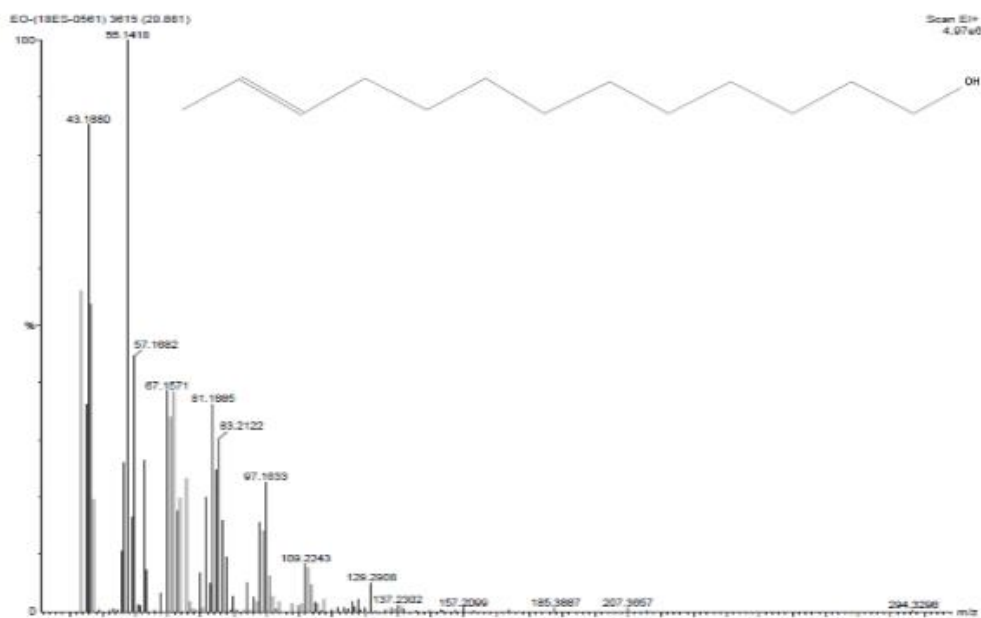
4. 1-hexacosanol



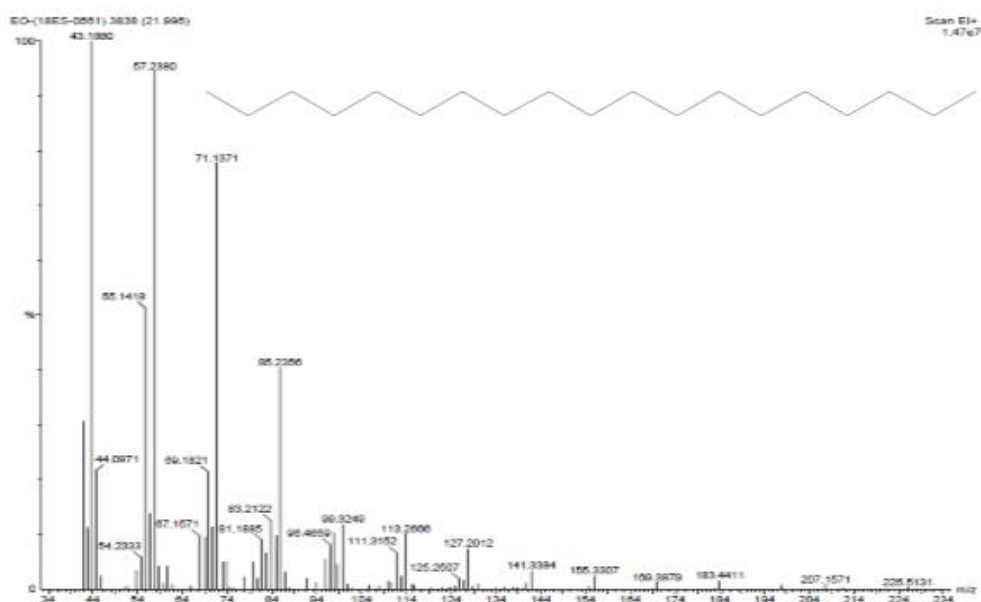
5. Lauroyl peroxide



6. Tetratetracontane



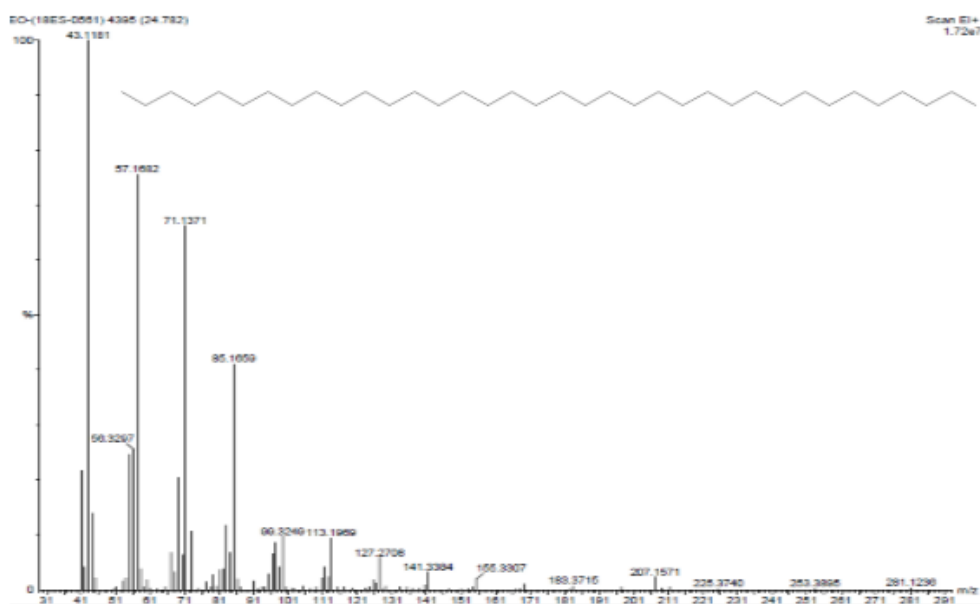
7. 11-tridecen-1-ol



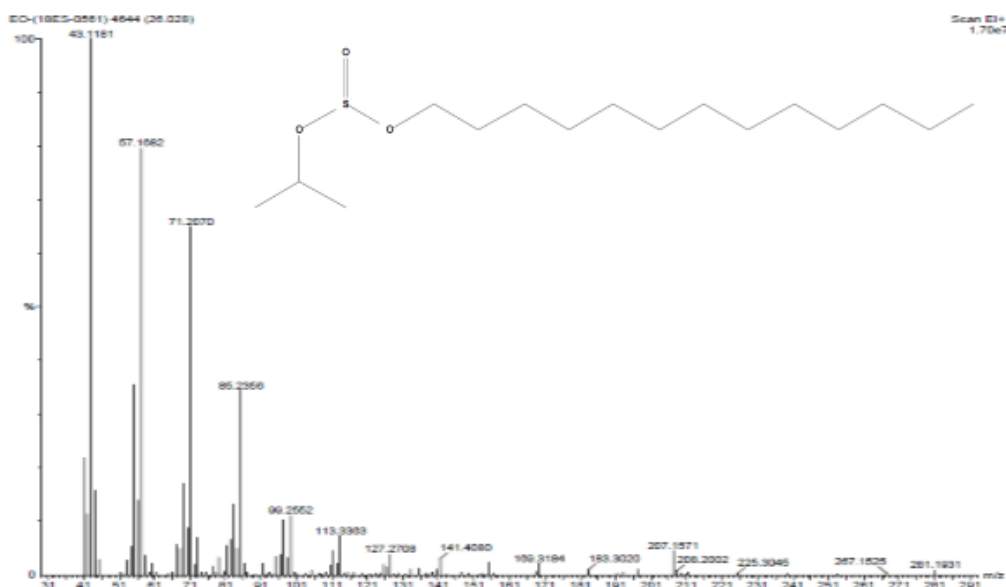
8. Nonadecane



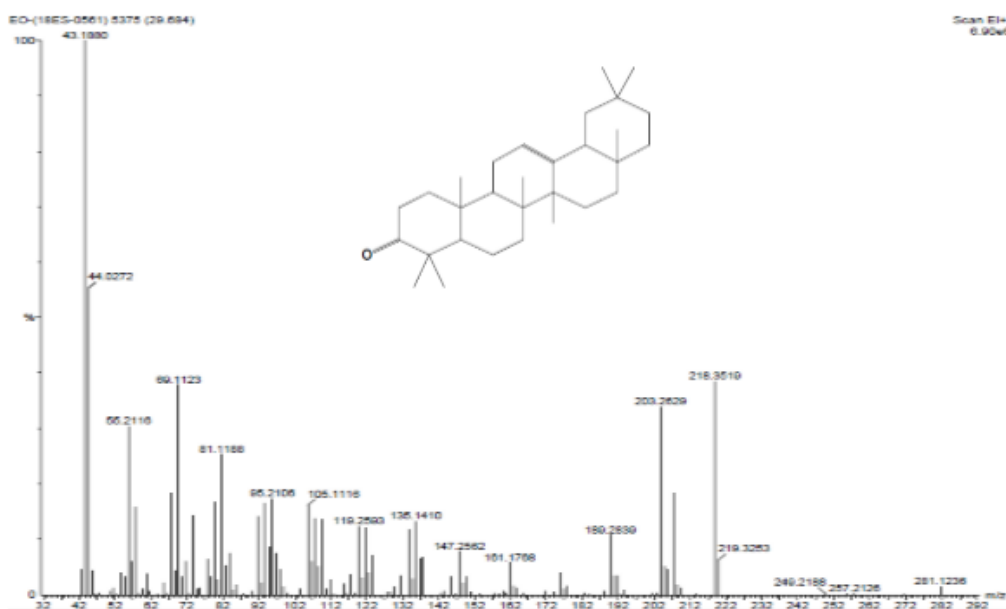
9. Sulfurous acid, 2-propyl tetradecyl ester



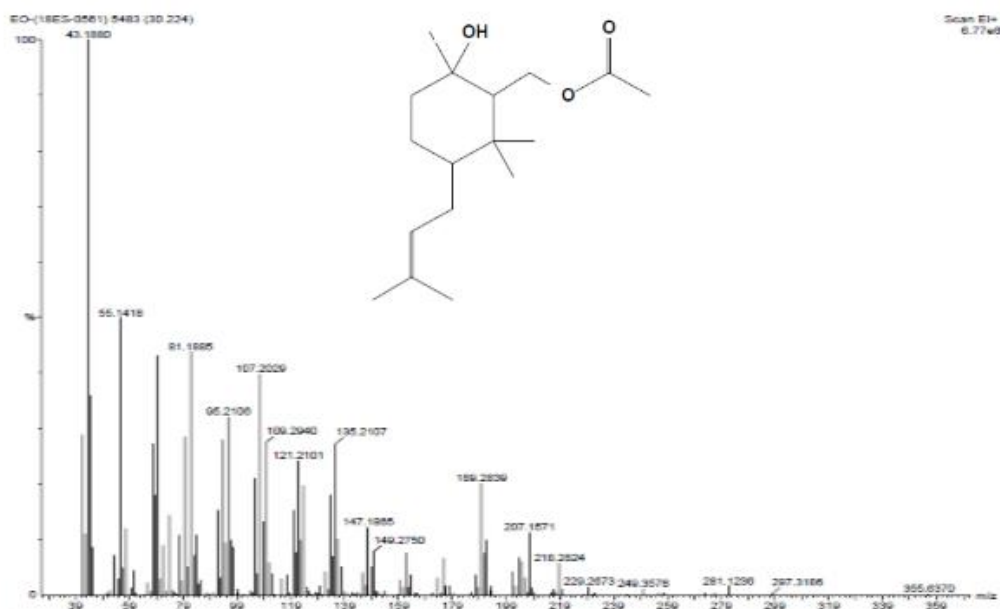
10. Hexatriacontane



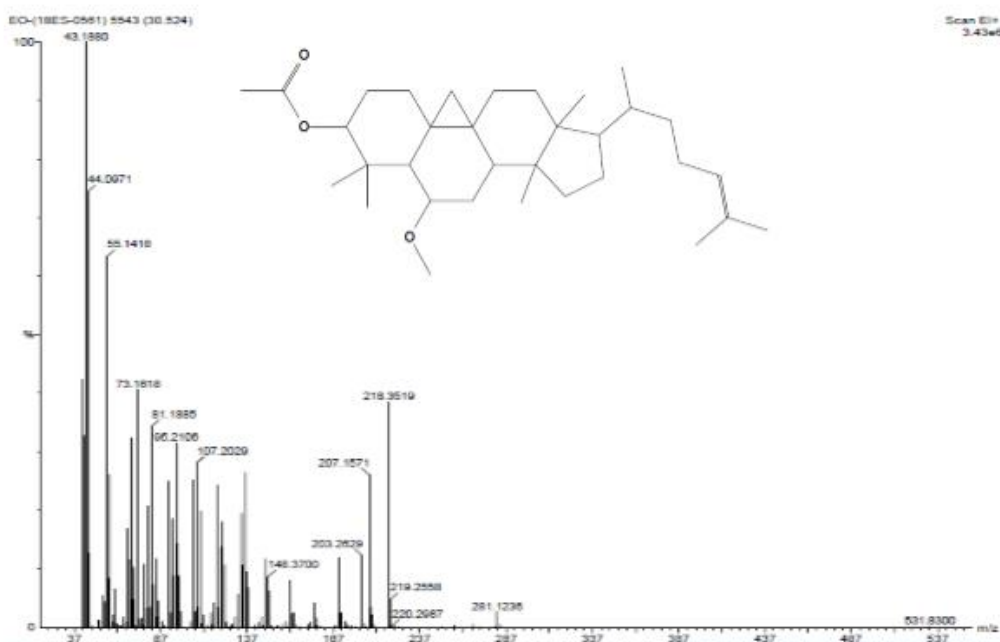
11. Sulfurous acid, 2-propyl tridecyl ester



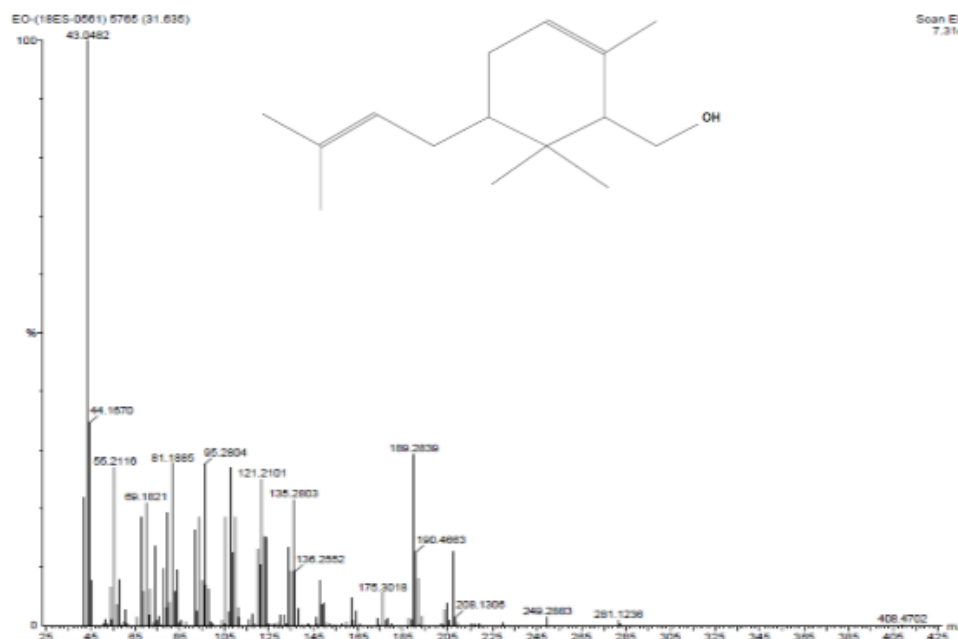
12. 4,4,6a,6b,8a,11,11,14b-Octamethyl-1,4,4a,5,6,6a,6b,7,8,8a,9,10,11,12,12a,14,14a,14b-octadecahydro-2H-picen-3-one



13. 2R-acetoxymethyl-1,3,3-trimethyl-4T-(3-methyl-2-buten-1-yl)-1T-cyclohexanol



14. 3-O-Acetyl-6-methoxy-cycloartenol



15. 2,4,4-Trimethyl-3-hydroxymethyl-5A-(3-methyl-but-2-enyl)-cyclohexene

CONCLUSION

Traditional medicines have played vital role and remain as high value drugs even in the era of modern medicine. Phyto-constituents are responsible for medicinal value of any plant hence, in this regard a study on phyto-components in hexane fraction (oil) of *S. ciliata* flower extract was performed using GCMS technique. The hexane extract yielded a yellow oil which was found to be rich in fatty alcohols, sesquiterpenoid, esters, fatty acids and higher alkanes. Most of the compounds identified in the oil associated with anti-cancerous, anti-microbial, anti-inflammatory and other biological properties. This study thus facilitates large scale isolation of new compounds that may serve as lead molecules for discovery of new drugs and other products.

CONFLICTS OF INTEREST

The authors declare no conflict of interest in the publication of this manuscript.

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REFERENCES

1. Tiwari, K.L., Jadhav, S.K., Joshi, V., An updated review on medicinal herb Genus *Spilanthes*, J. Chin. Integr. Med. 9, 1170–1178, (2011).
2. Preetha T.S, Mohan S.N, Najeena P.M, Deepthi S.R., Chemical Fingerprinting of *Spilanthes acmella* L. (Murr.) -An acutely threatened medicinal plant of pharmaceutical importance by HPTLC, FTIR and UV-Vis spectroscopic tools, World J. Pharm. Pharm. Sci., 3, 1275–1287, (2014).
3. Lagnika L, Amoussa A.M.O, Adjileye R.A.A, Laleye A, Sanni A., Antimicrobial, antioxidant, toxicity and phytochemical assessment of extracts from *Acmella uliginosa*, a leafy-vegetable consumed in Bénin, West Africa, BMC Complement Altern Med., 16(34), (2016).
4. Paulraj J, Govindarajan R, Palpu P., The genus *Spilanthes* ethnopharmacology, phytochemistry, and pharmacological properties: a review. Adv. Pharmacol. Sci., <http://dx.doi.org/10.1155/2013/510298>, (2013).
5. Singh, M, Chaturvedi, R., Screening and quantification of an antiseptic alkylamide, spilanthol from in vitro cell and tissue cultures of *Spilanthes acmella* Murr. Ind. Crops Prod., 36, 321–328, (2012).

6. Bhat Z.S, Jaladi N, Khajuria R.K, Shah Z.H, Arumugam N, Comparative analysis of bioactive N-alkylamides produced by tissue culture raised versus field plantlets of *Spilanthes ciliata* using LC-Q-TOF (HRMS), J. Chromatogr. B., 195-203, (2016).
7. Leng T.C, Ping N. S, Lim B. P, Keng C.L., Detection of bioactive compounds from *Spilanthes acmella* (L.) plants and its various in vitro culture products, J.Med.Aromat.Plants Sci., 5: 371–378, (2011).
8. Jirovetz L, Buchbauer G, Wobus A, Shafi M.P, Abraham G.T., Essential Oil Analysis of *Spilanthes acmella* Murr. Fresh Plants from Southern India, J. Essent. Oil Res., 17, 429-431 (2005).
9. Benelli G, Pavela R, Drenaggi E, Maggi F., Insecticidal efficacy of the essential oil of jambú (*Acmella oleracea* (L.) R.K. Jansen) cultivated in central Italy against filariasis mosquito vectors, houseflies and moth pests, Journal of Ethnopharmacology, <https://doi.org/10.1016/j.jep.2018.08.030>, (2018).
10. Dubey S, Maity S, Singh M, Saraf S.A, Saha S., Phytochemistry, pharmacology and toxicology of *Spilanthes acmella*: a review. Adv. Pharmacol. Sci., <http://dx.doi.org/10.1155/2013/423750>, (2013).
11. Prachayasittikul S, Suphapong S, Worachartcheewan A, Lawung R, Ruchirawat S, Prachayasittikul V., Bioactive Metabolites from *Spilanthes acmella* Murr., Molecules, 14, 850-867, (2009).
12. Baruah R.N, Leclercq P.A., Characterization of the Essential Oil from Flower Heads of *Spilanthes acmella*, J Essent Oil Res, 5:6, 693-695 (1993)
13. Phrutivorapongkul A, Chaiwon A, Vejabhikul S, Netisingha W, Chansakaow S., AN ANESTHETIC ALKAMIDE AND FIXED OIL FROM ACMELEA OLERACEA, J Health Res., 22(2): 97-99, (2008).
14. Maimulyanti A, Prihadi A.R., Chemical composition of essential oil and hexane extract and antioxidant activity of various extracts of *Acmella uliginosa* (Sw.) Cass flowers from Indonesia, Agriculture and Natural Resources, 50, (2016)
15. Silveira N, Saar J, Santos A.D.C, Barison A, Sandjo L.P, Kaiser M, Schmidt T.J, Biavatti M.W., A New Alkamide with an Endoperoxide Structure from *Acmella ciliata* (Asteraceae) and Its in Vitro Antiplasmodial Activity, Molecules, 21, 765; doi:10.3390/molecules21060765, (2016).
16. Begum J, Bhuiyan Md.N.I, Chowdhury J.U., ESSENTIAL OIL FROM INFLORESCENCE OF SPILANTHES CALVA D.C., Bangladesh J. Bot. 37(2), 217-218, (2008).
17. Naine S.J, Devi C.S, Mohanasrinivasan V, Doss C.G.P, Kumar D.T., Binding and molecular dynamic studies of sesquiterpenes (2R-acetoxymethyl-1,3,3-trimethyl-4t-(3-methyl-2-buten-1-yl)-1tcyclohexanol) derived from marine Streptomyces sp. VITJS8 as potential anticancer agent, Appl Microbiol Biotechnol., 100(6), 2869-2882, (2015).
18. Kang W, Elitzer S, Noh K, Bednarek T and Weiss M., Myocardial pharmacokinetics of ebastine, a substrate for cytochrome P450 2J, in rat isolated heart, Br J Pharmacol., 163, 1733–1739, (2011).
19. Balasundaram A, Ragupathy R, Sankar S, Thiyagarajan M, Ravi L, Karuppasamy R, Veerappapillai S., Investigation of Phytochemicals and Computational Approach for the Evaluation of Therapeutic Properties of Ethanolic Leaf Extract of *Callistemon citrinus*, Int. J. Pharm. Sci. Rev. Res., 20, 110-116, (2016).
20. Vivekraj P, Vijayan, A, Anandgideon V., Analysis of Phytochemical constituents of the chloroform extracts of *Abutilon hirtum* (Lam.) Sweet using GC-MS Method, Int. J. Pharm Res., 5(8), 167, (2015).
21. Pubchem.ncbi.nlm.nih.gov/compound/Arachidic_acid
22. Pubchem.ncbi.nlm.nih.gov/compound/pentadecanoic_acid
23. Azzouz M, Kennel P.F, Warter J-M, Poindron P, Borg J., Enhancement of Mouse Sciatic Nerve Regeneration by the Long Chain Fatty Alcohol, N-Hexacosanol., EXPERIMENTAL NEUROLOGY 138, 189–197 (1996)
24. Pubchem.ncbi.nlm.nih.gov/compound/Lauroyl_peroxide
25. Barretto D.A, Vootla S.K., GC-MS ANALYSIS OF BIOACTIVE COMPOUNDS AND ANTIMICROBIAL ACTIVITY OF CRYPTOCOCCUS RAJASTHANENSIS KY627764 ISOLATED FROM BOMBAY MORI GUT MICROFLORA, Int. J. Adv. Res. 6(3), 525-538 (2018)
26. Noweck K, Grafahrend W., Ullmann's Encyclopedia of Industrial Chemistry, Vol 14, Weinheim: Wiley-VCH, 117-141, (2012)
27. Pubchem.ncbi.nlm.nih.gov/compound/nonadecane
28. Xuanji X, Zengjun G, Hui Z, Xia L, Jun L, Dandan Li, Jun Li., Chemical composition, in vitro antioxidant activity and α -glucosidase inhibitory effects of the essential oil and methanolic extract of *Elsholtzia densa* Benth. Natural Product Research, 30(23), 2707–2711, (2016)
29. Setzer W.N, Setzer M.C., Plant-Derived Triterpenoids as Potential Antineoplastic Agents, Mini Reviews in Medicinal Chemistry, 3, 540-556, (2003).
30. Naine S.J, Devi C.S, Mohanasrinivasan V, Doss C.G.P, Kumar D.T., Binding and molecular dynamic studies of sesquiterpenes (2R-acetoxymethyl-1,3,3-trimethyl-4t-(3-methyl-2-buten-1-yl)-1tcyclohexanol) derived from marine Streptomyces sp. VITJS8 as potential anticancer agent, Appl Microbiol Biotechnol, DOI 10.1007/s00253-015-7156-2 (2015)
31. Sugumar N, Karthikeyan S., Preliminary Phytochemical Screening GC-MS and FTIR Profiling of Methanolic Extract of Leaves of *Eupatorium triplinerve* Vahl, Int. J. Multidiscip. Res. Dev., 2(8), 335-340, (2015).



32. Karthik R, Saravanan R, Ebenezer R.K, Sivamalai T., Isolation, Purification, and Characterization of Avian Antimicrobial Glycopeptide from the Posterior Salivary

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Gland of *Sepia pharaonis*, Appl Biochem Biotechnol., 175:1507–1518, (2015).

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